Supplementary Table 1. List of genes co-amplified in cases with NCSTN gene amplification (TCGA, Nature 2012)

Official gene symbol	Number of cases in ``Breast Invasive Carcinoma (TCGA, Nature 2012)'' study where both the gene and NCTSN are amplified	Number of cases in ``Breast Invasive Carcinoma (TCGA, Nature 2012)'' study where the gene is amplified	Number of cases in the study that report amplified genes	p-value corresponds to statistical significance of the value in the second column	p-value in the sixth column corresponds to the statistical significance of number of cases where a gene is amplified in cases when nicastrin is not amplified
ARHGAP30	38	45	825	<< 0.01	~1
CD244	38	42	825	<< 0.01	~1
CD48	38	40	825	<< 0.01	~1
DEDD	38	44	825	<< 0.01	~1
F11R	38	45	825	<< 0.01	~1
ITLN1	38	42	825	<< 0.01	~1
ITLN2	38	44	825	<< 0.01	~1
KLHDC9	38	44	825	<< 0.01	~1
LY9	38	42	825	<< 0.01	~1
NIT1	38	44	825	<< 0.01	~1
PFDN2	38	44	825	<< 0.01	~1
PVRL4	38	45	825	<< 0.01	~1
SLAMF6	38	38	825	<< 0.01	~1
SLAMF7	38	40	825	<< 0.01	~1
TSTD1	38	45	825	<< 0.01	~1
USF1	38	45	825	<< 0.01	~1
PEX19	39	39	825	<< 0.01	~1
СОРА	40	40	825	<< 0.01	~1
NHLH1	40	40	825	<< 0.01	~1
SUMO1P3	40	40	825	<< 0.01	~1
VANGL2	40	40	825	<< 0.01	~1

Supplementary Table 2. TP53 and PIK3CA mutation correlation with NCSTN amplification in the TCGA, Nature 2012 dataset

Official gene symbol	Number of cases in ``Breast Invasive Carcinoma (TCGA, Nature 2012)" study where gene is mutated and NCTSN is amplified	Number of cases in ``Breast Invasive Carcinoma (TCGA, Nature 2012)" study where the gene is mutated.	Total number of cases in the study where mutated genes are reported	p-value corresponds to statistical significance of the value in the second column	p-value corresponds to statistical significance of number of cases where a gene is mutated in cases when nicastrine is not amplified
TP53	15	187	507	0.0482	0.9799
PIK3CA	4	178	507	0.9971	0.0113

Supplementary Table 3.
Predictors of 10-year disease free survival on multivariate analysis in a Cox regression model

Characteristic	Cox p value	Hazard Ratio	
Grade	0.411	1.625	
Stage	<0.001	1.967	
Size	0.291	1.479	
Nicastrin RNAScope H-score	0.044	1.352	

Supplementary Table 4. Correlation of Nicastrin RNAScope H-score with clinical and tumor parameters

Variable/biomarker	Nicastrin RNAScope H-Score		<i>p</i> -value	
	Low (<120) Number (%)	High (≥120) Number (%)		
Patients age (years)				
<u>≤50</u>	50 (33.1)	83 (51.2)	0.005	
>50	98 (64.9)	77 (47.5)	0.005	
Grade				
1	1 (0.7)	0 (0.0)		
2	19 (12.6)	12 (7.4)	0.175	
3	131(86.8)	150 (92.6)		
stage				
1	99 (65.6)	93 (58.1)		
2	35 (23.2)	47(29.4)	0.381	
3	17 (11.3)	20 (12.5)		
ize				
<1.5cm	46 (30.9)	35(22.9)	0.117	
≥1.5cm	103(69.1)	118(77.1)	0.117	
Pleomorphism				
2	4 (2.7)	0 (0.0)	0.036	
3	144 (97.3)	161 (100)	0.000	
litosis				
	16(10.8)	7(4.3)		
	28(18.9)	21 (13.0)	0.023	
	140(70.3)	133(82.6)		
licastrin Membrane Score	14 (10.0)	14 (11.0)		
)	14 (13.2)	14 (11.3)		
1	51 (48.1)	45 (37.9)	0.055	
2	31 (29.2)	37 (28.2)	0.056	
3	10 (9.4)	28 (22.6)		
licastrin Cytoplasmic Score				
0	11 (10.3)	7(5.6)		
1	32(29.9)	36 (29.0)	0.414	
2	49 (45.8)	56 (45.2)	0.414	
3	15 (14.0)	25 (20.2)		
Ci67				
Negative	28 (18.9)	11(7.0)	0.002	
Positive	120 (81.1)	146 (93.0)	0.002	
E-cadherin				
legative	30 (24.0)	18 (12.1)	0.01	
Positive	95 (76.0)	131 (78.9)		
BRCA1cvto				
	49 (29.9)	44 (29.9)		
Negative	49 (29.9)	44 (27.7)		

