

# Supplementary Information

## Addressing Misclassification Bias in Vaccine Effectiveness Studies with an application to Covid-19

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### Load packages/functions

```
library(tidyverse)
library(runjags)
library(rjags)
library(rootSolve)
library(parallel)
library(kableExtra)
source("R/functions.R")
testjags()
```

```
## You are using R version 4.2.2 (2022-10-31 ucrt) on a windows machine,
## with the RTerm GUI
## JAGS version 4.3.1 found successfully using the command 'C:/Program
## Files/JAGS/JAGS-4.3.1/x64/bin/jags-terminal.exe'
## The rjags package is installed
```

# Results from simulation study

Table 1: Results from simulation studies

	Sp	Se V+	Se V-	True OR	Bias - BM Adjusted	Bias - Unadjusted	
Non-differential misclassification	0.99	0.925	0.925	0.1	1.01 (0.82-1.20)	1.32 (1.12-1.53)	
	0.99	0.925	0.925	0.2	1.01 (0.87-1.17)	1.16 (1.02-1.32)	
	0.99	0.975	0.975	0.1	1.01 (0.83-1.20)	1.29 (1.10-1.49)	
	0.99	0.975	0.975	0.2	1.00 (0.87-1.15)	1.13 (1.00-1.28)	
	1.00	0.925	0.925	0.1	1.01 (0.85-1.18)	1.03 (0.87-1.21)	
	1.00	0.925	0.925	0.2	1.00 (0.89-1.15)	1.03 (0.91-1.17)	
	1.00	0.975	0.975	0.1	1.01 (0.85-1.18)	1.01 (0.86-1.18)	
	1.00	0.975	0.975	0.2	1.00 (0.89-1.15)	1.01 (0.89-1.15)	
	Differential missclassification	0.99	0.925	0.975	0.1	1.01 (0.83-1.20)	1.23 (1.05-1.43)
		0.99	0.925	0.975	0.2	1.00 (0.88-1.16)	1.08 (0.96-1.23)
0.99		0.975	0.925	0.1	1.01 (0.82-1.21)	1.38 (1.17-1.59)	
0.99		0.975	0.925	0.2	1.00 (0.87-1.16)	1.21 (1.06-1.38)	
1.00		0.925	0.975	0.1	1.01 (0.86-1.19)	0.96 (0.82-1.13)	
1.00		0.925	0.975	0.2	1.00 (0.89-1.15)	0.96 (0.85-1.09)	
1.00		0.975	0.925	0.1	1.01 (0.85-1.17)	1.09 (0.92-1.26)	
1.00		0.975	0.925	0.2	1.00 (0.89-1.14)	1.09 (0.96-1.23)	

BM = Bayesian model; Se V- = sensitivity in unvaccinated; Se V+ = sensitivity in vaccinated; Sp = specificity; OR = Odds Ratio  
 Bias statistics are median (5th - 95th percentiles) of the 1000 simulations ran for each scenario

## Code and MCMC output of illustrative case

### Gibbs sampling set-up.

The MCMC sampling required 100 000 iterations with 50 000 burnin and thin interval 25.

```
inits1 <- list(".RNG.name" = "base::Mersenne-Twister", ".RNG.seed" = 100022)
inits2 <- list(".RNG.name" = "base::Mersenne-Twister", ".RNG.seed" = 300022)

n_thin <- 25
n_burnin <- 50000
n_samples <- 100000
```

### Data from Chung et al. (2021).

A TND study conducted in Ontario, Canada, investigated the effectiveness of mRNA Covid-19 vaccines (bnt162b2 and mrna-1273) against symptomatic SARS-CoV-2 infection. (Chung et al. 2021) The study used linked data from provincial SARS-CoV-2 laboratory testing, Covid-19 vaccination, and health administrative datasets.

