Supplementary information

AP4 suppresses DNA damage, chromosomal instability and senescence via inducing *MDC1/Mediator of DNA damage Checkpoint 1* and repressing *MIR22HG/*miR-22-3p

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Fig. S1: *AP4* inactivation induces senescence and apoptosis in CRC cells. A AP4 detection by Western blot analysis. β -actin served as a loading control. **B** Phase contrast images of the indicated cell lines. Scale bars: 50 µm. **C** Colony formation assays of *AP4* WT and KO single cell clones obtained from DLD-1 and SW480 cells. Three independent clones were analyzed. **D** Detection of senescent cells using pH6 β -gal staining after 48 hours of transfection. Quantification of a total of 120 cells in 3 fields. Scale bars: 100 µm. **E** Quantification of apoptotic cells by Annexin V / PI detection 48 hours after transfection in the indicated cells (left panel). The flow charts of DLD-1 and SW480 cells with indicated transfections are presented in the right panel.

F MTT assay results of indicated cell lines 72 hours after seeding into a 96-well plate. **G** Proliferation was determined by impedance measurement. In panels **D-G** the mean <u>+</u> SD (n = 3) is provided. *: p < 0.05, **: p < 0.01, ***: p < 0.001, ****: p < 0.0001.



Figure S2

Fig. S2: AP4 inactivation induces DNA damage in CRC cells. A Detection of γ H2AX foci. Quantification of a total of 100 cells in 5 fields. Scale bars: 20 µm. B Western blot analysis of γ H2AX 48 hours after transfection in the indicated cells. C Comet assay and quantification of DNA tail moment of a total of 150 cells in 10 fields. Scale bars: 20 µm. D Western blot analysis 48 hours after transfection of c-*MYC*–specific siRNA. E FACS analysis of DLD-1/SW480 *AP4* WT/KO pRTR-c-*MYC*-VSV pools. Pools with more than 80% GFP-positive cells were selected. F γ H2AX detection after c-*MYC* induction by DOX for indicated durations. Quantification of a total of 100 cells in 5 fields. Scale bars: 20 µm. In panels A the mean \pm SD (n = 5), C the mean \pm SD (n = 10) and F the mean \pm SD (n = 5) are provided. *: p < 0.05, **: p < 0.01, ***: p < 0.001.



Fig. S3: AP4 directly represses *MIR22HG***. A** qPCR analysis of *MIR22HG* and *p21* in the indicated clones of SW480 cells. **B** qPCR analysis of miR-22-3p expression (left panel) and nascent *MIR22HG* mRNA (right panel) in the indicated cells. **C** Western blot analysis of MYC-VSV and AP4 expression after c-*MYC* induction by DOX for indicated durations. **D** qPCR analysis of *MIR22HG* (left panel) and miR-22-3p (right panel) after c-MYC induction by DOX for indicated durations by DOX for indicated durations. **E** qPCR analysis of nascent *MIR22HG* mRNA 48 hours after c-*MYC* induction by DOX. **F** qPCR analysis of *SNAI1* (left panel) and *p21* (right panel) after c-*MYC* induction by DOX for indicated durations. **G** qPCR analysis of *MIR22HG* 48 hours after transfection. **H** β-gal detection at pH 6 48 hours after transfection. Quantification of a total of 120 cells in 3 fields. Scale bars: 50 μm. In panels **A-B**, **D-H** the mean <u>+</u> SD (n = 3) is provided with *: p < 0.05, **: p < 0.01, ***: p < 0.001.





Fig. S4: *MDC1* is directly and indirectly regulated by AP4. A qPCR analysis of *MDC1* expression. **B** Western blot analysis. **C** qPCR analysis 48 hours after transfection. **D** Western blot analysis 48 hours after transfection. **E** MDC1 foci detection in untreated cells. Quantification of a total of 120 cells in 3 fields. Scale bars: 20 µm. **F** qPCR analysis of nascent *MDC1* mRNA in the indicated cell lines. **G** qPCR analysis of *MDC1* 48 hours after transfections. **H** Western blot analysis 48 hours after transfection. **I** qPCR analysis of nascent *MDC1* mRNA 48 hours after c-MYC induction by DOX. **J** Western blot analysis after induction of c-MYC by DOX for the indicated periods. In panels **A**, **C**, **E**-**G** and **I** the mean <u>+</u> SD (n = 3) is provided with *: p < 0.05, **: p < 0.01, ***: p < 0.001.



Fig. S5: MDC1 mediates effects of AP4 on DNA damage and senescence. A Detection of MDC1 and γ H2AX foci by immunocytochemistry 48 hours after silencing *MDC1*. Scale bars: 20 µm. Quantification of a total of 120 cells in 3 fields. **B** MDC1 and γ H2AX foci were detected by immunocytochemistry 48 hours after ectopic expression of MDC1. Scale bars: 20 µm. Quantification of a total of 120 cells in 3 fields. **C** β -gal staining 48 hours after silencing or ectopic expression of MDC1, respectively.

