

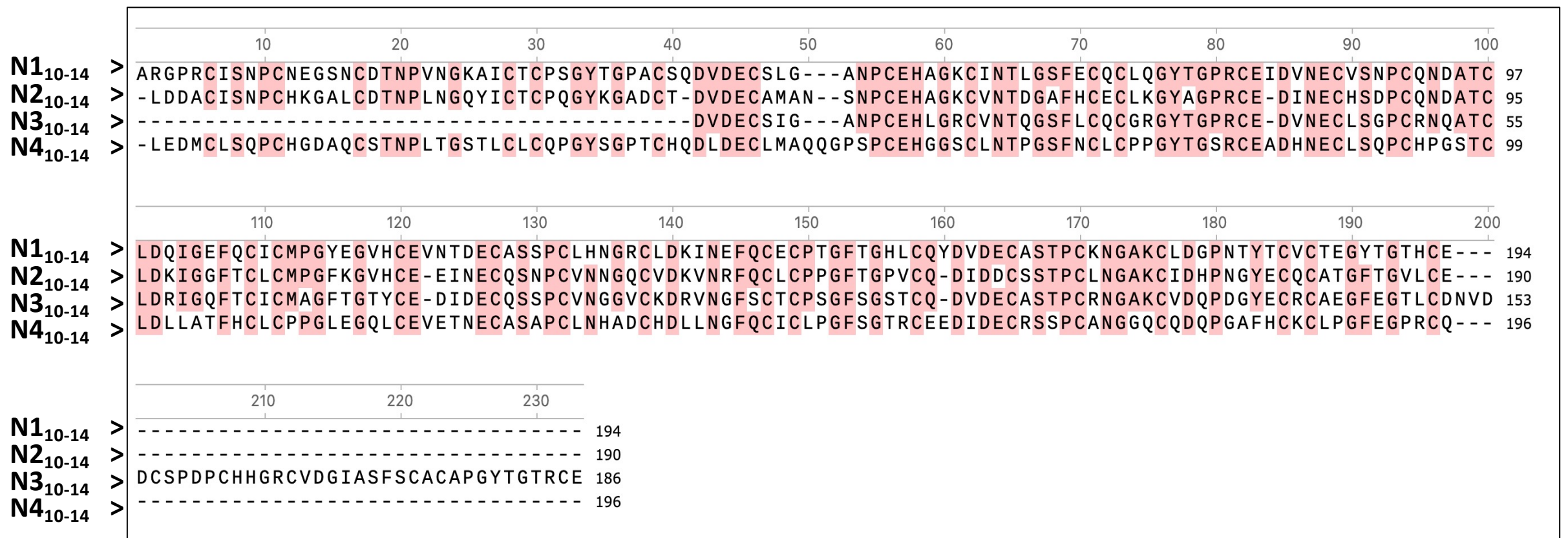
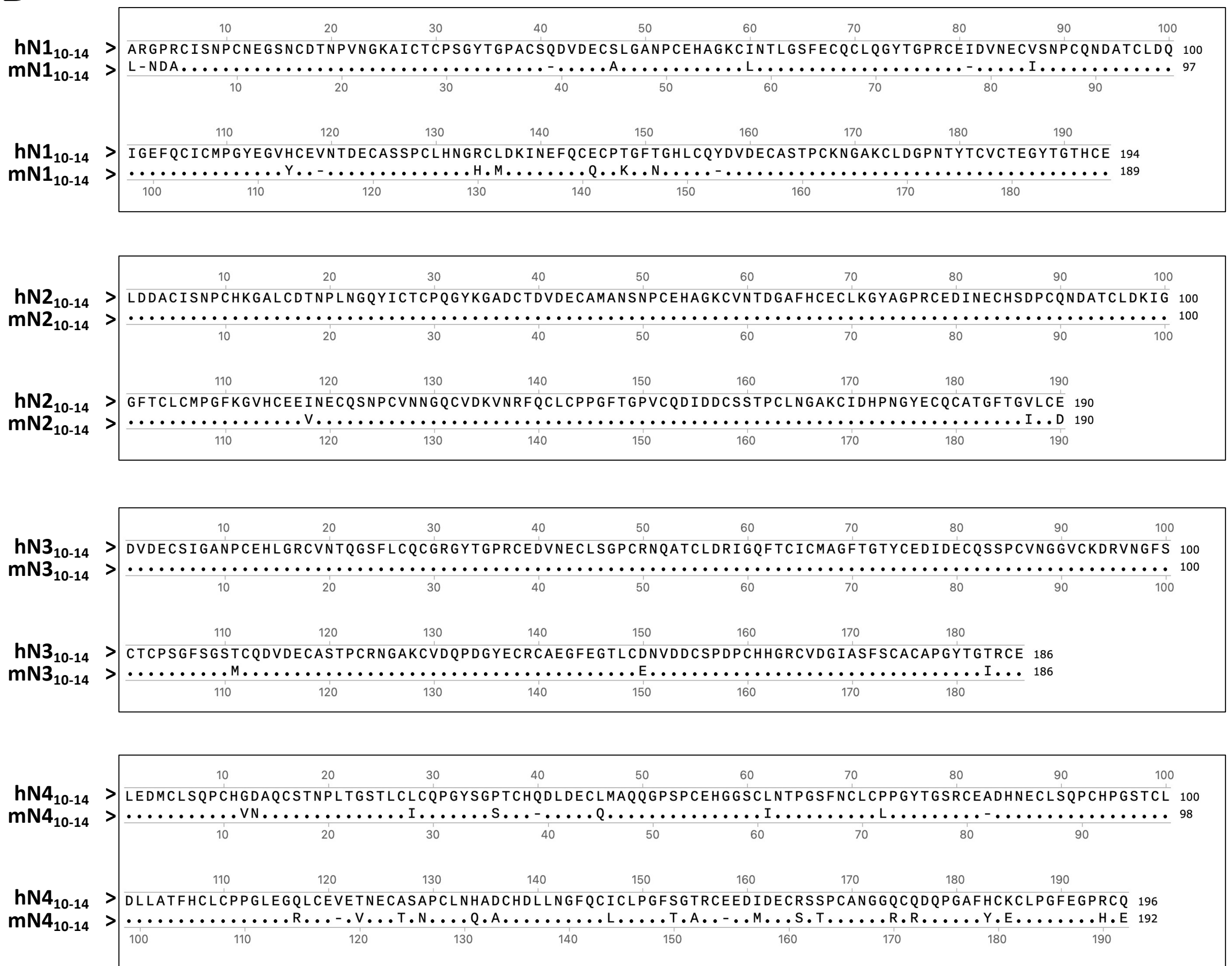
A**Human Notch core binding domain alignment****B****Human/mouse Notch core binding domain alignment**

