Figure S1



**Figure S1, related to Figure 1. Flow-cytometry analysis of tumor-infiltrating immune cells in mouse NB models.** Representative flow cytometric gating strategy for 975A2 tumors to define tumor-infiltrating lymphoid cells (**A**), myeloid cells (**B**) and naïve and memory T cells (**C**).

Figure S2



D E



F



**Figure S2, related to Figure 2. Immune content of cisplatin- and vincristine-treated 975A2 tumors.** Flow-cytometry analysis of the immune compartment in 975A2 tumors treated with cisplatin (**A-C**) and vincristine (**D-F**) for 7 days. Levels of significance for comparison between samples were determined by two-tailed Student's t test. CTR, vehicle control; CDDP, cisplatin; VINC, vincristine. Non-significant data are shown.





F % of MHC II<sup>+</sup> CD11c<sup>+</sup> 100

G



Η GZMB GZMB 100 40 % of CD8 75 30 % of NK 20 50 10 25 0 0 CTR+ MTX-OXP-CTR-MTX OXP-È Ŕ Ä Ä 7 Days 7 Days

**Figure S3 related to Figure 2. Immune content of drug-treated 975A2 tumors.** Flow-cytometry analysis of the immune content in 975A2 tumors 1 day (**A-C**) and 7 days (**D-F**) after treatment with the indicated drugs. (**G-H**) Flow-cytometry analysis of the granzyme B expression in tumor-infiltrating CD8<sup>+</sup> T cells (**G**) and NK cells (**H**) in 975A2 tumors 7 days after treatment. Levels of significance for comparison between samples were determined by ANOVA. CTR, vehicle control; DX, doxorubicin; IRI, irinotecan; MTX, mitoxantrone; OXP, oxaliplatin; GZMB, granzyme B. Non-significant data are shown.

Figure S4





G





75

50-

25

0-

CTR

aTGFB

7 Days



#### Figure S4 related to Figure 3. Effect of anti-TGF<sup>β</sup> treatment on immune content of 975A2

**tumors.** (A-E) Flow-cytometry analysis of the immune content in 975A2 tumors 1 day (A-C) and 7 days (D-E) after the start of treatment. (F-G) Flow-cytometry analysis of the activation status of tumor-infiltrating CD8<sup>+</sup> T cells (F) and NK cells (G) in 975A2 tumors 7 days after the start of treatment. Levels of significance for comparison between samples were determined by two-tailed Student's t test. CTR, vehicle control; aTGF $\beta$ , anti-TGF $\beta$ ; GZMB, granzyme B. Non-significant data are shown.







F



splenocytes (relative) 5. 4. 3-2 1 0 Ē aTGFβ-aPD-1-

0.0006

G





**Figure S5 related to Figure 4. Effect of combined drug treatment on the recall and activation of CD8<sup>+</sup> T cells and NK cells. (A-C)** Flow-cytometry analyses of IFNγ and granzyme B expression of CD8<sup>+</sup> T cells and NK cells from splenocytes co-cultured 24 hours with drug-treated 975A2 MDOTS. (**D**) Schematic representation of the microfluidic device. (**E-G**) Representative images of red-labeled splenocyte migration in microfluidic devices versus drug-treated and untreated 975A2 MDOTS after 24 hours of co-culture. The number of splenocytes from tumor-bearing mice migrating versus treated and untreated 975A2 MDOTS was assessed by ImageJ software. Data are shown as fold change  $\pm$  SD. Levels of significance were determined by ANOVA. CTR, vehicle control; DX, doxorubicin; IRI, irinotecan; OXP, oxaliplatin, aTGFβ, anti-TGFβ; aPD-1, anti-PD-1; DT, doxorubicin and anti-TGFβ; DP, doxorubicin and anti-PD-1; DTP, doxorubicin, anti-TGFβ and anti-PD-1. IT, irinotecan and anti-TGFβ; OP, oxaliplatin and anti-PD-1; ITP, irinotecan, anti-TGFβ and anti-PD-1. OT, oxaliplatin and anti-TGFβ; OP, oxaliplatin and anti-PD-1; OTP, oxaliplatin, anti-TGFβ and anti-PD-1; GZMB, granzyme B. Statistically significant P values are shown.















