

**Targeting cancer stem cell OXPHOS with tailored ruthenium complexes as a
new anti-cancer strategy**

Sonia Alcalá^{1,2}, Lara Villarino³, Laura Ruiz-Cañas^{1,2}, José R. Couceiro³, Miguel Martínez-Calvo³, Adrián Palencia-Campos^{1,2}, Diego Navarro^{1,2}, Pablo Cabezas-Sainz⁵, Iker Rodriguez-Arabaolaza^{1,4}, Alfonso Cordero-Barreal^{1,2}, Lucia Trilla-Fuertes^{6,7}, Juan A. Rubiolo⁵, Sandra Batres-Ramos^{1,2}, Mireia Vallespinos^{1,2}, Cristina González-Páramos^{1,8,9}, Jéssica Rodríguez³, Angelo Gámez-Pozo^{6,7}, Juan Ángel Fresno Vara^{6,10}, Sara Fra Fernández¹¹, Amparo Benito Berlinches^{2,12}, Nicolás Moreno Mata¹¹, Ana María Torres Redondo¹³, Alfredo Carrato^{2,10}, Patrick C. Hermann¹⁴, Laura Sánchez⁵, Susana Torrente¹⁵, Miguel Ángel Fernández-Moreno^{1,8,9}, José L. Mascareñas^{3,*} and Bruno Sainz, Jr.^{1,2,10,*}

