Defenses		Billoth and (D)	Creation	Deer	Timespan		
Reference	Method (1)	Method (2)	Species	Dose	dose to feeding	Most relevant results	Additional observations
Pampiglione 1985 [1]	treated subjects	mice	An. stephensi	140 to 28,000 μg/kg (once, subcutaneous)	12 hours	3-day mortality: 60% in the 280 μg/kg group Controls 20% (significance not reported)	
lakubovich 1989 [2]	membrane	in vitro mixture (blood + ivermectin)	An. stephensi An. sacharovi	1 to 50 ppm	N/A	[time of mortality assessment not stated] An. Stephensi LC100 1ppm An. sacharovi LC100 50 ppm	
lakubovich 1989 [2]	treated subjects	rabbits	An. stephensi An. Sacharovi An. atroparvus	340 μg/Kg (once, subcutaneous)	not available	 An. stephensi 4-day-mortality: 93% (significant) An. sacharovi No difference with control An. atroparvus No difference with control 	
Jones 1992 [3]	membrane	blood from treated dogs	An. quadrimaculatus	10 to 2,500 μg/Kg (once, orally)	4 hours	2-day mortality: 92% in the 500 μg/kg group Controls 3.4% (significant)	
Jones 1992 [3]	treated subjects	dogs	An. quadrimaculatus	10 to 2,500 μg/Kg (once, orally)	4 hours	2-day mortality: 98.6% in the 10 μg/kg group Controls 4.3%	
Gardner 1993 [4]	treated subjects	dogs	An. quadrimaculatus	6 to 24 μg/Kg (once, orally)	4 hours	24-hour mortality: 66.9% in the 12 µg/kg group Controls: 3.9% (significant) 24-hour LC50: between 6 and 12 ng/ml	24-hour Lethal DOSE 50: 9.9 μg/kg Significant decrease in oviposition and egg- hatching from survivors
Bockarie 1999 [5]	field collections	MDA for LF	An. punctulatus An. koliensis	400 μg/Kg (once, orally) + 6 mg/kg DEC	≤ 4 days	3-day mortality: 61% if collected ≤4 days post treatment Controls: 1-10% (significant) 9-day mortality: 100% if collected ≤4 days post treatment Controls: 18-23% (significant)	70% of deaths within 24 hours of collection. Survival rates similar for both species
Foley 2000 [6]	treated subjects	human volunteer (1)	Anopheles farauti	250 μg/Kg (once, orally)	Same day to 44 days	3-day mortality: 100% if fed on the same day Controls: 4% (significant) 3-day mortality: 80% if fed 7 days post treatment 9-day mortality: 100% if fed 7 days post treatment Controls: 4-6% (significant)	
Fritz 2009 [7]	membrane	in vitro mixture (blood + ivermectin)	An gambiae An arabiensis	0.01-1000 ppb	N/A	Survival rates similar for both species [time of mortality assessment not stated]-LC50: 19.8 ppb [time of mortality assessment not stated]-LC95: 77.7 ppb	
Fritz 2009 [7]	treated subjects	cattle	An gambiae An arabiensis	600 μg/kg (once, subcutaneous)	1 to 23 days	3-day mortality: 100% if fed 1 day post treatment Controls: 10% (significance not reported) 3-day mortality: 62% if fed 13 days post treatment 9-day mortality: 88% if fed 13 days post treatment Controls: 10-38% (significance not reported)	No oviposition or significantly reduced oviposition from mosquitoes fed up to 17 days post-treatment
Chaccour 2010 [8]	treated subjects	human volunteers (25)	An. gambiae	200 μg/kg (once, orally)	1 and 14 days	3-day mortality: 84% if fed 1 day post treatment Controls: 38% (significant) 9-day mortality: 96% if fed 1 day post treatment Controls: 73% (significant) No difference in mortality when fed 14 days after treatment.	A non-measured proportion of mosquitoes in the ivermectin group were noted to show lack of movement coordination, lethargy, inability to fly, and paralysis.