Figure S1: Comparison of Notch putative core binding domains within and between species. (A) Amino acid alignment of EGFs 10-14 of the four human Notch receptors. Pink highlighting indicates identity, dashes indicate lack of alignment. (B) Amino acid alignment of EGFs 10-14 of each human Notch protein with its murine ortholog. Dots indicate amino acid identity.

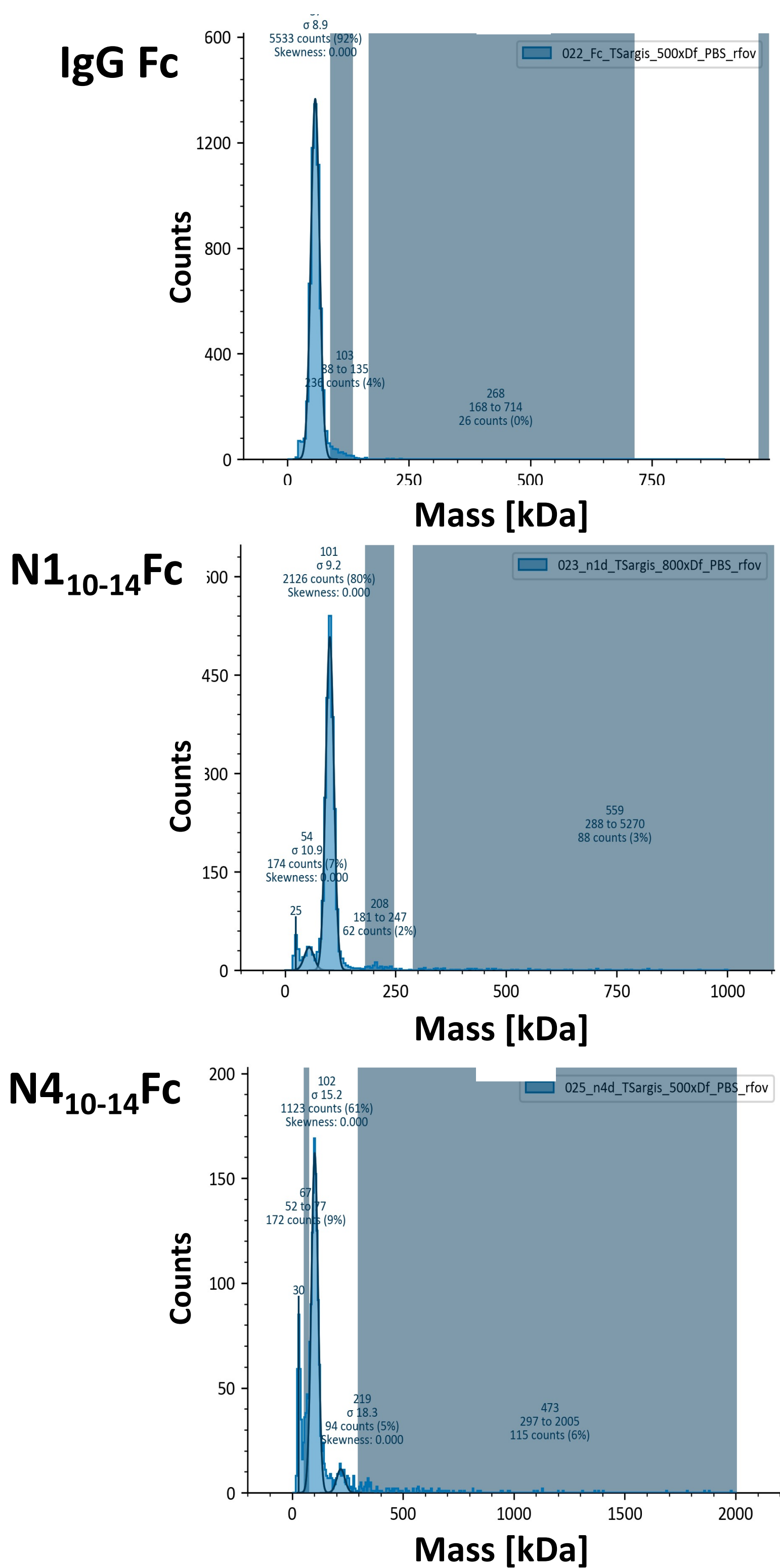


Figure S2: Mass photometry. molecular mass distribution of purified IgG Fc, N1₁₀₋₁₄Fc, and N4₁₀₋₁₄Fc as measured by mass photometry. Blue shaded regions indicate detectable oligomerization states and their respective percentage of population.