Figure S6 related to Figure 5. Treatment of mitoxantrone in combination with TGF $\beta$  and PD-1 blockade delays growth of subcutaneously transplanted 975A2 tumors and reshapes intratumoral immune infiltrate. (A) Schematic representation of the drug treatment and timing of tumor immune infiltrate analysis. (B) Tumor growth of 975A2 injected subcutaneously in C57BL/6 mice and treated as indicated. Significance at day 28 (Mann Whitney test). (C) Weight of explanted tumors at day 28 after cell inoculation. (D-E) Flow-cytometry analysis of the immune compartment in 1 day-MTX-treated 975A2 tumors. Levels of significance for comparison between samples were determined by two-tailed Student's t test. (F) Chemokine expression in 1 day-MTX-treated 975A2 tumor lysates by protein array. Relative chemokine expression based on densitometric analysis is shown on the right. (G-I) Flow-cytometry analysis of the immune compartment. (L-M) Flowcytometry analysis of the activation status of tumor-infiltrating CD8<sup>+</sup> T cells (L) and NK cells (M) in 975A2 tumors 12 days after treatment is shown. Levels of significance for comparison between samples in G-M were determined by ANOVA. CTR, vehicle control; MTX, mitoxantrone; MTP, mitoxantrone, anti-TGF $\beta$  and anti-PD-1; GZMB, granzyme B. Statistically significant P values are shown.



Figure S7 related to Figure 5. Tumor growth of drug-treated 975A2 and 9464D. Tumor growth of 975A2 (A) and 9464D (B) injected subcutaneously in C57BL/6 mice and treated as indicated. The average growth of the control tumors is indicated in each graph with a dotted line. CTR, vehicle control; MTX, mitoxantrone; aTGF $\beta$ , anti-TGF $\beta$ ; aPD-1, anti-PD-1; MT, mitoxantrone and anti-TGF $\beta$ ; MP, mitoxantrone and anti-PD-1; MTP, mitoxantrone. Levels of significance for comparison between samples were determined by Mann Whitney test. Non-significant data are shown.



#### Figure S8 related to Figure 5. Expression of calreticulin in 9464D tumors.

Representative example of calreticulin (CALR) staining in 9464D tumors grown subcutaneously in C57BL/6 mice. Brown, calreticulin-expressing cells. Nuclei were counterstained with hematoxylin (blue). Original magnifications, x20. Scale bars,  $30 \,\mu$ m. IHC density was assessed by ImageJ software by measuring 5 different areas of the slide on five independent mice for each condition, and the mean  $\pm$  SD of integrated density are reported in the histogram. Levels of significance for comparison between samples were determined by two-tailed Student's t test.



#### Figure S9 related to Figure 5 and Figure S6. Immune content of drug-treated 9464D and 975A2

**tumors.** Flow-cytometry analysis of the immune content in 9464D and 975A2 tumors (upper and lower panels, respectively) 1 day (**A-C**) and 12 days (**D-E**) after treatment with the indicated drugs. (**F**) Flow-cytometry analysis of the activation status of tumor-infiltrating CD8<sup>+</sup> T cells in 9464D and 975A2 tumors (upper and lower panels, respectively) 12 days after treatment. Levels of significance for comparison between samples were determined by two-tailed Student's t test (**A-C**) and ANOVA (**D-F**). CTR, vehicle control; MTX, mitoxantrone; MTP, mitoxantrone, anti-TGF $\beta$  and anti-PD-1; GZMB, granzyme B. Non-significant data are shown.