Reference	method (1)	method (2)	species	Dose	Timespan dose to feeding	Most relevant results	Additional observations
Kobylinski 2010 [9]	membrane	in vitro mixture (blood + ivermectin)	An. gambiae	0.5 to 64 ng/ml	N/A	3-day mortality: 75% in the 32 ng/ml group Controls 10-20% (significance not reported) 3-day mortality: 90% in the 64 ng/ml group Controls 10-20% (significance not reported) [5 day]-LC50: 22.4 ng/ml (18-26.9)	Sublethal concentrations significantly reduced mosquito re-blood feeding rates. Multiple sublethal blood meals further increased mortality. Adult size of An. gambiae did not alter susceptibility to ivermectin
Sylla 2010 [10]	field collections	MDA for onchocerciasis	An. gambiae An. arabiensis	150 µg/Kg (once, orally)	3 groups: pre-, 1 to 6 days and ≥ 7 days post MDA	An. gambiae 5-day mortality: 70% if collected 2 days post MDA Controls: 16-22% (significant) No difference with controls if collected after day 6. An. arabiensis 5-day mortality: 50% if collected 1-6 days post MDA Controls: <20% (significant)	
Kobylinski 2011 [11]	field collections	MDA for onchocerciasis	An. gambiae	150 µg/Kg (once, orally)	1-12 days post MDA		Sporozoite rate reduced by 79% in mosquitoes collected 2 weeks following MDA in treated villages vs sporozoite rate increased by 246% in control villages (significant)
Butters 2012 [12]	membrane	in vitro mixture (blood + ivermectin)	An. gambiae	5 different concentration (not stated)	N/A		Sublethal concentrations induced significant knockdown and inhibited recovery. No effect on the re-blood feeding rate.
Fritz 2012 [13]	membrane	in vitro mixture (blood + ivermectin)	An. arabiensis	0.1 to 100 ppb	N/A	9-day mortality: 70% in the 10 ppb group 9-day mortality: 100% in the 100 ppb group Controls 10% (significance not reported) [9-days]-LC50: 7.9 ppb (6.2 - 9.9)	LC95: 128.1 ppb (62.1 - 264.4)
Kobylinski 2012 [14]	membrane	in vitro mixture (blood + ivermectin)	An. gambiae	not stated	N/A	7-day-LC50 = 15.9 ng/ml (14.6, 17.3)	Sublethal concentrations significantly inhibited <i>P. falciparum</i> sporogony.
Bastiaens 2012 [15]	treated subjects	Swiss mice, Wistar rats and Cynomolgus monkeys	An. stephensi	mice and rats: 400 μg/Kg, Monkeys: 200 and 400 μg/Kg	1-5 days	Similar results for all three species. 3-day mortality: 70-100% if fed 1-2 days post treatment Controls: 0-28% (significant)	
Naz 2013 [16]	field collections	cattle	An. culicifacies, An. stephensi	200 μg/Kg (once, subcutaneous)	1 to 28 days post dose	An. culicifacies3-day mortality: 65% if fed 1 day post treatment Controls: 9%9-day mortality: 80% if fed 1 day post treatment Controls: 17% An. stephensi3-day mortality: 80% if fed 1 day post treatment Controls: 10%9-day mortality: 80% if fed 1 day post treatment Controls: 25%	Significance only reported for the 12-day cumulative mortality

