INTERNATIONAL RESEARCH JOURNAL OF PHARMACY



www.irjponline.com

ISSN 2230 - 8407



Research Article

IN VITRO ANTI-ARTHRITIC ACTIVITY OF ETHANOLIC EXTRACT OF *CALLICARPA MACROPHYLLA* FLOWER

Santosh Kumar Gupta^{*1}, Amit Gupta², A.K. Gupta¹, Dhirendra Pakash¹, Vedpal¹ ¹Agra Public Pharmacy College, Agra, U.P., India ²Jaipur College of Pharmacy, Jaipur, Rajasthan, India E-mail:santosh1015@gmail.com

Article Received on: 11/01/13 Revised on: 02/02/13 Approved for publication: 10/03/13

DOI: 10.7897/2230-8407.04332

IRJP is an official publication of Moksha Publishing House. Website: www.mokshaph.com © All rights reserved.

ABSTRACT

The present study is aimed to evaluate the *in-vitro* anti-arthritic activity of ethanolic extract of *Callicarpa macrophylla* flower using inhibition of protein denaturation model and human red blood cell Membrane stabilization model. Diclofenac sodium was used as a standard drug. Results revealed that the ethanolic extract of *Callicarpa macrophylla* at different concentrations possessed significant anti-arthritic activity as compared to standard drug used as Diclofenac sodium. The results obtained in the present investigation Indicate that ethanolic extract of *Callicarpa macrophylla* flower showed anti-arthritic activity.

Key words: Callicarpa macrophylla, anti-inflammatory, Anti-arthritic, protein denaturation, Membrane stabilization.

INTRODUCTION

Rheumatoid arthritis is an autoimmune disease in which there is joint inflammation, synovial proliferation and destruction of articular cartilage¹ Inflammation is a bodily response to injury, infection or destruction characterized by heat, redness, pain, swelling and disturbed physiological functions. Inflammation is a normal protective response to tissue injury caused by physical trauma, noxious chemical or microbial agents. It is the body response to inactivate or destroy the invading organisms, to remove the irritants and set the stage for tissue repair. It is triggered by the release of chemical mediators from injured tissue and migrating cells² It is a common disease having peak incidence in 3rd to 4th decades of life with 3-5 times higher preponderance in female.³ Its prevalence depends upon age.⁴ The commonly used drug for management of inflammatory conditions are non-steroidal anti-inflammatory drugs (NSAIDs), which have several adverse effects especially gastric irritation leading to formation of gastric ulcers.^{5,6} Herbal drugs constitute a major part in all the traditional system of medicine. Herbal medicine is a triumph of popular therapeutic diversity.⁷ The factors responsible for the continued and extensive use of herbal remedies in India are their effectiveness, easy availability, low cost, comparatively less toxic effects and shortage of practitioners of modern medicine in rural areas.⁸ Number of synthetic medicines has been derived from medicinal herbs.9 The major merits of herbal medicine seem to be their perceived efficacy, low incidence of serious adverse effects and low cost.

Callicarpa macrophylla of family Verbenaceae, is an indigenous plant of India, have with a wide spectrum of therapeutic properties. Its leaves are reported to have anti-inflammatory, analgesic and antipyretic effects.^{10,11} while roots have are anti-inflammatory and analgesic effects.¹² Its stems of C. macrophylla has been evaluated for its anti-fungal activity and results are very significant.¹³

Previously we had reported the presence of glycosides, saponins, flavanoids, tannins and carbohydrates in the aqueous extract of stems of C. macrophylla Vahl. While their alcoholic extracts have significant glycoside, flavanoid, tannins, carbohydrates and steroid content.¹⁴

MATERIALS AND METHODS

Plant material

The flower of plant *Callicarpa macrophylla* were collected from local area of agra, U.P., India, and authenticated by Dr P N Sharma, department of Botany, Dr. B R Ambedkar university, Agra, U.P. where a voucher specimen No. has been submitted. (Voucher specimen No. BRAU3171)

Preparation of plant extract

Collected flower of *Callicarpa macrophylla* were converted into moderately coarse powder and extracted with solvent ethyl alcohol for 27 hours by soxhlet. The solvent was removed under reduced pressure.

Drugs and chemicals

Diclofenac sodium was obtained from Medley Pharmaceutical Pvt. Ltd., Jammu, J&K, India. Double distilled water from all-glass still was used throughout the study.