Quantification of a total of 120 cells in 3 fields. Scale bars: 50 µm. **D** MTT assay results 48 hours after silencing *MDC1*. **E** MTT assay results 48 hours after ectopic expressing MDC1. Detection of **F** γ H2AX foci by immunocytochemistry, **G** β -gal staining and **H** comet assay in DLD-1 *AP4* WT1 cells 48 hours after transfection. Quantification of DNA tail moment of a total of 150 cells in 10 fields. In panel **A-E**, the mean <u>+</u> SD (n = 3), in panel **F-G**, the mean <u>+</u> SD (n = 5) and in panel **H**, the mean <u>+</u> SD (n = 10) are provided with **: p < 0.01, ***: p < 0.001, ****: p < 0.0001.



Figure S6

Fig. S6: MDC1 mediates effects of AP4 on chromosomal instability and HR. A Examples and quantification of micronuclei after DAPI staining. Quantification of a total of 50 cells in 3 fields. Scale bars: 20 μ m. B Kinetic evaluation of micronucleus formation 48 hours after transfection with the indicated oligonucleotides or vectors. C The indicated cells were co-transfected with pDR-GFP, pCBAScel plasmids. A pcDNA-mCherry plasmid was also co-transfected as a control of transfection efficiency. The percentage of cells expressing GFP among mCherry-positive cells was measured by flow cytometry 72 hours after transfection. D-G The percentage of the indicated cells expressing GFP was measured by flow cytometry after 72 hours of transfections with the indicated plasmids or oligonucleotides. In panels A-G, the mean <u>+</u> SD (n = 3) is provided with *: p < 0.05, **: p < 0.01, ***: p < 0.001.



Figure S7

Fig. **S7**: AP4 confers resistance towards Etoposide via MDC1. Α Immunocytochemistry detection of MDC1 and yH2AX foci 12 hours after addition of 20 µM Etoposide. Quantification of a total of 120 cells in 3 fields. Scale bars: 50 µm. B MTT assay results 12 hours after 20 µM Etoposide treatment. MTT assay results of cells transfected with C the indicated oligonucleotides or D expression plasmids for 48 hours and then subjected to treatment with Etoposide for another 12 hours. In panels **A-D** the mean + SD (n = 3) is provided with *: p < 0.05, **: p < 0.01, ***: p < 0.001.



Figure S8

Fig. S8: AP4 confers resistance towards 5-FU via MDC1. A The indicated cells were treated with increasing concentrations of 5-FU for 48 hours. Then the IC_{50} was determined by an MTT assay. B-E The indicated cells were transfected with indicated oligonucleotides for 48 hours and subsequently treated with increasing concentrations

of 5-FU for another 48 hours. After treatments, the IC₅₀ was determined by an MTT assay. **F-I** Colony formation assay of the indicated cells transfected with the indicated oligonucleotides or plasmids for 48 hours and then subjected to treatment with 10 μ M 5-FU for another 48 hours. For the last 48 hours fresh DOX was added. After treatments, cells were cultured for 3 weeks. In panels **A-I**, the mean <u>+</u> SD (n = 3) is provided with *: p < 0.05, **: p < 0.01, ***: p < 0.001.



Fig. S9: Kaplan–Meier analysis of the association between relapse free survival and c-MYC, AP4, MIR22HG and MDC1 mRNA expression. Analysis of A c-MYC, B AP4, C MIR22HG and D MDC1 in patients that received chemotherapy (n = 87) represented in the GSE14333 dataset. The significance was calculated with the log-rank test. Below the graphs, the number of patients at risk with high or low expression of the indicated mRNA at the respective time point is provided. HR, hazard ratio.

Fig. S10: Original blots.











uncropped gel of Fig 4F



uncropped gel of Fig 4H and S4J





uncropped gel of Fig 9F



Uncropped gel for Figure S3C



Table S1: Sequence information for miR-22-3p mimic and control mimic.

	Sequence information (5'-3')
hsa-miR-22-3p mimic	AAGCUGCCAGUUGAAGAACUGU
Control mimic	UCACCGGGUGUAAAUCAGCUUG

Table S2: Sequence information for guide RNAs used for *AP4* deletion.

	Sequence information (5'-3')	
Guide RNA 1 Forward	CACCGACCAGGAGCGGCGGATTCGG	
Guide RNA 1 Reverse	AAACCCGAATCCGCCGCTCCTGGTC	
Guide RNA 2 Forward	CACCGGCGTCTCCGCTCGTTGCTGT	
Guide RNA 2 Reverse	AAACACAGCAACGAGCGGAGACGCC	
Guide RNA 3 Forward	CACCGCGCATGCAGAGCATCAACGC	
Guide RNA 3 Reverse	AAACGCGTTGATGCTCTGCATGCGC	

Table S3: List of Antibodies.