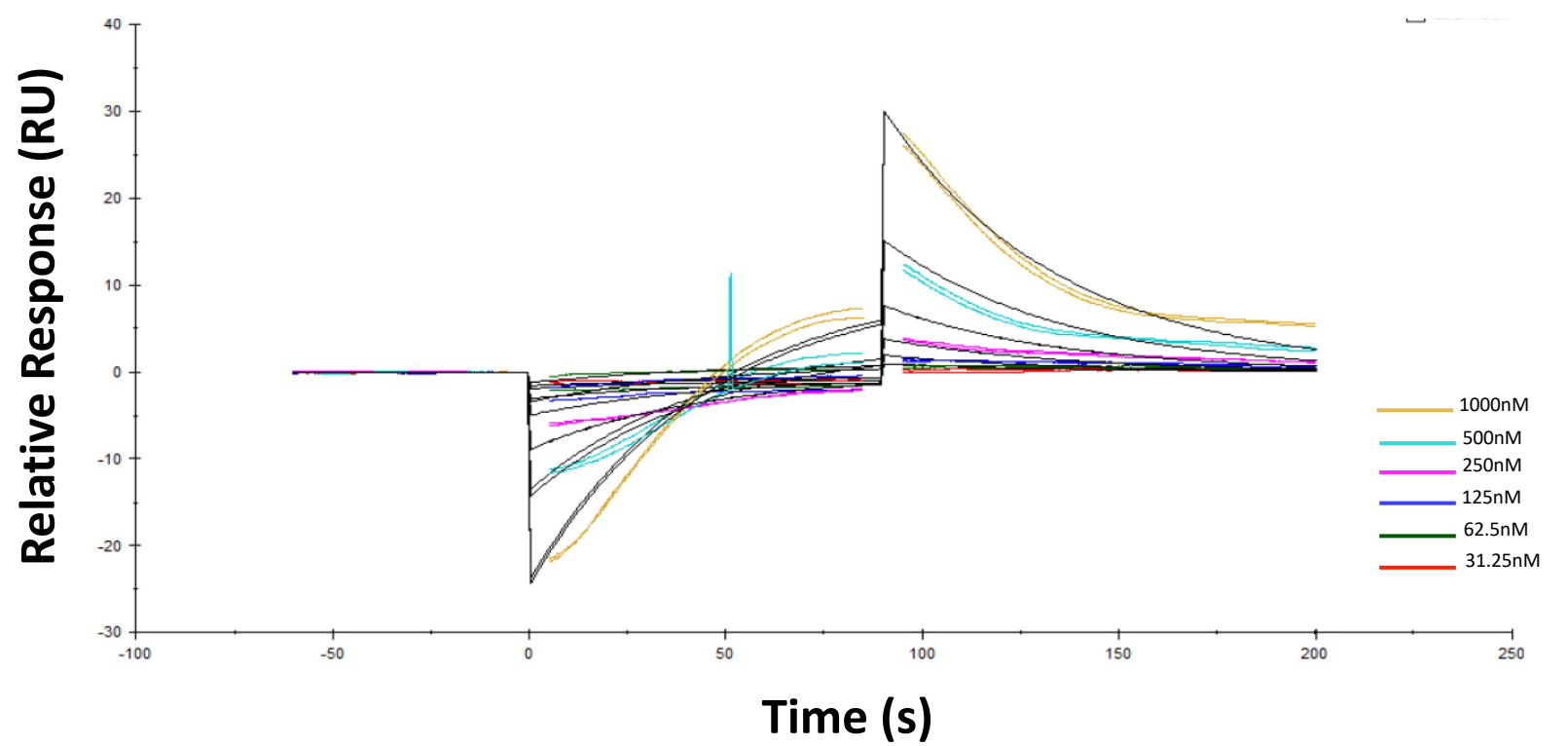
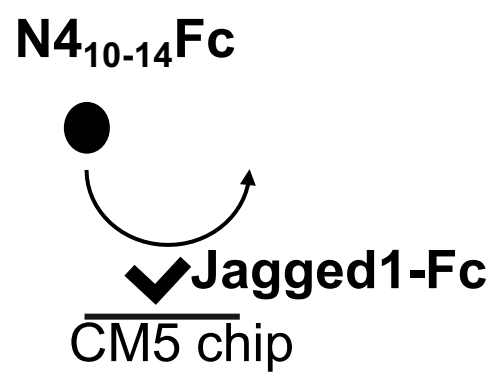
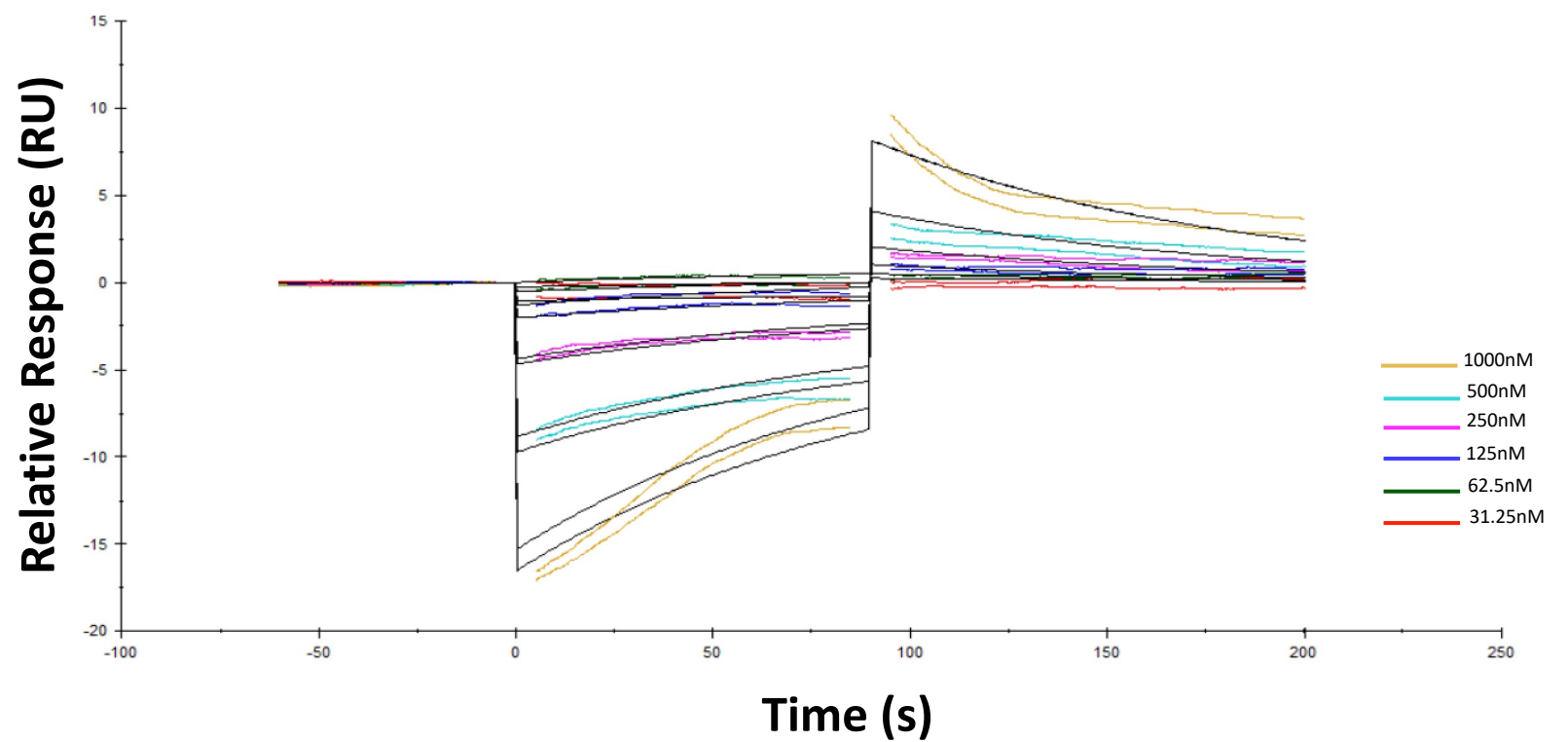
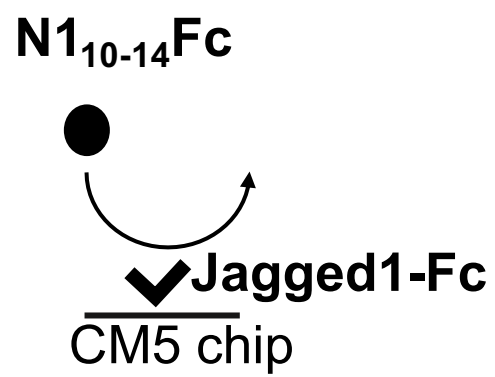