Reference	method (1)	method (2)	species	Dose	Timespan dose to feeding	Most relevant results	Additional observations
Yamada 2013 [17]	membrane	in vitro mixture (blood + ivermectin)	An. arabiensis	0.5 to 7.5 ppm (blood meal repeated daily)	N/A	24-hour mortality: 90% at 7.5 ppm 2-day mortality: 98% at 7.5 ppm 3-day mortality: 100% at 7.5 ppm	Ivermectin knocked down females almost immediately after blood feeding and killed most within 12 hours
Alout 2014 [18]	field collections	MDA for onchocerciasis or LF	An. gambiae	150 μg/Kg (once, orally) +/- albendazole	Pre- and 1- 12 days post MDA	3-day mortality: 65% if collected 2 days post MDA 5-day mortality: 68% if collected 2 days post MDA Controls: 16-22% (significant) Survivorship reduced by 33.9% for one week post MDA	Sporozoite rates significantly reduced by >77% for two weeks following the MDAs Parity rates were significantly reduced for more than two weeks after the MDAs
Kobylinski 2014 [19]	membrane	in vitro mixture (blood + ivermectin)	An. dirus An. minimus An. campestris An. sawadwonrporni	-	N/A	An. dirus [7 days]-LC50 = 55.6 ng/ml An. minimus [7 days]-LC50 = 16.3 ng/ml An. campestris [7 days]-LC50 = 26.4 ng/ml An. sawadwongporni [7 days]-LC50 =27.1 ng/ml	Preliminary data suggests that ivermectin is sporontocidal to <i>Plasmodium vivax</i> in <i>An.dirus</i>
Ouedraogo 2015 [20]	membrane	blood from treated volunteers	An. gambiae, An. funestus	200 µg/Kg (once or twice, orally)	1-7 days	An. gambiae 3-day mortality: 33% if fed <u>1 day</u> post treatment (<u>1 dose</u>) 3-day mortality: 31% if fed <u>3 days</u> post treat. (<u>2 doses</u>) Controls: 6% (significant) 10-day-mortality: 59% if fed <u>1 day</u> post treatment (1 dose) 10-day-motality: 66% if fed <u>3 days</u> post treat. (2 doses) Controls: 21% (significant) 10-days-LC50: 5.97 ng/ml An. funestus 3-day mortality: 19% if fed <u>1 day</u> post treatment (<u>1 dose</u>) 3-day mortality: 22% if fed <u>3 days</u> post treat. (<u>2 doses</u>) Controls: 3% (significant) 10-day-mortality: 40% if fed <u>1 day</u> post treatment (1 dose) 10-day-mortality: 51% if fed <u>3 days</u> post treat. (2 doses) Controls: 5% (significant)	The artemether-lumefantrine-ivermectin combination was well-tolerated 4- to 7-fold increased mortality in mosquitoes feeding 1 day after ivermectin Day 7 ivermectin plasma levels were positively associated with body mass index and female gender
Derua [21]	treated subjects	human volunteers	An. gambiae	150–200 µg/kg	24 hours	3-day mortality: 66.2% Controls 4% (significant) 9-day mortality: 95% Controls 12% (significant)	None of the An. gambiae in the ivermectin group laid eggs
Kobylinski 2015 [22]	membrane	in vitro mixture (blood + ivermectin)	An. dirus	not stated	N/A		Sublethal concentrations significantly inhibited <i>P. vivax</i> sporogony.
Poché 2015 [23]	treated subjects	Cattle	An. Coluzzi	100 & 200 µg/kg	1-21 days	 100 μg/kg 3-day mortality 45-63% if fed < 7 days post treatment 9-day mortality 65-94% if fed < 7 days post treatment (significant differences only one day post-treatment) 200 μg/kg 3-day mortality 53-77% if fed < 7 days post treatment 9-day mortality 85-100% if fed < 7 days post treatment (significant differences until 7 days post-treatment) 	Anopheles coluzzii carrying the kdr mutation

Reference	method (1)	method (2)	species	Dose	Timespan dose to feeding	Most relevant results	Additional observations
Seaman 2015 [24]	membrane	in vitro mixture (blood + ivermectin)	An. gambiae	11.75 ng/ml	N/A		Mosquito ivermectin susceptibility increased with age and previous bloodfeeding. Likely midgut interactions resulting from ivermectin ingestion: blood meal digestion physiological responses, midgut microflora, and innate immune responses. Gene transcription consistently affected by ivermectin ingestion.
Derua 2016 [25]	field collections	Larvae	An. gambiae	0.001-10 ppm	N/A	24-hour mortality: 38.4% with 0.1 ppm 24-hour mortality: 100% with 1 and 10 ppm	Cx. quinquefasciatus larvae approximately 10- fold more susceptible than <i>An. gambiae</i>

Studies assessing the efficacy of ivermectin to kill mosquitoes taking a loaded blood meal. Only studies based on blood meals are included, i.e. no impregnated cotton or sugary solutions. Studies using more than one feeding methodology were separated by experiment. Efforts were made to present the results in a uniform manner. 3-day and 9-day cumulative mortality were chosen as main outcomes given the duration of the gonotrophic and sporogonic cycles. If these were not available, the authors were contacted, and calculated when raw data was provided. If no response was obtained or if the data was not available, the 3-day and 9-day cumulative mortality were extrapolated from the Kaplan-Meier curves in every publication. If this was not possible, the cumulative mortality is reported together with the original time of assessment. The LC50 is reported with the time used for its assessment. Four studies do not use mortality as a primary endpoint (Kobylinski 2011 & 2015, Butters 2012 and Seaman 2015), but are included because of the importance of their results (sporozoite rate, knockdown/recovery and effect of senescence). One study on *Anopheles/Culex* larvae is included for potential implications of residual ivermectin in latrines.

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