Supplementary Material

Supplementary Figures S1-S6

Supplementary Tables S1-S3

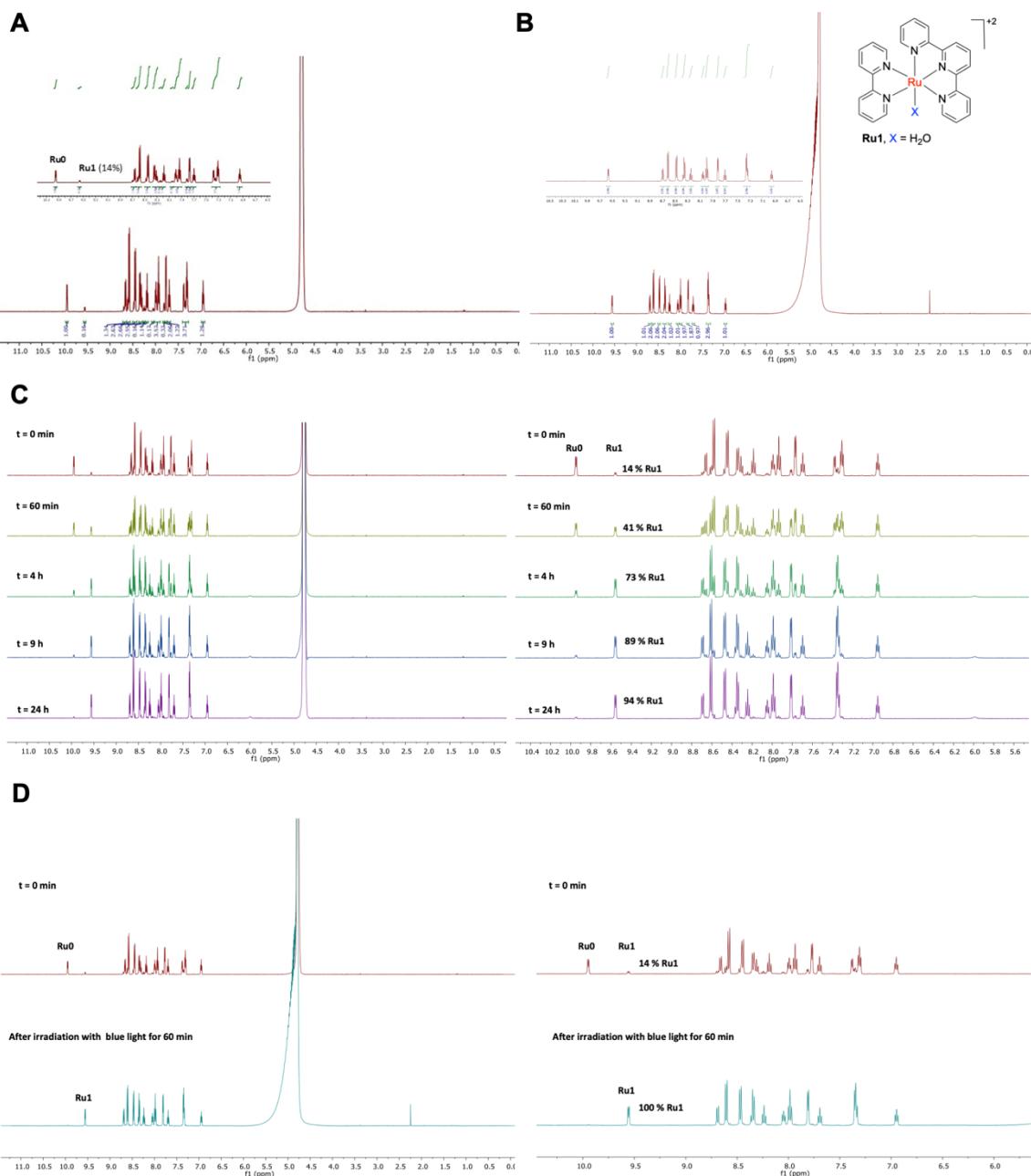


Fig. S1 NMR studies of the aquation of $[\text{Ru}(\text{terpy})(\text{bpy})\text{Cl}]^+$ complex (Ru0) using deuterium oxide. **A** $^1\text{H-NMR}$ of $[\text{Ru}(\text{terpy})(\text{bpy})\text{Cl}]^+$ complex (Ru0) in water at $t = 0$. **B** $^1\text{H-NMR}$ of $[\text{Ru}(\text{terpy})(\text{bpy})]^+ \text{H}_2\text{O}^{+2}\text{Cl}^{-2}$ complex (Ru1) in water. **C** $^1\text{H-NMR}$ of $[\text{Ru}(\text{terpy})(\text{bpy})\text{Cl}]^+$ complex (Ru0) in water at different times (starting concentration of $\text{Ru0} = 2\text{mM}$). **D** $^1\text{H-NMR}$ of $[\text{Ru}(\text{terpy})(\text{bpy})\text{Cl}]^+$ complex (Ru0) after dissolving in water ($t = 0$ min) and after irradiation with visible light for 60-120 min (starting concentration of $\text{Ru0} = 2\text{mM}$).