In testing positive subjects (cases) 57 were vaccinated and 51 220 unvaccinated. In subjects testing negative (negative-controls) 3 817 were vaccinated and 251 541 unvaccinated

```
y <- matrix(
  c(51220, 251541, 57, 3817),
  nrow = 2,
  byrow = T,
  dimnames = list(c("V-", "V+"), c("T+", "T-"))
)
y
```

```
##      T+      T-
## V- 51220 251541
## V+   57   3817
```

```
N <- apply(y, 1, sum)
N
```

```
##      V-      V+
## 302761  3874
```

## Bayesian models written in JAGS

```
# Bayesian model assuming perfect classification -----
bm_1t_perf <- " model {
  for (i in 1:2) {
    # likelihood
    y[i,1] ~ dbin(pi[i], N[i])
    # priors for prevalence parameters
    pi[i] ~ dbeta(2,2)
  }
  # Computing OR/VE
  OR <- (pi[2]/(1-pi[2])) / (pi[1]/(1-pi[1]))
  VE <- (1-OR)*100
  #data# N, y
  #inits#
  #monitor# pi, OR, VE
}
"
```

```

# Bayesian model for non-differential misclassification -----
bm_1t_nondif <- " model {

  for (i in 1:2) {

# likelihood
y[i,1] ~ dbin(prob[i], N[i])
prob[i] <- pi[i]*Se + (1-pi[i])*(1-Sp)

# priors for prevalence parameters
pi[i] ~ dbeta(2,2)
}

# priors for Se and Sp
Se~dbeta(HPSe[1], HPSe[2])
Sp~dbeta(HPSp[1], HPSp[2])

# Computing OR/VE

OR <- (pi[2]/(1-pi[2])) / (pi[1]/(1-pi[1]))
VE <- (1-OR)*100

#data# N, y, HPSe, HPSp
#inits#
#monitor# Se, Sp, pi, OR, VE

}
"

```

## Model 1: perfect classification

```
res_perfect <- run.jags(  
  bm_1t_perf,  
  n.chains = 2,  
  inits = list(inits1, inits2),  
  burnin = n_burnin,  
  sample = n_samples,  
  thin = n_thin  
)
```