Supplementary Figure Legends

Supplementary Figure 1. (a) FACS analysis of hybridoma supernatants (Genovac). Antibodies were raised by genetic immunisation of WISTAR rats using the cDNA, encoding for the ECD of nicastrin (34-669 a.a). This cDNA was cloned into a proprietary GENOVAC expression vector developed to maximise an immune response in mammals and to allow screening of the resulting immune sera. The immunisation constructs were introduced into the animals using a Genegun. Following DNA boosts, test sera were taken from the animals and tested for the presence of antibodies in a transient transfection assay in flow cytometry against the corresponding GENOVAC screening vector. Labels bellow the histograms annotate the MAb clone reference number. (b) FACS analysis assessing the binding efficacy of the 21 anti-nicastrin MAb clones. MDA-MB-231 cells were grown to 70-80% confluence. Cells were incubated with the supernatants obtained from the first round of hybridoma cloning and with the rat IgG2b as the isotype control antibody at an arbitrary concentration of 50 µg/ml. The concentration of the mAbs in the preparations was at this phase unknown. Secondary anti-Rat-FITC conjugated antibody was used. FACS analysis was performed using the FACSCalibur, BD. The labels in the top left-hand corner of the histograms annotate the MAb clone reference number. Blue: Rat IgG2b isotype control; Green: MAb supernatants. (c) Anti-nicastrin monoclonal antibodies affect MDA-MB-231 cell invasion. MDA-MB-231 cells were treated with indicated anti-NCT MAbs and subjected to the transwell invasion assay for 72 h. Bars represent mean number of invaded cells ± SEM from three separate experiments, each in triplicate. Statistical difference from IgG2b isotype control antibody. 1H4, *p = 0.01; 2H6, *p = 0.0005; 6H5, *p = 0.01; 6H11, *p = 0.01; 9G7, *p = 0.01; 10C5, *p = 0.02; 10C11, *p = 0.004. Anti-nicastrin PcAb (100 µg/ml) was used as a positive control, p < 0.001.

Supplementary Figure 2. K_d determination (a, b) Whole IgG 10C11 (a) and 2H6 (b) were directly immobilised to a BIACore 3000 CM5 chip and NCT-Fc fusion protein was flowed over at a concentration range of 1000-62.5 nM. Replicate injections and double referencing was used and the data fitted to a 1:1 binding model using BIAevaluation v2.0 to determine the kinetic constants and K_d . Clone 10C11 has a significantly better association rate compared to clone 2H6, whereas 2H6 has a slower dissociation rate. Kinetic values are

shown in Figure 1-Table 1. (c,d) IgG clones 10C11 and 2H6 (c) and 1E2 and 10C5 (d) were analyses by flow cytometry on MDA-MB-231 cells at a concentration range of 200 mg/ml to 19 ng/ml. The mean fluorescent intensity (MFI) was fitted to a sigmoidal binding model to determine the cellular binding K_d which are incorporated in Table 1-Figure 1.

Supplementary Figure 3. (a) Binding of anti-NCT MAbs to endogenous cell surface nicastrin of mouse and rat cells. Non-permeabilized mouse melanocytes and rat fibroblast cells were incubated with 50 µg/ml of 2H6 and 10C11 antibodies, followed by the incubation with the secondary anti-rat FITC antibody. Rat IgG antibody was used as control. Binding was assessed by FACS. (b) Mouse weights (grams) during the metastatic in vivo mouse model experiment. (c-f) Biochemistry analysis from mouse serum collected at termination of animals in the metastatic in vivo mouse model experiment. Kidney function was assessed through creatinine and urea measurements, while liver function test were assessed through alanin aminotransferase (ALT) and alkaline phosphatise (Alk Phos).

Supplementary Figure 4. (a-e) Mann-Whitney Test was used to compare all conditions to one another for each time point and all p values summarised, highlighting significant differences (p<0.05).

Supplementary Figure 5. Nicastrin gene amplification and high mRNA levels predict worse survival of breast cancer patients. (a) Kaplan Meier overall survival curves showing worse outcome of patients with nicastrin gene amplification in the Nature TCGA 2012 dataset (cBio Portal for Cancer Genomics). (b) Correlation of nicastrin gene amplification and mRNA levels in the Nature TCGA 2012 dataset. (c) Bright field microscopy images (magnification x40) of nicastrin RNAScope analyses on paraffin embedded breast cancer tissue microarrays. Representative images are show for four different levels of the H-score. Cut-off value = 120. (d) Kaplan Meier analysis showing worse disease free survival of breast cancer patients with the Nicastrin RNAScope scope \geq 120, p = 0.019.

