

# Additional file 1 to “A random effects meta-analysis model with Box-Cox transformation”

## 1 R code

Our Bayesian analyses in the simulation study and the application were implemented by using the R software. In particular, we used `rstan` package which provides an interface to Stan. The Stan is a programming language for implementing Bayesian inference with the MCMC sampling. We below provide R functions `nremeta` and `bcremeta` for the Bayesian estimations of the normal random effects model and the proposed model respectively.

### 1.1 R function for normal random effects model: `nremeta`

**Description:** Function to implement Bayesian estimation of normal random effects model using MCMC sampling.

**Usage:** `nremeta(ns, yi, vi, iter.num, chain.num, burnin, thin)`

**Arguments:** `ns` = number of studies; `yi` = vector of observed treatment effect estimates; `vi` = vector of observed within-study variances for treatment effect estimates; `iter.num` = number of samples drawn within each chain of MCMC; `chain.num` = number of chains in MCMC; `burnin` = number of burn-in samples within each chain of MCMC; `thin` = thinning interval between consecutive samples within each chain of MCMC.

**Returned list elements:** `theta` = posterior mean, standard deviation (s.d.), median, lower and upper limit of 95 per cent credible interval for overall mean effect; `theta.samp` = samples of overall mean effect from its posterior distribution; `tau` = posterior mean, s.d., median, lower and upper limit of 95 per cent credible interval for square root of between-study variance; `I2` = posterior mean, s.d., median, lower and upper limit of 95 per cent credible interval for  $I^2$ ; `pred` = mean, s.d. and median of samples from predictive distribution, and lower and upper limit of 95 per cent prediction interval; `pred.samp` = samples from predictive distribution.

### Source code:

```
library(rstan)

make.df <- function(parsamp)
{
  no.parsamp <- na.omit(parsamp)
  dataf <- data.frame(
    mean=mean(no.parsamp), sd=sd(no.parsamp)
    ,median=quantile(no.parsamp,probs=0.500)[[1]]
    ,CIlower=quantile(no.parsamp,probs=0.025)[[1]]
    ,CIupper=quantile(no.parsamp,probs=0.975)[[1]]
  )
}
```

```

    return(dataf)
  }

normal <- '
  data {
    int<lower=0> k;
    real yi[k];
    real<lower=0> vi[k];
  }
  parameters {
    real theta;
    real<lower=0> tau;
  }
  model {
    for(i in 1:k)
      yi[i] ~ normal(theta,sqrt(tau*tau+vi[i]));
    theta ~ normal(0,10000);
    tau ~ uniform(0,10);
  }
}'
nmeta <- stan(model_code=normal)

nremeta <- function(ns,yi,vi,iter.num,chain.num,burnin,thin)
{
  standata <- list(k=ns,yi=yi,vi=vi)
  res <- stan(fit=nmeta,data=standata,iter=iter.num,chains=chain.num,warmup=burnin,thin=thin)

  ntheta <- extract(res,par="theta",permuted=TRUE)$theta
  ntau <- extract(res,par="tau",permuted=TRUE)$tau

  wi <- 1/vi
  nsigma <- sum(wi*(ns-1))/(sum(wi)^2-sum(wi^2))
  nI2 <- ntau^2/(ntau^2+nsigma)*100
  npred <- rnorm(length(ntheta),mean=ntheta,sd=ntau)

  ntheta.df <- make.df(ntheta)
  ntau.df <- make.df(ntau)
  nI2.df <- make.df(nI2)
  npred.df <- make.df(npred)
  ntheta2.df <- data.frame(samples=ntheta)
  npred2.df <- data.frame(samples=npred)

  return(list(
    theta=ntheta.df,theta.samp=ntheta2.df,tau=ntau.df
    ,I2=nI2.df,pred=npred.df,pred.samp=npred2.df))
}

```

## 1.2 R function for random effects model with Box-Cox transformation: bcremeta

**Description:** Function to implement Bayesian estimation of random effects model with Box-Cox transformation using MCMC sampling.