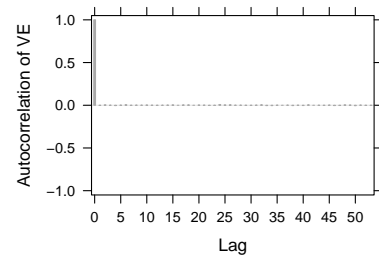
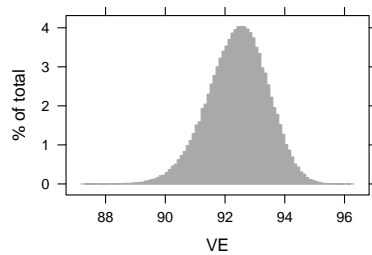
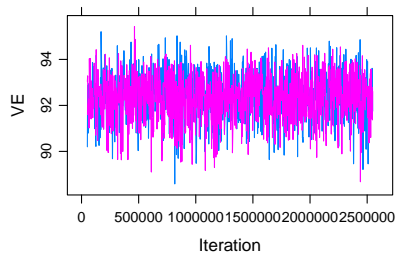
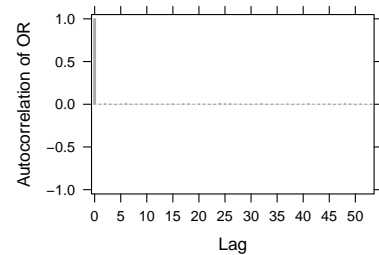
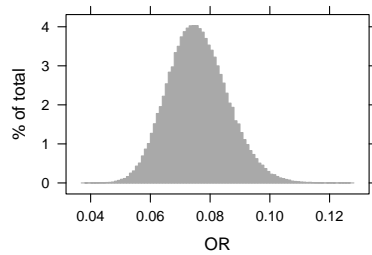
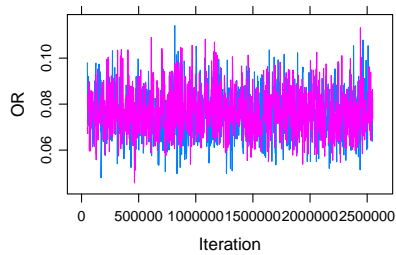
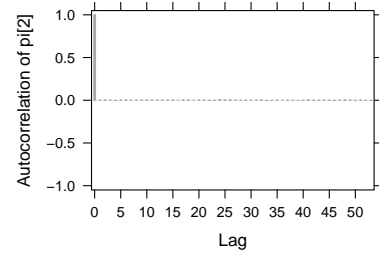
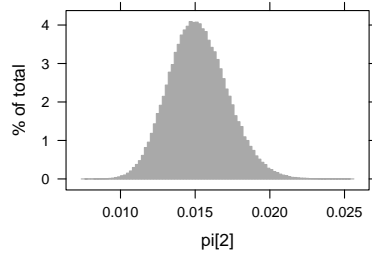
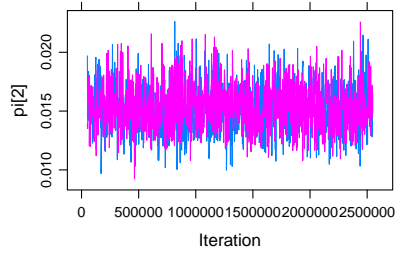
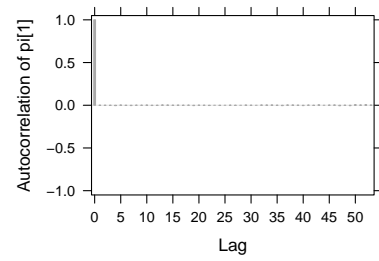
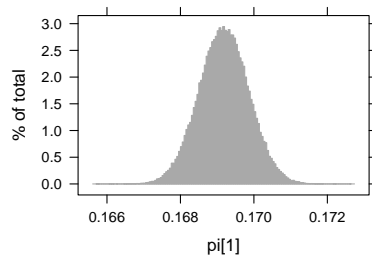
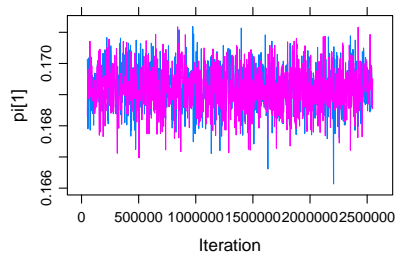
```
## Compiling rjags model...  
## Calling the simulation using the rjags method...  
## Adapting the model for 1000 iterations...  
## Burning in the model for 50000 iterations...  
## Running the model for 2500000 iterations...  
## Simulation complete  
## Calculating summary statistics...  
## Calculating the Gelman-Rubin statistic for 4 variables....  
## Finished running the simulation
```

```
round(summary(res_perfect), 3)
```

##	Lower95	Median	Upper95	Mean	SD	Mode	MCerr	MC%ofSD	SSeff	AC.250	psrf
## pi[1]	0.168	0.169	0.171	0.169	0.001	NA	0.000	0.7	20000	0.002	1
## pi[2]	0.012	0.015	0.019	0.015	0.002	NA	0.000	0.7	20405	0.002	1
## OR	0.057	0.075	0.096	0.076	0.010	NA	0.000	0.7	20382	0.002	1
## VE	90.409	92.460	94.269	92.404	0.996	NA	0.007	0.7	20382	0.002	1

```
plot(  
  res_perfect,  
  plot.type = c("histogram", "trace", "autocorr"),  
  vars = c("pi", "OR", "VE"),  
  layout = c(4, 3)  
)
```

```
## Generating plots...
```



## Model 2: sensitivity and specificity from Kostoulas, Eusebi, and Hartnack (2021)

Kostoulas and colleagues (Kostoulas, Eusebi, and Hartnack 2021) used a Bayesian latent class model to estimate the diagnostic accuracy of RT-PCR and lateral flow immunoassay tests for Covid-19. The sensitivity of RT-PCR was 0.68 (95% PrI=0.63-0.73), while the specificity was 0.99 (95% PrI=0.98-1.00).

