Supplemental Information

Stathmin 1/2-triggered microtubule loss mediates Golgi fragmentation in mutant SOD1 motor neurons

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Supplemental Table S1. Primary and secondary antibodies

			Catalog
Primary Antibodies	Dilution IF (WB)	Supplier	n°/reference
mouse anti-β-actin	1:5000	Sigma	A1978
goat anti-Choline Acetyl			
Transferase (ChAT)	1:200	Chemicon	Ab144P
mouse anti-Clathrin heavy			
chain	1:1000	Becton Dickinson	610499
mouse anti-β-COP	(1:1000)	Abcam	Ab6323
rabbit anti-β-COP	(1:1000)	Dr. R. Duden	Duden et al., 1991
mouse anti-GM130	1:300 (1:500)	Becton Dickinson	610823
mouse anti-GS28	1:300 (1:1000)	Becton Dickinson	611185
mouse anti-GS15	1:300 (1:1000)	Becton Dickinson	610961
rabbit anti-L1	(1:2000	Dr. M. Schäfer	Schäfer et al. 2010
rabbit anti-Myc	1 : 400 (1:1000	Cell Signaling	2272
mouse anti-p115	(1:1000)	Becton Dickinson	612260
rabbit anti-Cu/Zn hSOD1	1:1000	Enzo	ADI-SOD-100
rabbit anti-stathmin 1	1:1000 (1:1000)	Dr. A. Sobel	Gavet et al., 1998
rabbit anti-stathmin 2	1:1000 (1:1000)	Dr. A. Sobel	Gavet et al., 1998
rabbit anti-stathmin 3	(1:1000)	Dr. A. Sobel	Gavet et al., 1998
rabbit anti-Sec23	1:1000	Abcam	ab50672
mouse anti-syntaxin 5a	1:10000	Abcam	ab96185
mouse anti-acetylated tubulin	(1:2000)	Sigma	T6793
mouse anti-βııı-tubulin	1:10000	Babco	TUJI (MMS-435P)
mouse anti- α -tubulin	1:2000 (1:5000)	Sigma	T9026
rabbit anti-detyr-tubulin	1:2000	Dr. A. Andrieux	Erck et al., 2005
rat anti-Tyr-tubulin	1:2000 (1:50000)	Dr. A. Andrieux	Erck et al., 2005
rat anti-Tyr-tubulin	1:5000	Millipore	YL1/2 (Mab1864)
rabbit anti-VAChT	1:2000	Sigma	V5387
mouse anti-Vti1a	1:1000 (1:1000)	Becton Dickinson	V85620

Secondary antibodies	Dilution	Supplier
goat anti-mouse IgG Alexa 488	1:500	Molecular Probes
goat anti-mouse IgG Cy3	1:2000	Jackson Lab.
goat anti-mouse IgG Alexa 633	1:2000	Molecular Probes
goat anti-rabbit IgG Alexa 488	1:500	Molecular Probes
goat anti-rabbit IgG Cy3	1:2000	Jackson Lab.
goat anti-rabbit IgG Alexa 633	1:2000	Molecular Probes
goat anti-rat IgG Alexa Cy3	1:2000	Molecular Probes



Golgi structure analyzed by GM130 immunolabeling.

A. Single confocal sections at the midplane of the nucleus show apparent reduction of GM130-labelled Golgi area in motor neurons from mice of the indicated genotypes (age 240 days). Scale bar 10 μ m.

B. Single confocal sections of lumbar spinal cord from non-transgenic control mice (age 130 days) and transgenic SOD1^{wt} mice (age 180 days) as well as presymptomatic mutant SOD1^{G85R} mice aged 180 days and SOD1^{G93A} mice aged 130 days are identified with the motor neuron marker VAChT. Golgi structure is analyzed with the marker GM130. Black and white images show GM130-labelled Golgi alterations in a fraction of mutant SOD1 motor neurons as compared to controls. Note that non-motor neuronal cells (negative for VaChT) show low GM130 labeling without overt structural Golgi alterations. Scale bar 10 μ m.

C. Confocal images showing GM130-labelled Golgi structure in NSC-34 cells transfected for 4 DIV with empty, wildtype SOD1 or mutant SOD1 plasmids.



Subcellular fractionation of spinal cords.

Western blot analyses shows purity of subcellular fractions P10, P100 and S100 from lumbar spinal cord attested by markers of membranes (L1), vesicles (Vti1a) and cytosol (GAPDH).



Flow cytometry analysis of cellular microtubules.

A. Flow cytometry. Diagram showing the cellular content in polymerized α -tubulin in NSC-34 cells after extraction of soluble proteins, incubation with anti- α -tubulin-FITC antibodies and flow cytometry analysis. Solid lines correspond to cells that had been treated with mock (control), Nocodazole or Taxol. Dotted lines correspond to cells incubated without antibodies. Median fluorescence values and their statistical difference with respect to control by chi-test : control 12.800, Nocodazole 4.770, T(x)=5.616 (*); Taxol 34.900, T(x)=5.730 (*).

B. Subcellular fractionation. Immunoblots showing total α -tubulin (T), soluble α -tubulin (S) and polymerized α -tubulin in NSC-34 motor neurons treated with mock (control), Nocodazole or Taxol. The fraction of polymerized tubulin (% P representing P/(S+P)) is indicated on the right. Differences are statistically significant by Mann-Whitney test.



Labeling of spinal cord sections from SOD1 G85R mice with antibodies against Choline Acetyltransferase (ChAT), Stathmins 1 or 2, Golgi SNARE GS28 and DAPI (nuclei).

Motor neurons were identified by ChAT immunoreactivity and large, faintly DAPIstained, nuclei, as indicated by arrows on merged images. Non-motor neuronal cells were identified by negative ChAT and positive DAPI labeling ; some of them are indicated by asterisks.

A. Confocal images show up-regulation of both Stathmin 1 and GS28 in motor neurons. Expression of Stathmin-1 and GS28 in other cell types of mutant SOD1 G85R spinal cord is below threshold. For quantitative immunofluorescence analyses non-saturated images were used and partially sectioned motor neurons (arrowhead) were excluded.

B. Confocal images showing up-regulation of Stathmin 2 and GS28 in a motor neuron.

Scale bar 20 μ m.



Subcellular localization of Stathmins.

Confocal images showing NSC-34 motor neurons transfected with Myc-tagged forms of Stathmin 1 or Stathmin 2 or control, and labeled with anti-Myc antibodies and the co-transfected marker MannosidaseII-GFP (MannII-GFP). Differential interphase contrast (DIC) images indicate cell contours. While Stathmin 1 is mainly cytosolic, Stathmin 2 localizes to the Golgi. Overexpression of Stathmin 1 or 2 causes Golgi disruption. Scale bar 10 μ m.

Supplemental References

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