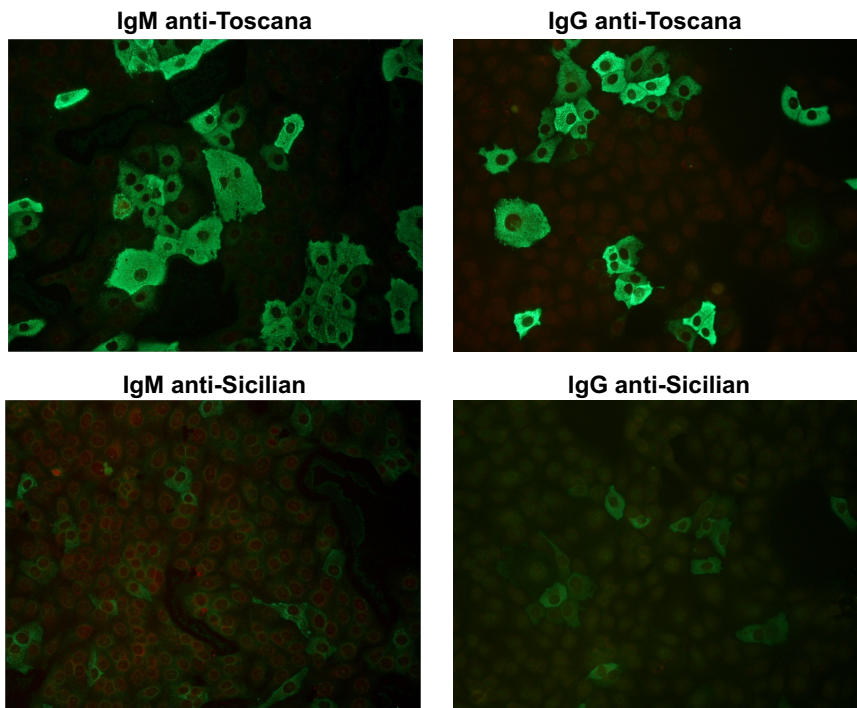


**Additional Figure 1: Toscana virus IgG and IgM Serology.**



Serology was performed with the Sandfly fever virus Mosaic 1 types Sicilian, Naples, Toscana, Cyprus IgG and IgM assay (EuroImmun, Luebeck, Germany).

## Additional Methods

### TOSV sequencing

Amplicons extending over a gap in sequence coverage in segment S were generated with opposite direction primer pairs (TOSV\_fwd\_Seq5, TOSV\_rev\_Seq7 and TOSV\_fwd\_Seq7, TOSV\_rev\_Seq5). PCR reactions were prepared including a no template control. 4  $\mu\text{L}$  Phusion HF Buffer (5X, final 1X), 0.5  $\mu\text{L}$  dNTP Mix (10 mM each, final 0.25 mM each), 0.75  $\mu\text{L}$  forward primer (10  $\mu\text{M}$ , final 0.375  $\mu\text{M}$ ), 0.75  $\mu\text{L}$  reverse primer (10  $\mu\text{M}$ , final 0.375  $\mu\text{M}$ ), 0.2  $\mu\text{L}$  Phusion Hot Start II High-Fidelity DNA Polymerase (2 U/ $\mu\text{L}$ , final 0.02 U/ $\mu\text{L}$ ), 12.8  $\mu\text{L}$  Water and 1  $\mu\text{L}$  template for a final volume of 20  $\mu\text{L}$  were mixed.

PCR cycling conditions were 98°C for 30 sec; 35 cycles of 98°C for 5 sec, 60°C for 10 sec, 72°C for 30 sec; and 72°C for 1 min.

PCR products were purified using the NucleoSpin Gel and PCR Clean-up Kit (MACHEREY-NAGEL). DNA concentration was measured using the QuantiFluor ONE dsDNA System (Promega) and Quantus Fluorometer (Promega). DNA fragment size was verified using the dsDNA Reagent Kit (Agilent) with the Fragment Analyzer (Agilent).

Equal amounts of DNA (0.5 ng each) from the previous step were pooled and used as template for the limited-cycle extension PCR. 4  $\mu\text{L}$  Phusion HF Buffer (5X, final 1X), 0.5  $\mu\text{L}$  dNTP Mix (10 mM each, final 0.25 mM each), 0.75  $\mu\text{L}$  primer TS-D501 (10  $\mu\text{M}$ , final 0.375  $\mu\text{M}$ ), 0.75  $\mu\text{L}$  primer TS-D701 (10  $\mu\text{M}$ , final 0.375  $\mu\text{M}$ ), 0.2  $\mu\text{L}$  Phusion Hot Start II High-Fidelity DNA Polymerase (2 U/ $\mu\text{L}$ , final 0.02 U/ $\mu\text{L}$ ), 12.5  $\mu\text{L}$  Water and 1.3  $\mu\text{L}$  template for a final volume of 20  $\mu\text{L}$  were mixed.

PCR cycling conditions were 98°C for 30 sec; 12 cycles of 98°C for 5 sec, 55°C for 30 sec, 72°C for 30 sec; and 72°C for 1 min.

PCR products were purified using the NucleoSpin Gel and PCR Clean-up Kit (MACHEREY-NAGEL). DNA concentration was measured using the QuantiFluor ONE dsDNA System (Promega) and Quantus Fluorometer (Promega). DNA fragment size was verified using the dsDNA Reagent Kit (Agilent) with the Fragment Analyzer (Agilent).

The indexed, purified PCR product was diluted to 4 nM, 5  $\mu\text{L}$  mixed with 5  $\mu\text{L}$  NaOH (0.2 M), incubated at RT for 5 min, 990  $\mu\text{L}$  HT1 added. The resulting 20 pM denatured library was diluted to 18 pM and sequenced on the MiSeq with the 150-cycle sequencing kit.

## PCR Primers

TOSV_903fwd_Seq5	CTTCCCTACACGACGCTCTTCCGATCTTGGCCCACAATACATGAGT
TOSV_1107rev_Seq7	GGAGTTCAGACGTGTGCTCTTCCGATCTTTTGGCATCAGCAGTAGT GAGA
TOSV_903fwd_Seq7	GGAGTTCAGACGTGTGCTCTTCCGATCTTGGCCCACAATACATGAGT
TOSV_1107rev_Seq5	CTTCCCTACACGACGCTCTTCCGATCTTTTGGCATCAGCAGTAGT GAGA
TS-D501	AATGATACGGCGACCACCGAGATCTACACTATAGCCTACACTCTTT CCCTACACGA CGCTCTTCCGATCT
TS-D701	CAAGCAGAAGACGGCATAACGAGATCGAGTAATGTGACTGGAGTTC AGACGTGTGCTCTTCCGATC