Assessment of in-vitro Anti-arthritic activity

Inhibition of albumin denaturation ^{15,16}

The reaction mixture (5 ml) consisted of 0.2 ml of egg albumin (from fresh hen's egg), 2.8 ml of phosphate buffered saline (PBS, pH 6.4) and 2 ml of varying concentrations of extract so that final concentrations become 50, 100, 200, 400, 800 µg/ml. Similar volume of double distilled water served as control. Then the mixtures were incubated at $37 \pm 2^{\circ}$ C in a BOD incubator for 15 mins and then heated at 70°C for 5 mins. After cooling, their absorbance was measured at 660 nm (SHIMADZU, UV 1800) by using vehicle as blank. Diclofenac sodium at the final concentration of (50, 100, 200, 400, 800µg/ml) was used as reference drug and treated similarly for determination of absorbance. The percentage inhibition of protein denaturation was calculated by using the following formula:

% of Inhibition = $100 \times [Vt / Vc - 1]$ Where, Vt = absorbance of test sample, Vc = absorbance of control.

Membrane stabilization test

Preparation of Red Blood cells (RBCs) suspension

Fresh whole human blood (10 ml) was collected and transferred to the heparin zed centrifuged tubes. The tubes were centrifuged at 3000 rpm for 10 min and were washed three times with equal volume of normal saline. The volume of the blood was measured and reconstituted as 10% v/v suspension with normal saline.^{17,18}

Heat induced hemolysis

The reaction mixture (2 ml) consisted of 1 ml of test drug solution and 1 ml of 10% RBCs suspension, instead of drug only saline was added to the control test tube. Aspirin was taken as a standard drug. All the centrifuge tubes containing reaction mixture were incubated in a water bath at 56°C for 30 min. At the end of the incubation, the tubes were cooled under running tap water. The reaction mixture was centrifuged at 2500 rpm for 5 min and the absorbance of the supernatants was taken at 560 nm. The experiment was performed in triplicates. Percent membrane stabilization activity was calculated by the formula mentioned above.^{18,19}

RESULT

Anti-arthritic effect of *Callicarpa macrophylla* was studied significantly by testing various in-vitro parameters. The effect of *Callicarpa macrophylla* on inhibition of protein denaturation and membrane stabilization is shown in table. *Callicarpa macrophylla* at different dose levels (50, 100, 200, 400 and 800µg/ml) provided significant protection against denaturation of proteins and hypotonic saline induced RBC membrane damage.

Test Sample	Conc. ((µg/ml)	% Protection
Ethanolic extract of	50	59.47
flower of Callicarpa	100	64.10
macrophylla	200	88.64
	400	98.21
	800	130.52
Effect of Diclofenac	50	107.24
Sodium	100	112.68
(Std. drugs)	200	147.46
	400	202.14
	800	238.96

 Table 1: In vitro Anti-arthritic activity by inhibition of Protein denaturation method

 Table 2: In vitro Anti-arthritic activity by Membrane stabilization method

Test Sample	Conc. ((µg/ml)	% Protection
Ethanolic extract of	50	12.8
flower of Callicarpa	100	19.82
macrophylla	200	30.72
	400	42.81
	800	54.84
Effect of Diclofenac	50	73.86
Sodium	100	84.06
(Std. drugs)	200	87.72
	400	90.03
	800	96.94

DISCSSUSION

There are certain problems associated with animal use in experimental pharmacological research such as ethical issues and the lack of rationale for their use when other suitable methods are available or could be investigated. Hence, in the present study the protein denaturation bioassay was selected for in vitro assessment of anti-arthritic property of ethanolic extract of *Callicarpa macrophylla*.

Denaturation of tissue proteins is one of the well documented causes of inflammatory and arthritic diseases. Production of auto-antigens in certain arthritic diseases may be due to denaturation of proteins in vivo.^{20,21} Agents that can prevent protein denaturation therefore, would be worthwhile for antiinflammatory drug development. The mechanism of denaturation probably involves alteration in electrostatic, hydrogen, hydrophobic and disulphide bonding.²² From the results of the present study it can be stated that *Callicarpa macrophylla* is capable of controlling the production of auto-antigens due to in vivo denaturation of proteins in rheumatic diseases.

Protective effect on heat and hypotonic saline-induced erythrocyte lysis is known to be a very good index of antiarthritic activity of any agent. ²³ Since the membrane of RBC is structurally similar to the lysosomal membrane, the effect of any substance on stabilization of RBC membrane may be extrapolated to the stabilization of lysosomal membrane.²³

Further studies are needed to elucidate other mechanisms of the in-vitro Anti- arthritic activity of the *Callicarpa macrophylla* extract and to identify the active constituents responsible for the anti-arthritic effect.