Cell/gr 9464D on MHC II<sup>+</sup> CD11c<sup>+</sup> x10<sup>2</sup> MTX MTX MTX MTX MTX MTX MTX

7 Days



# Figure S10 relative to Figure 6. Immune content of drug-treated 9464D grown in the adrenal gland. (A-C) Flow-cytometry analysis of the immune content in 9464D 7 days after the start of treatment with the indicated drugs is shown. (D) Flow-cytometry analysis of the activation status of tumor-infiltrating CD8<sup>+</sup> T cells in 9464D tumors 7 days after the start of treatment is shown. Levels of significance for comparison between samples were determined by ANOVA. Non-significant data are shown. CTR, vehicle control; MTX, mitoxantrone; MTP, mitoxantrone, anti-TGF $\beta$ and anti-PD-1; GZMB, granzyme B.



# Figure S11 relative to Figure 7. Mitoxantrone recalls activated CD8<sup>+</sup> T cells and NK cells in some PDOTS when combined with TGFβ and PD-1 blockade.

(A) Representative images of PDOTS derived from P4 NB patient. Original magnification, x20. Scale bar, 30  $\mu$ m. (**B and C**) Flow-cytometry analysis of granzyme B expression by CD8<sup>+</sup> T cells (**B**) and NK cells (**C**) of autologous PBMCs co-cultured for 24 hours with drug-treated PDOTS. (**D**) Representative images of the migration of red-labeled autologous PBMCs in microfluidic devices to drug-treated and untreated PDOTS after 24 hours of co-culture. The number of PBMCs migrating versus drug-treated and untreated PDOTS was assessed by ImageJ software. Data are shown as fold change ± SD. Levels of significance for comparison between samples were determined by two-tailed Student's t test. MTX, mitoxantrone; CTR, vehicle control; MTP, mitoxantrone, anti-TGF $\beta$  and anti-PD-1; GZMB, granzyme B. Non-significant data are shown.



Figure S12. Mitoxantrone TME of drug-treated 9464D grown in the adrenal gland. (A-B) Representative flow-cytometry analyses (left) of MHC class I (A) and PD-L1 (B) in MDOTS derived from 9464D orthotopic model treated 24 hours with MTX. Mice treated with vehicle or MTX are shown as grey and red plots, respectively. The graphs on the right represent the fold changes in MFI  $\pm$  SD from three independent experiments. Levels of significance for comparison between samples were determined by the two-tailed Student's *t* test. CTR, vehicle control; MTX, mitoxantrone; MFI, mean fluorescence intensity. Statistically significant, *P* values are shown.