We plugged-in this prior information in our model by using a Beta(226.16, 105.93) prior for the sensitivity and a Beta(287.48, 2.14) prior for the specificity.

```
HPSe <- findbetaqq2(  
  percentile.value1 = 0.63,  
  percentile1 = 0.025,  
  percentile.value2 = 0.73,  
  percentile2 = 0.975  
)  
HPSe
```

```
## [1] 226.16 105.93
```

```
round(qbeta(c(0.025, 0.5, 0.975), # check  
  HPSe[1], HPSe[2]), 2)
```

```
## [1] 0.63 0.68 0.73
```

```
HPSp <- findbeta2(  
  themedian = 0.99,  
  percentile = 0.975,  
  lower.v = FALSE,  
  percentile.value = 0.98  
)  
HPSp
```

```
## [1] 606.34 6.45
```

```
round(qbeta(c(0.025, 0.5, 0.975), # check  
  HPSp[1], HPSp[2]), 3)
```

```
## [1] 0.980 0.990 0.996
```

```
res_kostoulas <- run.jags(  
  bm_1t_nondif,  
  n.chains = 2,  
  inits = list(inits1, inits2),  
  burnin = n_burnin,  
  sample = n_samples,  
  thin = n_thin  
)
```

```
## Compiling rjags model...  
## Calling the simulation using the rjags method...  
## Adapting the model for 1000 iterations...
```



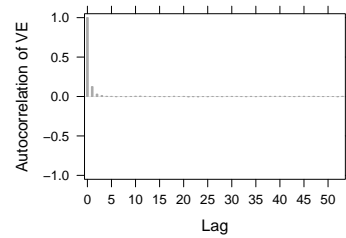