REFERENCES

- Singh M, Soni P, Upmanyu N, Shivhare Y. *In-vitro* Anti-arthritic Activity of *Manilkara zapota* Linn. Asian J Pharm Tech 2011:1(4);123-24.
- Chandra S, Chatterjee P, Dey P, Bhattacharya S. Evaluation of Anti-Inflammatory Effect of Ashwagandha: A Preliminary Study in Vitro. Pharmacog J 2012;4(29):47-9. <u>http://dx.doi.org/10.5530/pj.2012.29.7</u>
- 3. Pandey S. Various techniques for the evaluation of anti-arthritic activity in animal models. 2010. J. Adv. Pharm. Tech. Res. 1(2):164-170.
- Mukherjee PK. Quality control of herbal drugs, Syndicate binders, New Delhi, 2002, pp. 13.
- Tripathi KD. Essentials of medical pharmacology. 6th ed. New Delhi:Jaypee Brother's Medical Publishers (P) Ltd.; 2008.
- Bennett PN, Brown MJ. Clinical pharmacology. New Delhi: Churchill Livingstone; 2005.
- Mukherjee PK. Quality control of herbal drugs, Syndicate binders, New Delhi, 2002, pp. 13.
- Agrawal SS, and Paridhavi M. Herbal drug technology. University press Pvt. Ltd., Hyderabad, 2007, pp.2.
- Singh AP. 2006. Distribution of steroid like compound in plant flora, Phcog. Mag. 2(6): 87-89.
- Yadav V, Jayalakshmi S, Patra A, Singla RK. Investigation of analgesic & anti-pyretic potentials of Callicarpa macrophylla Vahl. leaves extracts. Webmedcentral: International Journal of Medicine and Molecular Medicine. 2012; 3(6):WMC003447.
- Yadav V, Jayalakshmi S, Patra A, Singla RK, Patra A. Preliminary assessment of anti-inflammatory activity of Callicarpa macrophylla Vahl. leaves extracts. Indo Global Journal of Pharmaceutical Sciences. 2011; 1(3): 219-222.
- Yadav V, Jayalakshmi S, Patra A, Singla RK, Patra A, Khan S. Assessment of anti-inflammatory and analgesic activities of Callicarpa macrophylla Vahl. roots extracts. Webmedcentral Pharmacology. 2012; 3(5): WMC003366.
- Yadav V, Jayalakshmi S, Patra A, Singla RK, Patra A. Ex vivo screening of stem extracts of Callicarpa macrophylla Vahl. for antifungal activity. Indo Global Journal of Pharmaceutical Sciences. 2012; 2(2): 103-107.
- Yadav VK, Satpathy S, Patra A. Pharmacognostical studies of Callicarpa macrophylla Vahl. stem. International Journal of Phytotherapy Research. 2012; 2(1): 35-41.
- Govindappa M, Naga Sravya S, Poojashri M N, Sadananda TS, Chandrappa C P. Antimicrobial, antioxidant and *in vitro* antiinflammatory activity of ethanol extract and active phytochemical screening of *Wedelia trilobata* (L.) Hitchc. J Pharmacognosy Phytother 2011;3(3):43-51.
- Chandra S, Dey P, Bhattacharya S. Preliminary in vitro assessment of anti-inflammatory property of Mikania scandens flower extract. J Adv Pharm Edu Res 2012;2(1):25-31.

- Sadique J, Rqobahs WA, Bughaith and ElGindi AR. The bioactivity of certain medicinal plants on the stabilization of RBC membrane system. Fitoterapia 1989; 60:525-32.)
- Sakat SS, Juvekar AR, Gambhire MN. *In-vitro* Antioxidant And Antiinflammatory Activity of Methanol Extract of *Oxalis Corniculata* Linn. IJPPS 2010;2(1):146-55
- Shinde UA, Phadke AS, Nari AM, Mungantiwar AA, Dikshit VJ and Saraf MN. Membrane stabilization activity a possible mechanism of action for the anti-inflammatory activity of Cedrus deodara wood oil. Fitoterapia 1999; 70:251-57. <u>http://dx.doi.org/10.1016/S0367-326X(99)00030-1</u>
- Opie EL. On the relation of necrosis and inflammation to denaturation of proteins. J Exp Med. 1962; 115: 597-608. <u>http://dx.doi.org/10.1084</u> /jem.115.3.597
- Umapathy E, Ndebia EJ, Meeme A, Adam B, Menziwa P, Nkeh-Chungag BN, Iputo JE. An experimental evaluation of Albuca setosa aqueous extract on membrane stabilization, protein denaturation and white blood cell migration during acute inflammation. J Med Plants Res. 2010; 4: 789-95.
- Deshpande V, Jadhav VM, Kadam VJ. In-vitro anti-arthritic activity of Abutilon indicum (Linn.) Sweet. IJPP 2009;2(4):644-45
- 23. Brown JH, Mackey HK. Inhibition of heat-induced denaturation of serum proteins by mixtures of non-steroidal anti-inflammatory agents and amino acids, Proc Soc Exp Biol Med 1968;128:225-28.

Cite this article as:

Santosh Kumar Gupta, Amit Gupta, A.K. Gupta, Dhirendra Pakash, Vedpal. In vitro Anti-arthritic activity of ethanolic extract of Callicarpa macrophylla flower. Int. Res. J. Pharm. 2013; 4(3):160-162

Source of support: Nil, Conflict of interest: None Declared