Fig S1

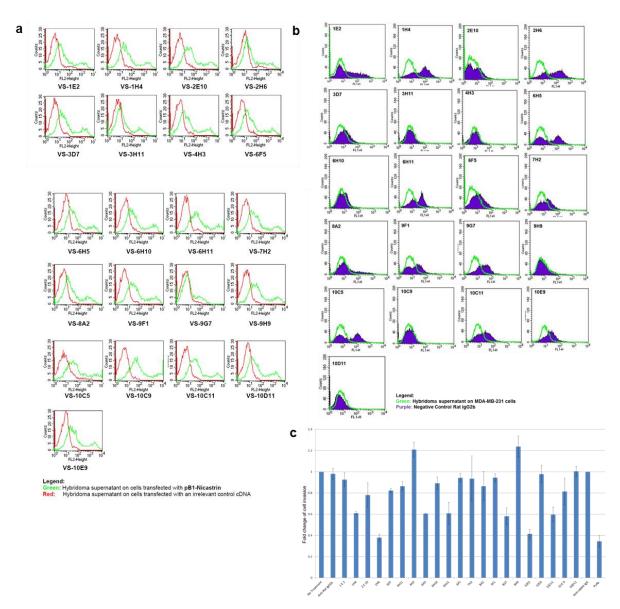
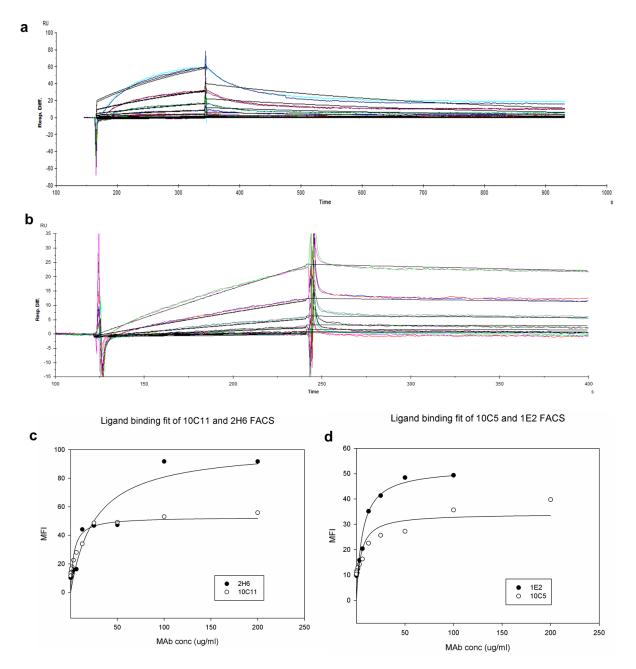


Fig S2







а	DAY 6	PBS	lsotype	2H6	10C11	GSI
a	PBS		1.0000	0.6126	0.0401	0.7551
	lsotype			0.6454	0.0104	0.8329
	2H6				0.0148	0.6601
	10C11					0.0062
	GSI					
b	DAY 12	PBS	lsotype	2H6	10C11	GSI
N N	PBS		0.0289	0.0541	0.0093	0.1807
	lsotype			0.5054	0.3282	0.1419
	2H6				0.2345	0.2824
	10C11					0.0813
	GSI					
С	DAY 18	PBS	lsotype	2H6	10C11	GSI
Ŭ	PBS		0.4557	0.1520	0.2810	0.1014
	lsotype			0.3357	0.9551	0.2949
	2H6				0.3282	0.4908
	10C11					0.2284
	GSI					
d	DAY 24	PBS	lsotype	2H6	10C11	GSI
	PBS		1.0000	0.2319	0.8665	0.4452
	lsotype			0.2345	0.7984	0.2824
	2H6				0.1605	0.9497
	10C11					0.2448
	GSI					
е	DAY 30	PBS	lsotype	2H6	10C11	GSI
	PBS		0.6282	0.0027	0.0813	0.0931
	lsotype			0.0006	0.0541	0.0513
	2H6				0.0740	1.0000
	10C11					0.4136
	GSI					

Fig S4.

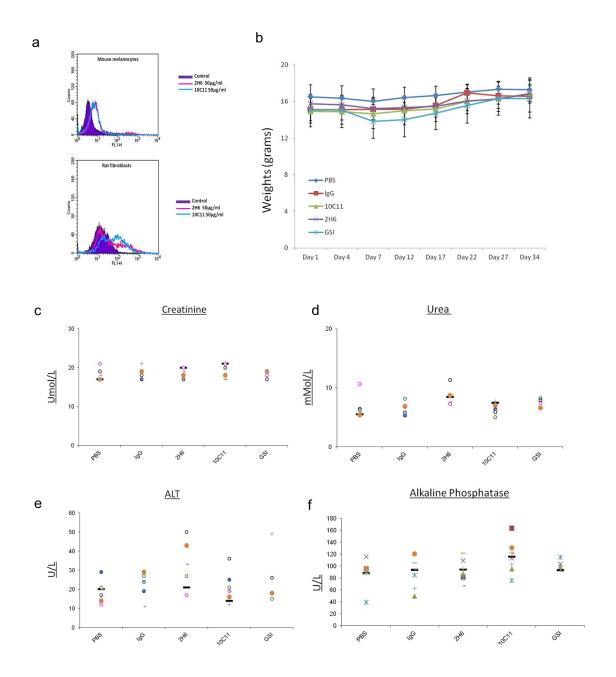


Fig S5