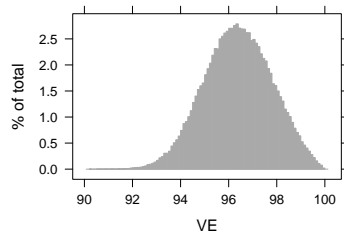
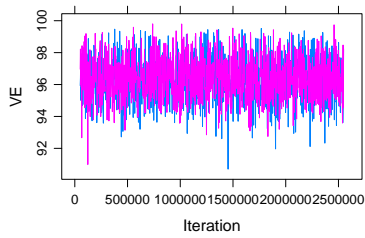
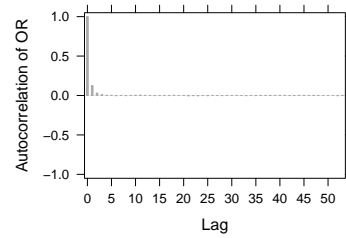
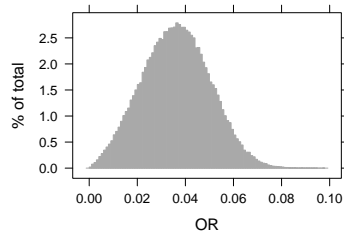
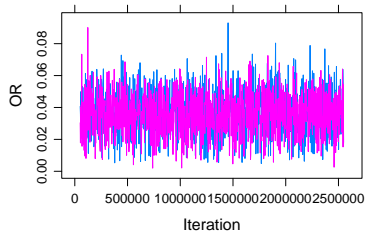
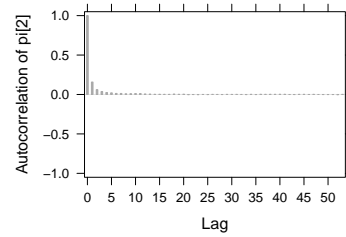
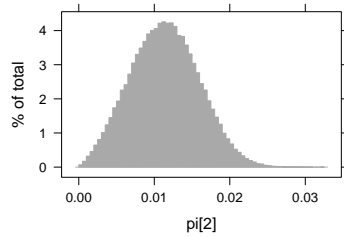
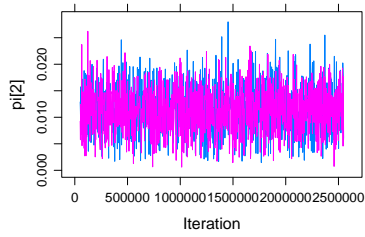
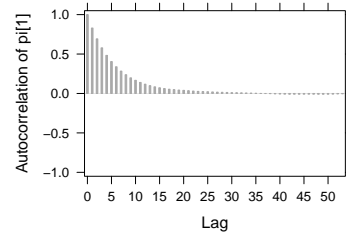
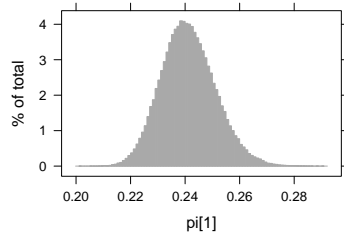
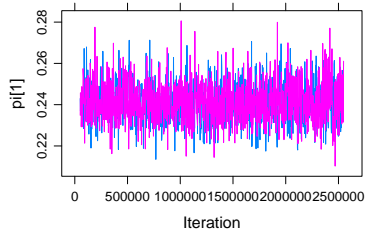
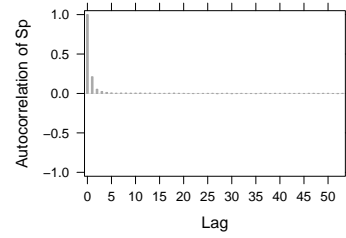
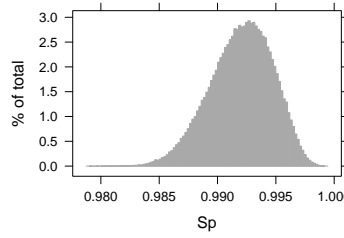
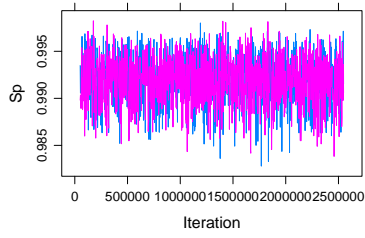
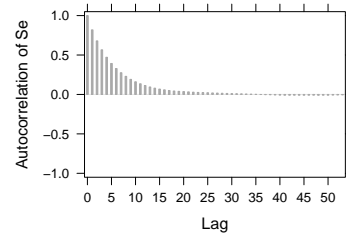
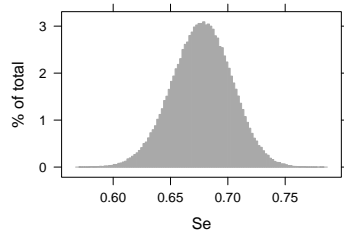
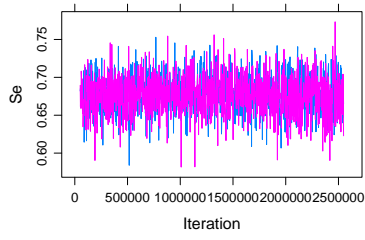
```
## Burning in the model for 50000 iterations...
## Running the model for 2500000 iterations...
## Simulation complete
## Calculating summary statistics...
## Calculating the Gelman-Rubin statistic for 6 variables....
## Note: Unable to calculate the multivariate psrf
## Finished running the simulation
```

```
round(summary(res_kostoulas), 3)
```