Epitope	Species	Catalog No.	Company	Use	Dilution	Source
Primary antibodies						
α-tubulin β-actin VSV	Human Human Human	# T-9026 # A2066 # V4888	Sigma-Aldrich Sigma-Aldrich Sigma-Aldrich	WB WB WB	1:1000 1:1000 1:7500	mouse rabbit rabbit
TFAP4	Human	# MCA4993Z	AbD Serotec	WB; ChIP	1:1000; 3 ua	mouse
MDC-1 Mdc-1 γ-H2AX	Human Mouse Human	# NB100-397 # ab217528 # JBW301	NOVUS Abcam Sigma-Aldrich	WB/IF IHC IF	1:1000 1:400 1:1000	rabbit rabbit mouse
γ-H2ax	Mouse	# 9718	Cell Signaling Technology	IHC	1:500	rabbit
p16 (M-156) Ki67	Mouse Mouse	# sc-1207 # E0468	Santa Cruz Dako	IHC IHC	1:100 1:500	rabbit rat Alexa Fluor®
F-actin	N.A.	# A12379	Thermo Fisher	IF	1:50	488 conjuga ted
Secondary antibodies						
Anti-mouse HRP	N.A.	# W4021	Promega	WB	1:10,000	goat
Anti-rabbit HRP	N.A.	# A0545	Sigma-Aldrich	WB	1:10,000	goat
Alexa Fluor Plus 555	N.A.	# A32727	Thermo Fisher	IF	1:1000	goat
Alexa Fluor Plus 488	N.A.	# A32731	Thermo Fisher	IF	1:1000	goat

Table S4: Oligonucleotides used for qPCR.

mRNA	Forward (5'-3')	Reverse (5'-3')
GAPDH	TGTTGCCATCAATGACCCCTT	CTCCACGACGTACTCAGCG
MDC1	TGCTCTTCACAGGAGTGGTG	GGGCACACAGGAACTTGACT
AP4	GCAGGCAATCCAGCACAT	GGAGGCGGTGTCAGAGGT
c-MYC	CTTCTCTCCGTCCTCGGATTCT	GAAGGTGATCCAGACTCTGACCTT
β-actin	TGACATTAAGGAGAAGCTGTGCTAC	GAGTTGAAGGTAGTTTCGTGGATG
MIR22HG	TTTGCAATAGGGGATTGCTT	TTTAATGTCTGCGCGGTACTC
p21	GGCGGCAGACCAGCATGACAGATT	GCAGGGGGCGGCCAGGGTAT
SNAI1	GCACATCCGAAGCCACAC	GGAGAAGGTCCGAGCACAC
mmu <i>-Ap4</i>	TCAAGCGCTTTATCCAGGAG	CAATGCCCTCATCCTTGTCT
mmu-Mdc1	CCACAAGAGCCAGGACCTTC	TGTAGCCAAGACTTCCCAAGG

Table S5: Oligonucleotides used for qChIP.

Gene	Forward (5'-3')	Reverse (5'-3')
<i>MIR22HG</i> (site A)	AGGGGGAGCAAATCACTGCG	CCGTGCATTCGCAGCTCGTG
<i>MIR22HG</i> (site B)	TGATGAGGCTGGAGGGTG	GGAGGGTAAGCAAGGAGGA
<i>MIR22HG</i> (site C)	AGGTCGGAGGTTGAGGAA	GTTGAGGCAGGCTGGAAG
MDC1 (site A and B)	GACAACCCACTACCGCTTGC	AAAGGCGCTCTGGCCTTACC
MDC1 (site C)	GAGATGACTTGTGGAATAGGAGGTAG	CCTTCCGGGACCTACCTCAG
SNAI1 16q22	GGAGTACTTAAGGGAGTTGGCGG CTACTCACTTATCCATCCAGGCTAC	GAACCACTCGCTAGGCCGT ATTTCACACACTCAGACATCACAG

Table S6: Oligonucleotides used for reporter plasmids.

Oligo	Forward (5'-3')	Reverse (5'-3')
	AAACTAGTGAACTCCACTACCCTTTTC	GGCTGCAGTAAGGCACAGAGTGA
MDC13-UTR	CCTC	ΑΤΑΤΤΤΑΤΤΤΑΤΟΑ
miR-22-3p	TCATGCTCAGATGTCATAAGATCTTTA	GCAACAGTCTGGCTAAAGATCTTA
ΔSMS	GCCAGACTGTTGC	TGACATCTGAGCATGA
miR-22-3p	AATTCACAGTTCTTCAACTGGCAGCTT	GTGTCAAGAAGTTGACCGTCGAA
antisense	CTGCA	G