MARKER	Fluorochrome	Clone	Company	Catalog	Application	Diluition
anti-mouse CD45	BV605	30-F11	<b>BD</b> Biosciences	563053	FACS	1:40
anti-mouse B220	APC	RA3-6B2	<b>BD</b> Biosciences	553092	FACS	1:40
anti-mouse CD3	BUV395	145-2C11	<b>BD</b> Biosciences	565992	FACS	1:40
anti-mouse CD4	PE CY7	GK1.5	<b>BD</b> Biosciences	563933	FACS	1:40
anti-mouse CD8	BUV805	53-6.7	<b>BD</b> Biosciences	612898	FACS	1:40
anti-mouse CD25	PE	PC61	<b>BD</b> Biosciences	553866	FACS	1:40
anti-mouse NK1.1	BV421	PK136	<b>BD</b> Biosciences	562921	FACS	1:40
anti-mouse FOXP3	PerCP CY5.5	FJK-16s	eBiosciences	45-5773-82	FACS	1:10
anti-mouse CD44	BV421	IM7	<b>BD</b> Biosciences	563970	FACS	1:40
anti-mouse CD62L	APC	<b>MEL-14</b>	<b>BD</b> Biosciences	561919	FACS	1:40
anti-mouse CD69	APC CY7	H1.2F3	<b>BD</b> Biosciences	561240	FACS	1:40
anti-mouse TGFβ	PE	/	<b>R&amp;D SYSTEMS</b>	IC1835P	FACS	1:10
anti-mouse CD11B	PerCP CY5.5	M1/70	<b>BD</b> Biosciences	550993	FACS	1:40
anti-mouse CD11C	BUV395	N418	<b>BD</b> Biosciences	744180	FACS	1:40
anti-mouse MHC II	APC	M5/114.15.2	BioLegend	107614	FACS	1:40
anti-mouse Ly6C	APC CY7	HK1.4	BioLegend	128026	FACS	1:40
anti-mouse Ly6G	PE CY7	1A8	BioLegend	127618	FACS	1:40
anti-mouse F4/80	PE	BM8	BioLegend	123109	FACS	1:40
anti-mouse CD103	BV421	2E7	BioLegend	121421	FACS	1:40
anti-mouse CD40	PE	1C10	eBiosciences	12-0401-82	FACS	1:40
anti-mouse CD86	PE CY7	GL1	eBiosciences	25-0862-82	FACS	1:40
anti-mouse IFNγ	PE CY7	XMG1.2	<b>BD</b> Biosciences	561040	FACS	1:10
anti-mouse GRZB	APC	NGZB	eBiosciences	17-8898-82	FACS	1:10
anti-mouse PD-1	APC	J43	<b>BD</b> Biosciences	562671	FACS	1:40
anti-mouse PD-L1	BV711	MIH5	<b>BD</b> Biosciences	563369	FACS	1:40
anti-mouse MHC-I	PerCP Cy5.5	C3H	Biolegend	114619	FACS	1:40
anti-mouse GD2	AF647	14.G2a	<b>BD</b> Biosciences	562096	FACS	1:40
anti-human CD45	BUV805	HI30	<b>BD</b> Biosciences	612891	FACS	1:40
anti-human CD3	<b>BB700</b>	SK7	<b>BD</b> Biosciences	566575	FACS	1:40
anti-human CD8	BUV395	RPA-T8	<b>BD</b> Biosciences	563795	FACS	1:40
anti-human NKp46	BV711	9E2/NKp46	<b>BD</b> Biosciences	563043	FACS	1:40
anti-human GZMB	BV510	GB11	<b>BD</b> Biosciences	563388	FACS	1:40
anti-mouse/human synaptophysin	/	SY38	Invitrogen	MA1-213	IHC	1:50
calreticulin	/	/	Abcam	ab2907	IHC	1:500
anti-mouse NK.1.1	/	PK136	Invitrogen	MA1-70100	IF	1:50
anti-mouse GZMB	/	D2H2F	Cell Signaling	17215	IF	1:100
Goat-anti-mIgG	Alexa-Fluor488	/	Invitrogen	A-11017	IF	1:500
Goat-anti-rIgG	Alexa-Fluor594	/	Invitrogen	A-11037	IF	1:500

 Table S1. List of antibodies

**Table S2:** Clinical and genetic characteristics of 7 newly diagnosed NB patients.

Patient	Age ad diagnosis (months)	Gender	Stage (INRG)	Histologic classification (INCP or Shimada)	Grade of NB differentiation (INCP or Shimada)	MYCN status	PDOTS formation	PDOTS responsiveness
P1	3	Male	L1	Favorable	Undifferentiated	Not amplified	No	/
P2	18	Male	L1	Intermediate	Undifferentiated	Not amplified	No	/
P3	13	Male	L2	Favorable	Undifferentiated	Not amplified	No	/
P4	35	Female	L2	Unfavorable	Undifferentiated	Not amplified	Yes	No
P5	22	Female	Μ	Unfavorable	Undifferentiated	Not amplified	Yes	Yes
<b>P6</b>	96	Male	Μ	Unfavorable	Undifferentiated	Not amplified	Yes	Yes
<b>P7</b>	1	Male	MS	Favorable	Undifferentiated	Not amplified	Yes	Yes

Patients analyzed are indicated in bold.