**Usage:** `bcremeta(ns, yi, vi, grid.alpmin, grid.lambda, iter.num, chain.num, burnin, thin)`

**Arguments:** `ns` = number of studies; `yi` = vector of observed treatment effect estimates; `vi` = vector of observed within-study variances for treatment effect estimates; `grid.alpmin` = vector of candidate values of grid search procedure for estimating shift parameter, which is specified as minimum values of  $\{(y_i + \alpha) : i = 1, \dots, k\}$ ; `grid.lambda` = vector of candidate values of grid search procedure for estimating transformation parameter; `iter.num` = number of samples drawn within each chain of MCMC; `chain.num` = number of chains in MCMC; `burnin` = number of burn-in samples within each chain of MCMC; `thin` = thinning interval between consecutive samples within each chain of MCMC.

**Returned list elements:** `median` = posterior mean, standard deviation (s.d.), median, lower and upper limit of 95 per cent credible interval for overall median effect; `median.samp` = samples of overall median effect from its posterior distribution; `IQR` = posterior mean, s.d., median, lower and upper limit of 95 per cent credible interval for IQR of random effects distribution ( $\xi_{75} - \xi_{25}$ ); `nIQR` = posterior mean, s.d., median, lower and upper limit of 95 per cent credible interval for normalised IQR of random effects distribution ( $(\xi_{75} - \xi_{25}) / (z_{75} - z_{25})$ ); `IQRrs` = posterior mean, s.d., median, lower and upper limit of 95 per cent credible interval for ratio of IQR squares ( $(\xi_{75} - \xi_{25})^2 / (\nu_{75} - \nu_{25})^2$ ); `pred` = mean, s.d. and median of samples from predictive distribution, and lower and upper limit of 95 per cent prediction interval; `pred.samp` = samples from predictive distribution; `tau` = posterior mean, s.d., median, lower and upper limit of 95 per cent credible interval for square root of between-study variance of random effects after transformation; `lambda` = estimate of transformation parameter; `alpha` = estimate of shift parameter; `alpmin` = estimate of  $\min\{(y_i + \alpha) : i = 1, \dots, k\}$ .

### Source code:

```
library(rstan)

make.df <- function(parsamp)
{
  no.parsamp <- na.omit(parsamp)
  dataf <- data.frame(
    mean=mean(no.parsamp), sd=sd(no.parsamp)
    ,median=quantile(no.parsamp,probs=0.500)[[1]]
    ,CIlower=quantile(no.parsamp,probs=0.025)[[1]]
    ,CIupper=quantile(no.parsamp,probs=0.975)[[1]]
  )
  return(dataf)
}

bc.like <- function(k,y,s2,lam,alp) { return(
  function(para) {
    mu <- para[1]
    tau <- para[2]
    ygm <- prod(y+alp)^(1/k)
    y1 <- ((y+alp)^lam-1)/(lam*ygm^(lam-1))
    phi2 <- s2/ygm^(2*lam-2)*(lam*ygm^(lam-1)*mu+1)^(2-2/lam)
    return(sum(-0.5*log(tau^2+phi2)-0.5/(tau^2+phi2)*(y1-mu)^2))
  }
}
```

```

)}}

boxcox <- '
data {
  int<lower=0> k;
  real yi[k];
  real ygm;
  real<lower=0> vi[k];
  real lambda;
  real alpha;
}
parameters {
  real mu;
  real<lower=0> tau;
}
model {
  real ylami;
  real phi2;
  for (i in 1:k) {
    ylami <- (pow(yi[i]+alpha,lambda)-1)/(lambda*pow(ygm,lambda-1));
    phi2 <- vi[i]/pow(ygm,2*lambda-2)*pow(lambda*pow(ygm,lambda-1)*mu+1,2-2/lambda);
    increment_log_prob(-0.5*log(tau*tau+phi2)-0.5/(tau*tau+phi2)*pow(ylami-mu,2));
  }
  mu ~ normal(0,10000);
  tau ~ uniform(0,10);
}',
bcmeta <- stan(model_code=boxcox)

bcmemeta <- function(ns,yi,vi,grid.alpmin,grid.lambda,iter.num,chain.num,burnin,thin)
{
  glambda <- grid.lambda[grid.lambda!=0]

  if(min(yi)>0) {

    mutau <- array(0,dim=c(4,length(glambda)))
    alpha <- 0
    ygmean <- prod(yi)^(1/ns)
    transf <- apply(as.matrix(glambda),1,function(a) {
      return((yi^a-1)/(a*ygmean^(a-1)))
    })
    transf.niqr <- apply(transf,2,function(a) {
      nq3 <- qnorm(0.75,0,1)
      nq1 <- qnorm(0.25,0,1)
      return(IQR(a)/(nq3-nq1))
    })
    bcox <- cbind(apply(transf,2,median),transf.niqr)

    for(l in 1:length(glambda)) {
      lambda <- glambda[l]
      if(lambda>0) {
        bclim <- -1/(lambda*ygmean^(lambda-1))