Figure S3: Binding of hJAG1-Fc to N1₁₀₋₁₄Fc and N4₁₀₋₁₄Fc. SPR measurements show binding affinity of N1₁₀₋₁₄Fc and N4₁₀₋₁₄Fc to hJAG1. Recombinant Fc-tagged hJAG1 was immobilized on the sensor chip using amine coupling and multi-cycle kinetic experiments were performed using increasing concentrations of either N1₁₀₋₁₄Fc or N4₁₀₋₁₄Fc. Binding was insufficient to allow for reliable calculation of K_d values.

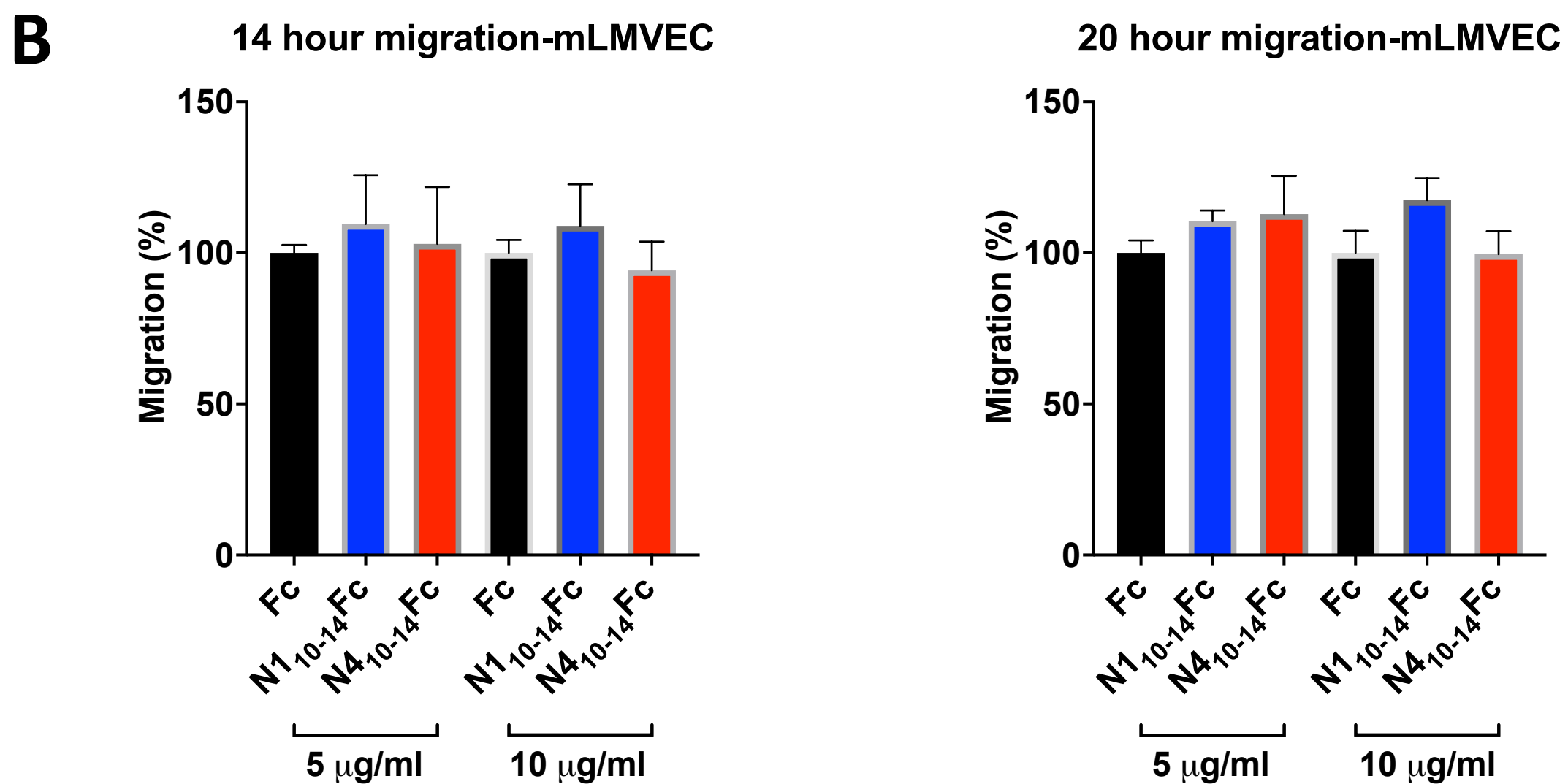
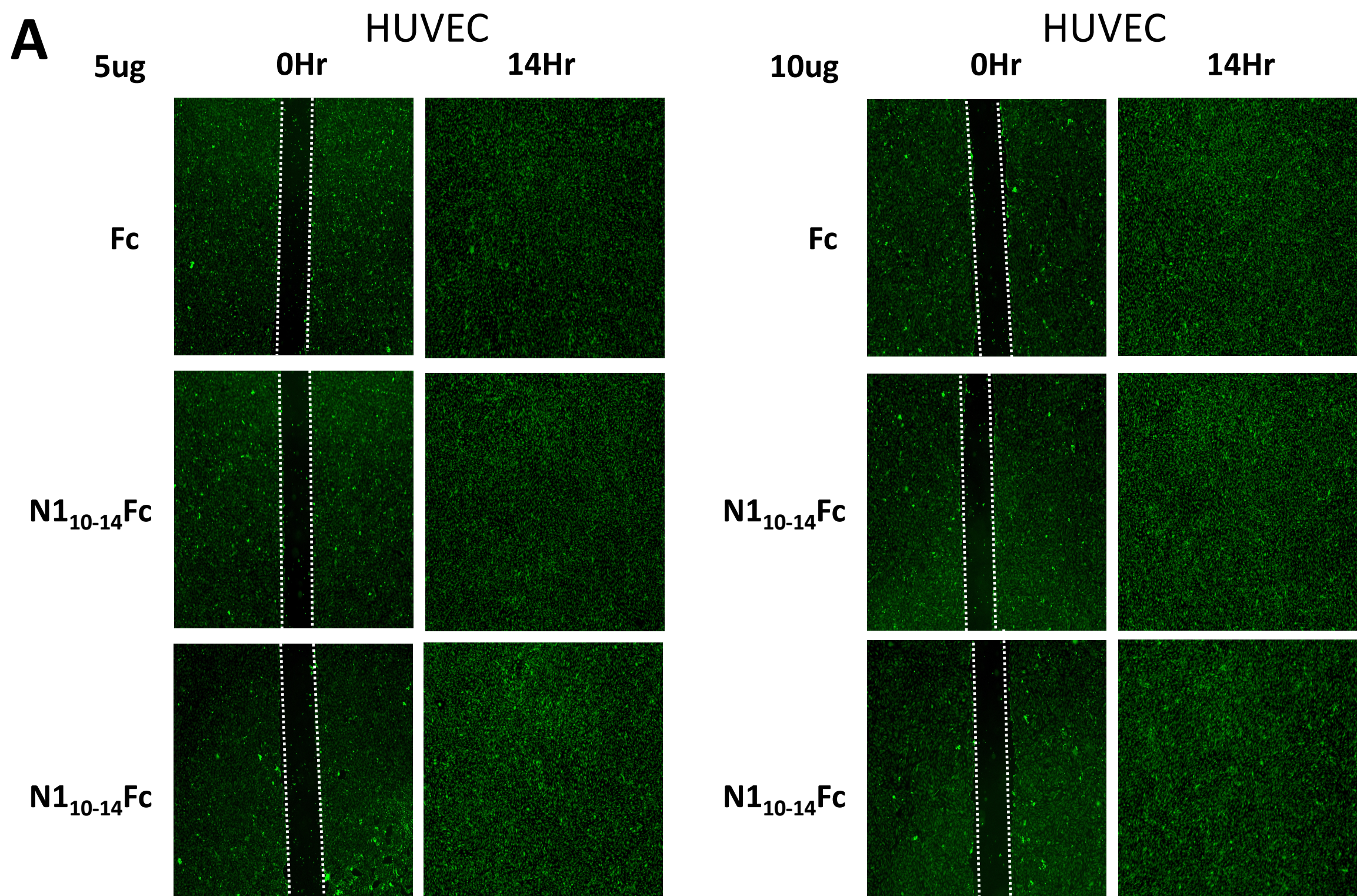


Figure S4: Wound healing assay of endothelial cells treated with Notch peptibodies. (A) HUVEC treated with cell-tracker CMFDA dye were seeded in 24-well plates and “scratch-wounded” using a 200 μ l pipette tip. After wounding, cells were treated with different concentrations (5 or 10 μ g/ μ l) of either IgG Fc, N1₁₀₋₁₄Fc or N4₁₀₋₁₄Fc. After 14 hours, microscopy was used to image cell migration into the scratch region. (B) Quantitation of identical experiments performed with mouse endothelial cells (mLMVEC) and assessed at 14 and 20 hours.