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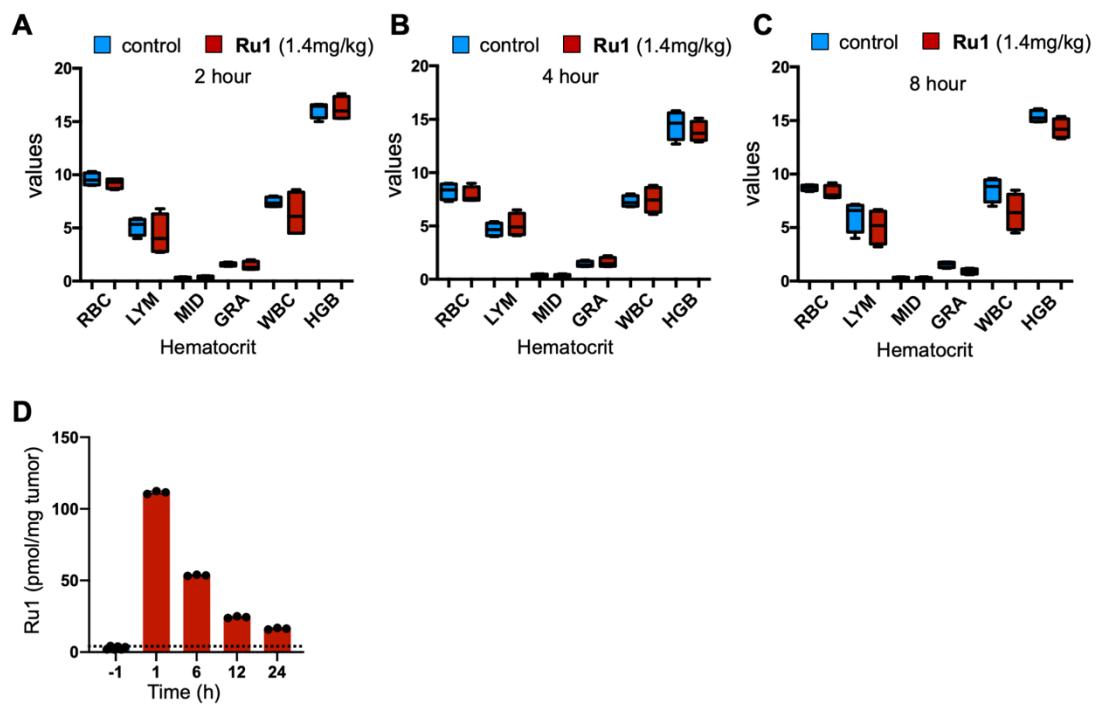


Fig. S2 Analysis of Ru1 toxicity *in vivo*. **A-C** Average values \pm SEM of indicated hematocrit parameters determined from blood of mice extracted 2h (A), 4h (B), or 8h (C) post treatment with diluent control (Ctl) or Ru1 (1.4mg/kg, r.o). No significant differences were found, as

determined by unpaired two-sided Student's t-test. **D** Picomoles of Ru1 per mg of tumor, determined by analyzing ruthenium with ICP-MS, from tumors extracted at indicated time points post treatment initiation. Dashed line indicates the background of the assay.

A

Gene	Description	Log2 Fold Change	p value	p adj
MT-ND5	MT Encoded NADH:Ubiquinone Oxidoreductase Core Subunit 5	-4.583453047	2.727E-21	2.507E-17
MT-ND4L	MT Encoded NADH:Ubiquinone Oxidoreductase Core Subunit 4L	-4.139355065	1.339E-19	6.842E-16
MT-ND6	MT Encoded NADH:Ubiquinone Oxidoreductase Core Subunit 6	-4.533419675	1.488E-19	6.842E-16
MT-ND4	MT Encoded NADH:Ubiquinone Oxidoreductase Core Subunit 4	-4.79740817	3.726E-19	1.370E-15
MT-CYB	MT Encoded Cytochrome B	-4.546259595	4.588E-19	1.406E-15
MT-ATP8	MT ATP Synthase Membrane Subunit 8	-3.980573335	1.279E-15	3.359E-12
MT-CO1	MT Encoded Cytochrome C Oxidase I	-4.436182642	2.545E-13	5.200E-10
MT-CO3	MT Encoded Cytochrome C Oxidase 3	-4.162005073	7.644E-13	1.405E-09
MT-ATP6	MT ATP Synthase Membrane Subunit 6	-4.127331673	1.441E-12	2.408E-09
MT-CO2	MT Encoded Cytochrome C Oxidase 2	-4.009943774	8.000E-09	1.131E-05
MT-ND3	MT Encoded NADH:Ubiquinone Oxidoreductase Core Subunit 3	-4.683257976	4.332E-07	5.310E-04
MT-TF	MT Encoded tRNA-Phe (UUU/C)	3.177659894	3.776E-06	4.340E-03
MT-TS1	MT Encoded tRNA-Ser (UCN) 1	-4.245403579	5.998E-06	6.487E-03
MT-TC	MT Encoded tRNA-Cys (UGU/C)	-4.141347134	1.300E-05	0.0125823