##	Lower95	Median	Upper95	Mean	SD	Mode	MCerr	MC%ofSD	SSEff	AC.250	psrf
## Se	0.626	0.678	0.728	0.677	0.026	NA	0.00	0.8	14469	0.160	1
## Sp	0.987	0.992	0.997	0.992	0.003	NA	0.00	0.7	20000	0.004	1
## pi[1]	0.223	0.241	0.262	0.241	0.010	NA	0.00	0.8	14725	0.167	1
## pi[2]	0.002	0.011	0.020	0.011	0.004	NA	0.00	0.7	20000	0.012	1
## OR	0.009	0.036	0.063	0.036	0.014	NA	0.00	0.7	20000	0.004	1
## VE	93.700	96.405	99.051	96.384	1.391	NA	0.01	0.7	20000	0.004	1

```
plot(
  res_kostoulas,
  plot.type = c("histogram", "trace", "autocorr"),
  vars = c("Se", "Sp", "pi", "OR", "VE"),
  layout = c(6, 3)
)
```

```
## Generating plots...
```



### Model 3: sensitivity and specificity from Stærk-Østergaard et al. (2021)

A recent report using Danish registries data used a Bayesian latent class model to estimate the diagnostic accuracy of RT-PCR and antigen tests for Covid-19.(Stærk-Østergaard et al. 2021) The specificity of RT-PCR was estimated to be close to 1.00. The sensitivity estimates were 0.957 (95% PrI=0.928-0.984). We plugged-in this prior information in our model by using a Beta(3040.61, 3.64) prior for the specificity and a Beta(168.66, 6.84) prior for the sensitivity.

```
# Sp
HPSp <- findbetaqq2(
  percentile.value1 = 0.9973,
  percentile1 = 0.025,
  percentile.value2 = 0.9997,
  percentile2 = 0.975
)
HPSp
```

```
## [1] 3040.61    3.64
```

```
round(qbeta(c(0.025, 0.5, 0.975),
            HPSp[1], HPSp[2]), 4)
```

```
## [1] 0.9973 0.9989 0.9997
```

```
# Se
HPSe <- findbetaqq2(
  percentile.value1 = 0.9279,
  percentile1 = 0.025,
  percentile.value2 = 0.9843,
  percentile2 = 0.975
)
HPSe
```

```
## [1] 168.66    6.84
```

```
round(qbeta(c(0.025, 0.5, 0.975),
            HPSe[1], HPSe[2]), 4)
```

```
## [1] 0.9279 0.9628 0.9843
```

```
res_staerk <- run.jags(
  bm_1t_nondif,
  n.chains = 2,
  inits = list(inits1, inits2),
  burnin = n_burnin,
  sample = n_samples,
  thin = n_thin
)
```

```
## Compiling rjags model...
```

```
## Calling the simulation using the rjags method...
```

```

## Adapting the model for 1000 iterations...
## Burning in the model for 50000 iterations...
## Running the model for 2500000 iterations...
## Simulation complete
## Calculating summary statistics...
## Calculating the Gelman-Rubin statistic for 6 variables....
## Note: Unable to calculate the multivariate psrf
## Finished running the simulation