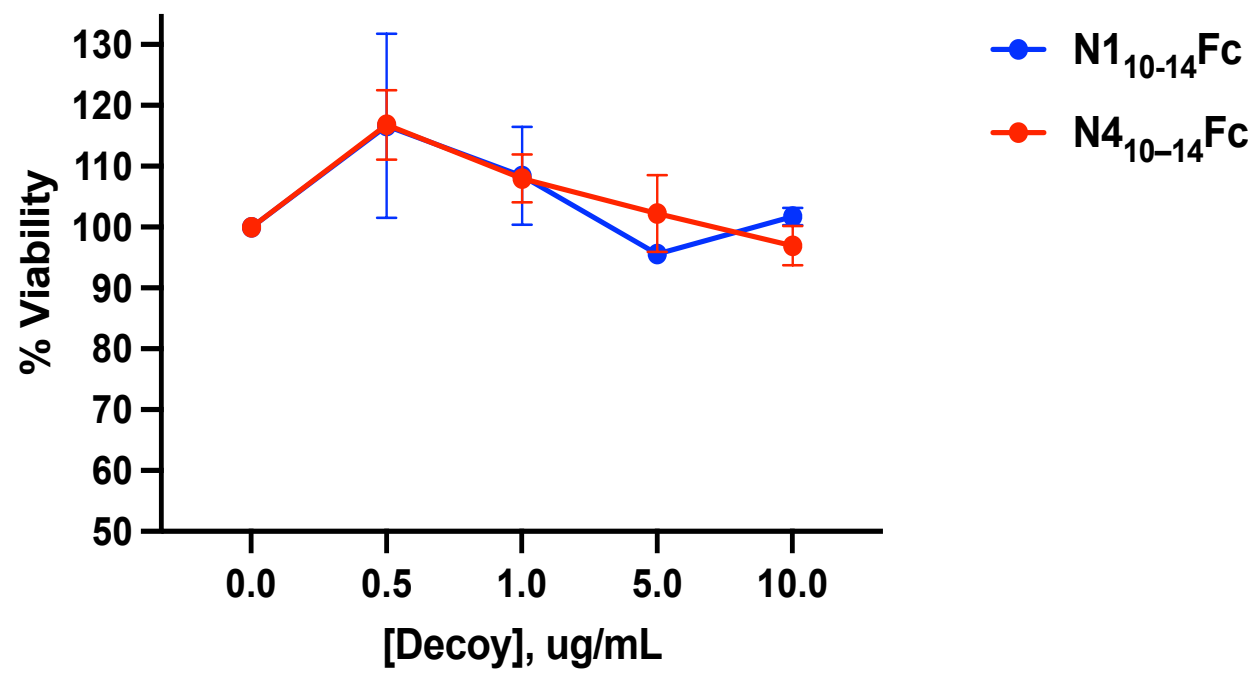
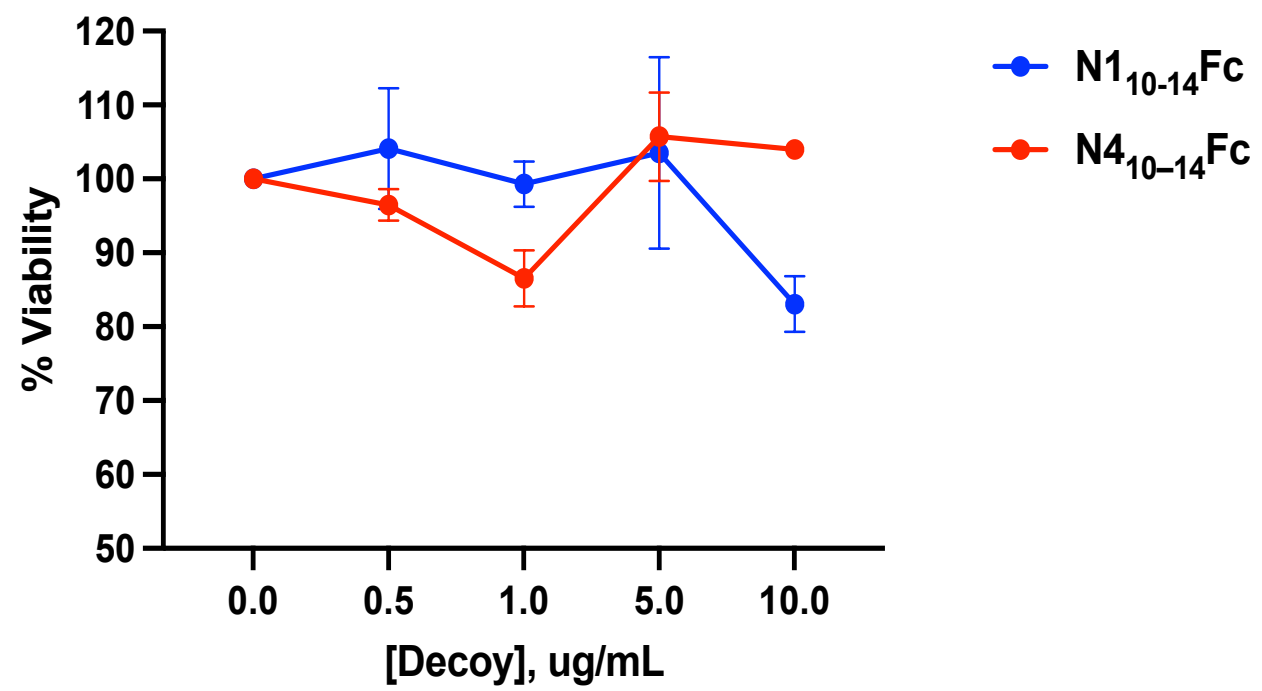
A**KopTK****B****T6E**