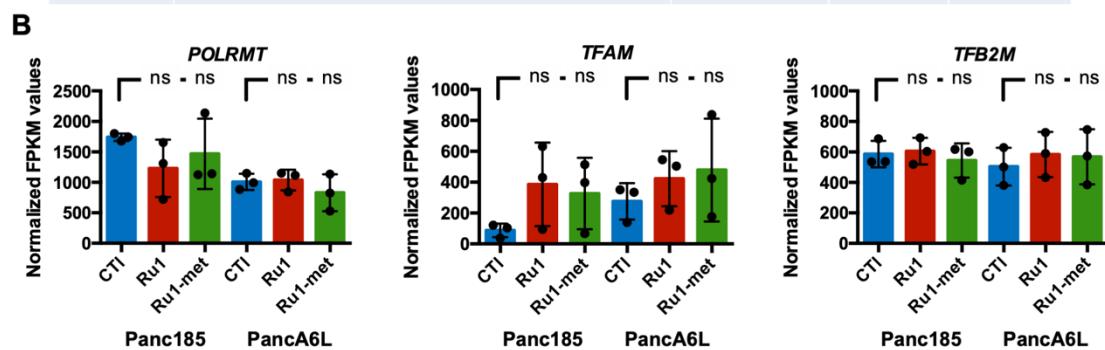


Fig. S3 *Ru1* negatively regulates MT-encoded genes. **A** Table summarizing the 14 mtDNA-encoded genes modulated in PancA6L spheres treated with *Ru1* (100μM, 24 h) compared to untreated Controls. Shown are the gene name, description, Log2 fold change, p value and p adjusted (adj). **B** Mean ± SD of normalized

Fragments Per Kilobase Million (FPKM) values for the indicated target genes in Ctl-, *Ru1* and *Ru1-met*-treated Panc185 or PancA6L spheres. (ns= not significant, as determined by one-way ANOVA with Dunnett post-test, compared to Control).