```

```
round(summary(res_staerk), 3)
```

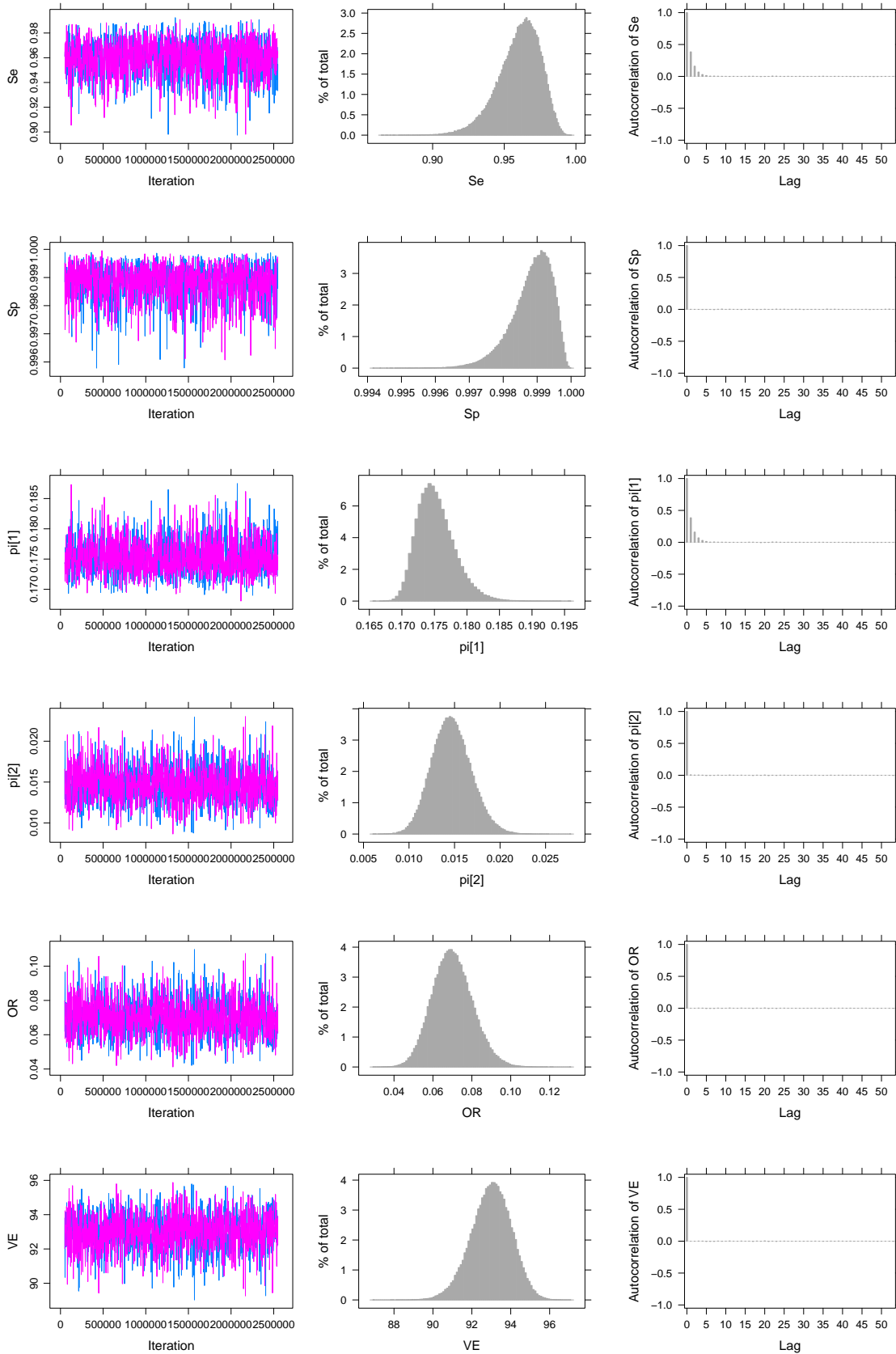
##	Lower95	Median	Upper95	Mean	SD	Mode	MCerr	MC%ofSD	SSEff	AC.250	psrf
## Se	0.931	0.962	0.987	0.960	0.015	NA	0.000	0.7	20000	0.000	1
## Sp	0.998	0.999	1.000	0.999	0.001	NA	0.000	0.7	20598	0.000	1
## pi[1]	0.170	0.175	0.181	0.175	0.003	NA	0.000	0.7	20000	0.002	1
## pi[2]	0.011	0.015	0.019	0.015	0.002	NA	0.000	0.7	20000	0.000	1
## OR	0.050	0.070	0.090	0.070	0.010	NA	0.000	0.7	20000	0.000	1
## VE	90.965	93.030	95.006	92.993	1.033	NA	0.007	0.7	20000	0.000	1

```

plot(
  res_staerk,
  plot.type = c("histogram", "trace", "autocorr"),
  vars = c("Se", "Sp", "pi", "OR", "VE"),
  layout = c(6, 3)
)

```

```
## Generating plots...
```



## Code availability

Code available at <https://github.com/paoloeusebi/tnd-vaccine-effectiveness/>

## References

- Chung, Hannah, Siyi He, Sharifa Nasreen, Maria E Sundaram, Sarah A Buchan, Sarah E Wilson, Branson Chen, et al. 2021. “Effectiveness of BNT162b2 and mRNA-1273 Covid-19 Vaccines Against Symptomatic SARS-CoV-2 Infection and Severe Covid-19 Outcomes in Ontario, Canada: Test Negative Design Study.” *BMJ* 374. <https://doi.org/10.1136/bmj.n1943>.
- Kostoulas, Polychronis, Paolo Eusebi, and Sonja Hartnack. 2021. “Diagnostic Accuracy Estimates for COVID-19 Real-Time Polymerase Chain Reaction and Lateral Flow Immunoassay Tests With Bayesian Latent-Class Models.” *American Journal of Epidemiology* 190 (8): 1689–95. <https://doi.org/10.1093/aje/kwab093>.
- Stærk-Østergaard, Jacob, Carsten Kirkeby, Lasse Engbo Christiansen, Michael Asger Andersen, Camilla Holten Møller, Marianne Voldstedlund, and Matthew J Denwood. 2021. “Evaluation of Diagnostic Test Procedures for SARS-CoV-2 Using Latent Class Models: Comparison of Antigen Test Kits and Sampling for PCR Testing Based on Danish National Data Registries.” *arXiv Preprint arXiv:2112.11298*.