```

```

low <- c(bclim+0.01,0)
upp <- c(Inf,Inf)
} else {
  bclim <- -1/(lambda*ygmean^(lambda-1))
  low <- c(-Inf,0)
  upp <- c(bclim-0.01,Inf)
}

like.init <- as.vector(bcox[l,]+0.001)
for(ll in 1:10) {
  tmp <- try(optim(
    par=like.init, fn=bc.like(ns,yi,vi,lambda,alpha),control=list(fnscale=-1)
    ,method="L-BFGS-B",lower=low,upper=upp),TRUE)

  if(class(tmp)=="try-error") {
    if(ll==10) {
      mutau[,1] <- c(0,0,lambda,-99999)
      break
    } else {
      like.init <- like.init+0.001
    }
  } else {
    mutau[,1] <- c(tmp$par,lambda,tmp$value)
    break
  }
}
}
mle <- c(mutau[1:3,order(-mutau[4,])[1]],alpha)

} else {

mutau <- array(0,dim=c(4,length(glambda)))
pml <- array(0,dim=c(5,length(grid.alpmin)))

for(m in 1:length(grid.alpmin)) {
  alpha <- grid.alpmin[m]-min(yi)
  ygmean <- prod(yi+alpha)^(1/ns)
  transf <- apply(as.matrix(glambda),1,function(a) {
    return(((yi+alpha)^a-1)/(a*ygmean^(a-1)))
  })
  transf.niqr <- apply(transf,2,function(a) {
    nq3 <- qnorm(0.75,0,1)
    nq1 <- qnorm(0.25,0,1)
    return(IQR(a)/(nq3-nq1))
  })
  bcox <- cbind(apply(transf,2,median),transf.niqr)

  for(l in 1:length(glambda)) {
    lambda <- glambda[l]
    if(lambda>0) {
      bclim <- -1/(lambda*ygmean^(lambda-1))