Figure S5: Viability assay of human and mouse T cell acute lymphoblastic leukemia (T-ALL) treated with Notch peptibodies. (A) human KopTK and (B) mouse T6E T-ALL cells were seeded at 8×10^3 cells per well in a 96-well plate in RPMI media and treated with the indicated concentration of IgG Fc, N1₁₀₋₁₄Fc or N4₁₀₋₁₄Fc for 72 hours. Viability in peptibody treated cells was assessed as the percent viable cells relative to the IgG Fc treated control group. Error bars indicate standard error of mean. For T-ALL experiments, n=3 for panel A and n=2 for panel B, no differences were significant.

IgG Fc (12.5 mg/kg)

N1₁₀₋₁₄Fc (12.5 mg/kg)

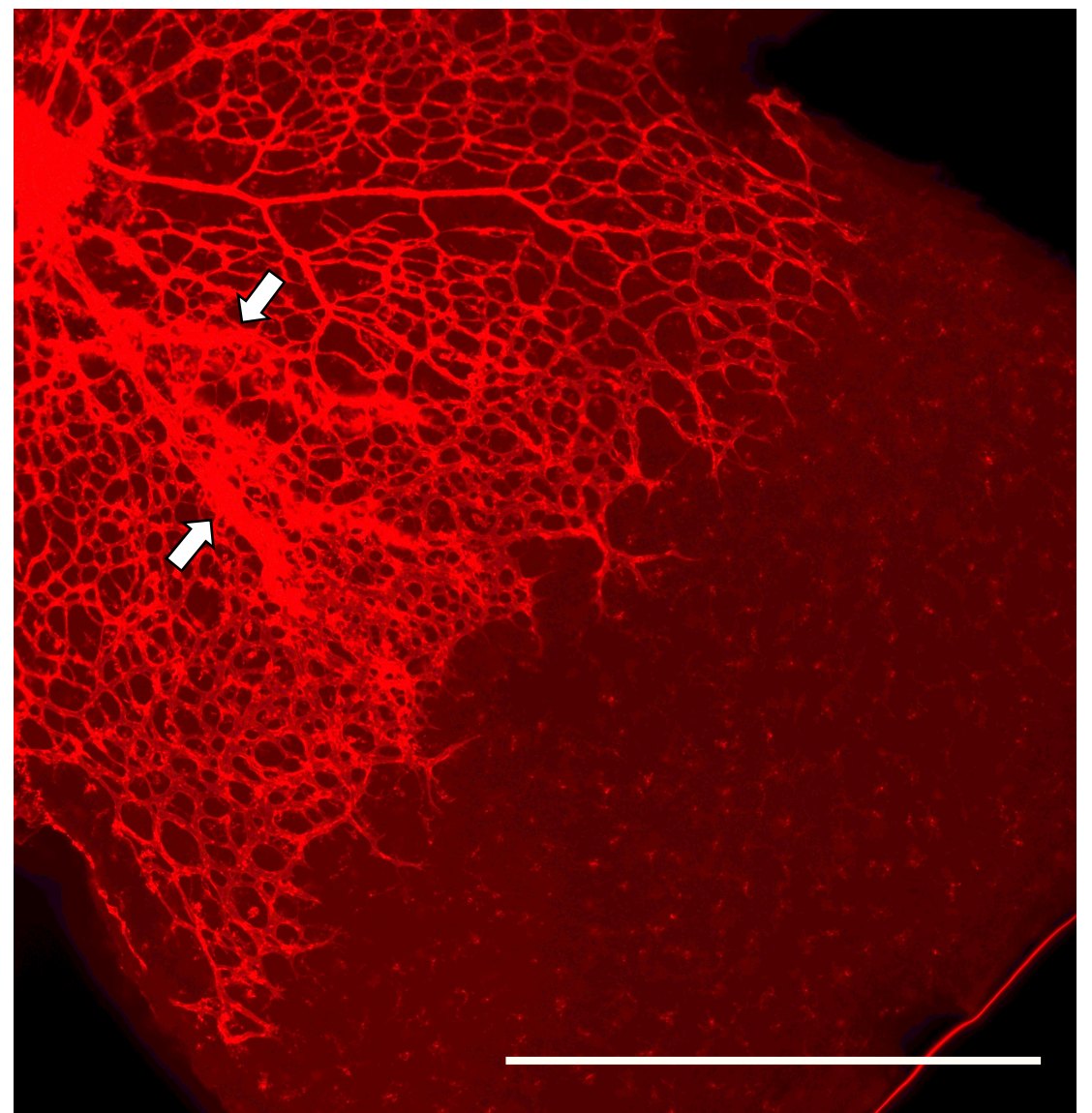
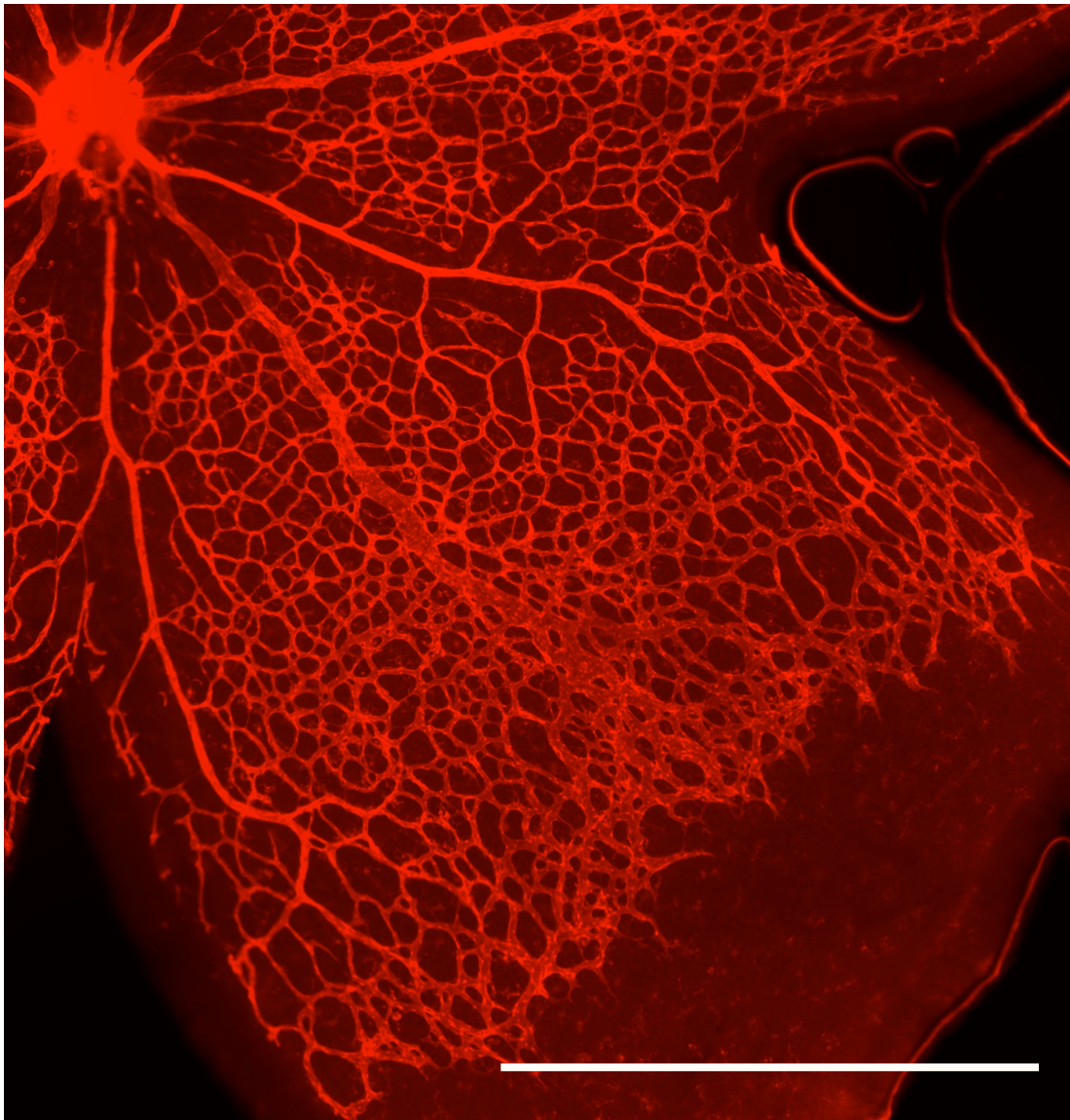


Figure S6: N1₁₀₋₁₄Fc induces abnormal enlargement of veins *in vivo*. C57BL/6 mice were injected intragastrically with 12.5 mg/kg of recombinant N1₁₀₋₁₄Fc decoy or IgG Fc for three days postnatally (P1-P3). (A) Representative images showing post-natal day 5 (P5) retinal vasculature stained with Isolectin B4 (red). (A) Arrowed region highlights enlarged veins. Scale bars: 1000 μ m.

TABLE 1: QPCR PRIMERS

Species	Gene	Forward Sequence (5'-3')	Reverse Sequence (5'-3')
Human	ACTB	CGAGGCCAGAGCAAGAGAG	CTCGTAGATGGGCACAGTGTG
Human	HEY1	ATCTGCTAAGCTAGAAAAAGCCG	GTGCGCGTCAAAGTAACCT
Human	HEY2	GCCCCCCTTGTCAGTATC	CCAGGGTCGGTAAGGTTTATTG
Human	HES1	CCTGTCATCCCCGTCTACAC	CACATGGAGTCGCGGTAA
Human	NRARP	TCAACGTGAACTCGTTCGGG	ACTTCGCCTGGTGATGAGAT
Human	RND1	CTATCCAGAGACCTATGTGCC	CGGACATTATCGTAGTAGGGAG
Human	CCL2	AGGTGACTGGGGCATTGAT	GCCTCCAGCATGAAAGTCTC
Human	CCL8	TTCTGTGCCTGCTGCTCATG	TTGGATGTTGGTGATTCTTGTGTAG
Human	NOTCH1	GTCATCTCCGACTTCATCTACC	CGGAATCAGAGCGTGAGTAG
Human	NOTCH4	AGATAAATGGGGGAAAAGTGGCG	CCTGGGCATCTTTATCGGCT

Table S1: Primers used in RT-qPCR. Table of sequences of qPCR primers.