```

```

    low <- c(bclim+0.01,0)
    upp <- c(Inf,Inf)
  } else {
    bclim <- -1/(lambda*ygmean^(lambda-1))
    low <- c(-Inf,0)
    upp <- c(bclim-0.01,Inf)
  }

  like.init <- as.vector(bcox[1,]+0.001)
  for(ll in 1:10) {
    tmp <- try(optim(
      par=like.init,fn=bc.like(ns,yi,vi,lambda,alpha),control=list(fnscale=-1)
      ,method="L-BFGS-B",lower=low,upper=upp),TRUE)

    if(class(tmp)=="try-error") {
      if(ll==10) {
        mutau[,1] <- c(0,0,lambda,-99999)
        break
      } else {
        like.init <- like.init+0.001
      }
    } else {
      mutau[,1] <- c(tmp$par,lambda,tmp$value)
      break
    }
  }
  }
  pmle[,m] <- c(mutau[,order(-mutau[4,])[1]],alpha)
}

mle <- pmle[c(1,2,3,5),order(-pmle[4,])[1]]
}

bclambda <- mle[3]
bcalpha <- mle[4]
bcalphmin <- bcalpha+min(yi)
ygmean <- prod(yi+bcalpha)^(1/ns)

standata <- list(k=ns,yi=yi,ygm=ygmean,vi=vi,lambda=bclambda,alpha=bcalpha)
init.list <- rep(list(list(mu=mle[1],tau=mle[2]+0.01)),chain.num)
res <- stan(fit=bcmeta,data=standata,iter=iter.num,chains=chain.num
  ,init=init.list,warmup=burnin,thin=thin)

bcmu <- extract(res,par="mu",permuted=TRUE)$mu
bctau <- extract(res,par="tau",permuted=TRUE)$tau

bcmed <- (bclambda*ygmean^(bclambda-1)*bcmu+1)^(1/bclambda)-bcalpha
bcxi25 <- (bclambda*ygmean^(bclambda-1)*(bcmu+bctau*qnorm(0.25,0,1))+1)^(1/bclambda)-bcalpha
bcxi75 <- (bclambda*ygmean^(bclambda-1)*(bcmu+bctau*qnorm(0.75,0,1))+1)^(1/bclambda)-bcalpha

bcIQR <- bcxi75-bcxi25

```

```

bcNIQR <- (bcxi75-bcxi25)/(qnorm(0.75,0,1)-qnorm(0.25,0,1))

bcsigma <- apply(as.matrix(bcmu),1,function(a) {
  phi2 <- vi/ygmean^(2*bclambda-2)*(bclambda*ygmean^(bclambda-1)*a+1)^(2-2/bclambda)
  wi <- 1/phi2
  return(sum(wi*(ns-1))/(sum(wi)^2-sum(wi^2)))
})

bctsd <- sqrt(bctau^2+bcsigma)
bcnu25 <- (bclambda*ygmean^(bclambda-1)*(bcmu+bctsd*qnorm(0.25,0,1))+1)^(1/bclambda)-bcalpha
bcnu75 <- (bclambda*ygmean^(bclambda-1)*(bcmu+bctsd*qnorm(0.75,0,1))+1)^(1/bclambda)-bcalpha
bcIQRrs <- (bcxi75-bcxi25)^2/(bcnu75-bcnu25)^2*100

bsamp1 <- rnorm(length(bcmu),mean=bcmu,sd=bctau)
if(bclambda<0)
  bsamp2 <- subset(bsamp1,bsamp1<(-1/(bclambda*ygmean^(bclambda-1))))
} else {
  bsamp2 <- subset(bsamp1,bsamp1>(-1/(bclambda*ygmean^(bclambda-1))))
}
bcpred <- (bclambda*ygmean^(bclambda-1)*bsamp2+1)^(1/bclambda)-bcalpha

bcmed.df <- make.df(bcmed)
bcIQR.df <- make.df(bcIQR)
bcNIQR.df <- make.df(bcNIQR)
bcIQRrs.df <- make.df(bcIQRrs)
bcpred.df <- make.df(bcpred)
bctau.df <- make.df(bctau)
bcmed2.df <- data.frame(samples=bcmed)
bcpred2.df <- data.frame(samples=bcpred)

return(list(
  median=bcmed.df,median.samp=bcmed2.df,IQR=bcIQR.df,nIQR=bcNIQR.df
  ,IQRrs=bcIQRrs.df,pred=bcpred.df,pred.samp=bcpred2.df,tau=bctau.df
  ,lambda=bclambda,alpha=bcalpha,alpmin=bcalphmin))
}

```