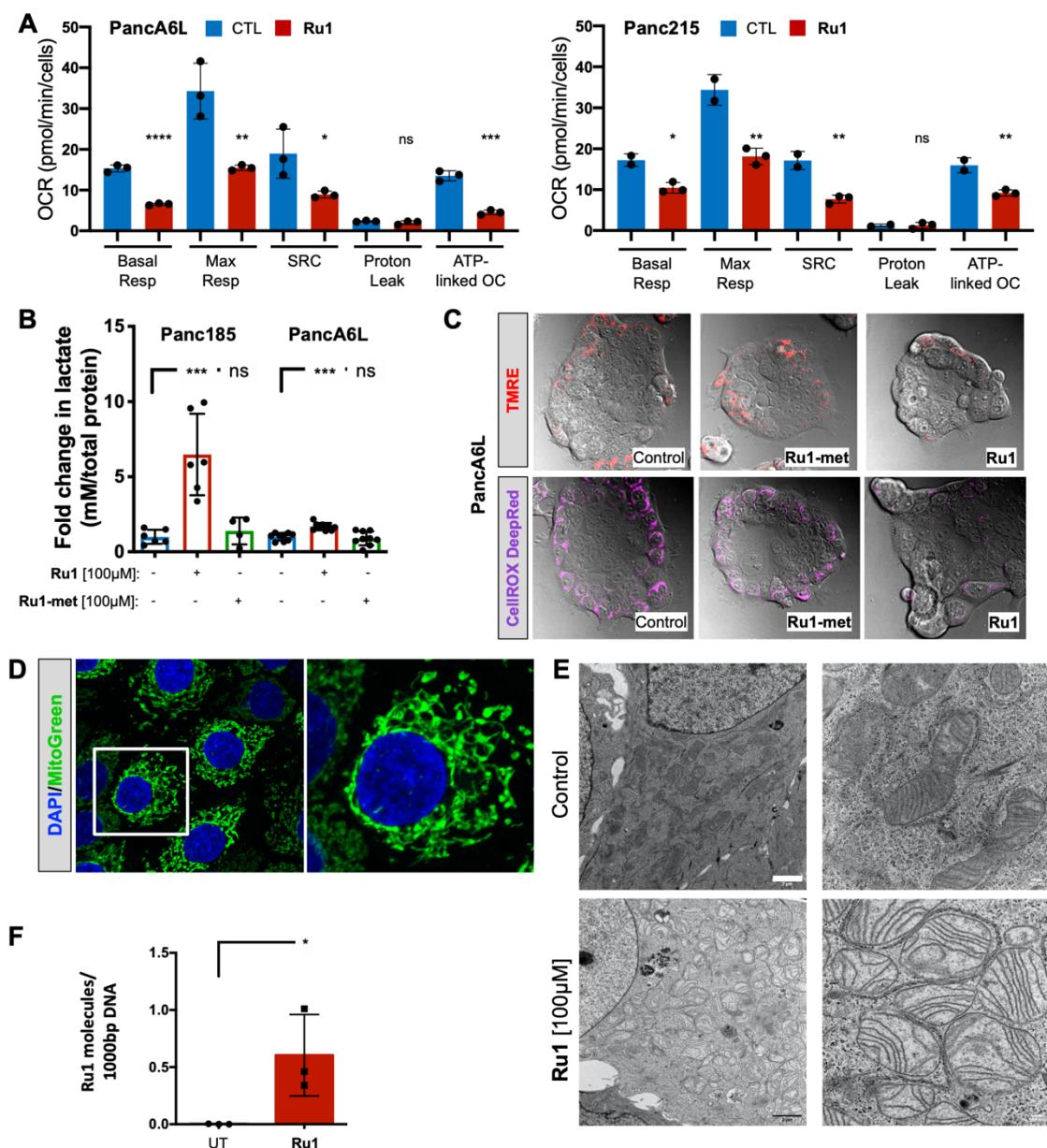


Fig. S4 *Ru1* affects PaCSC oxygen consumption and mitochondrial properties and morphology. **A** Measured and calculated mean \pm SD oxygen consumption rate (OCR) parameters (Resp = Respiration; Max = Maximum; SRC = Spare Respiratory Capacity; OC = Oxygen consumption) in Ctl- and *Ru1*-treated PancA6L or Panc215 spheres ($n = 3$ biological replicates with 3 readings). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, ns = not significant, as determined by unpaired two-sided Student's t-test. **B** Mean fold-change \pm SD in lactate (mM/total protein) in untreated (-), *Ru1* (100 μ M) or *Ru1-met* (100 μ M)-treated Panc185 and PancA6L cells compared to control, set as 1.0. *** $p < 0.001$, ns = not significant, as determined by one-way ANOVA with Dunnett post-test, compared to Control. **C**

Representative IF confocal images of TMRE (mitochondria membrane potential) or CellROX DeepRed (ROS) staining in untreated (Control), *Ru1* (100 μ M) or *Ru1-met* (100 μ M)-treated PancA6L cells (24 h). **D** Representative fluorescence confocal images of MitoGreen (mitochondrial mass) and DAPI (Blue) staining in *Ru1* (100 μ M)-treated PancA6L cells (24 h). Scale bar = 10 μ M. **E** Representative transmission electron micrographs of Control (untreated) or *Ru1* (100 μ M)-treated Panc185 cells (24 h). Mitochondria are better defined in the Control compared to *Ru1*-treated samples. **F** Amount of *Ru1* molecules per 1000bp of DNA, determined by ICP, in mtDNA isolated from gradient-purified mitochondria from untreated (CTL) or *Ru1*-treated (100 μ M, 2 h) Panc185 cells.

Fig. S5 GQ in the mtDNA determined with G4 hunter.

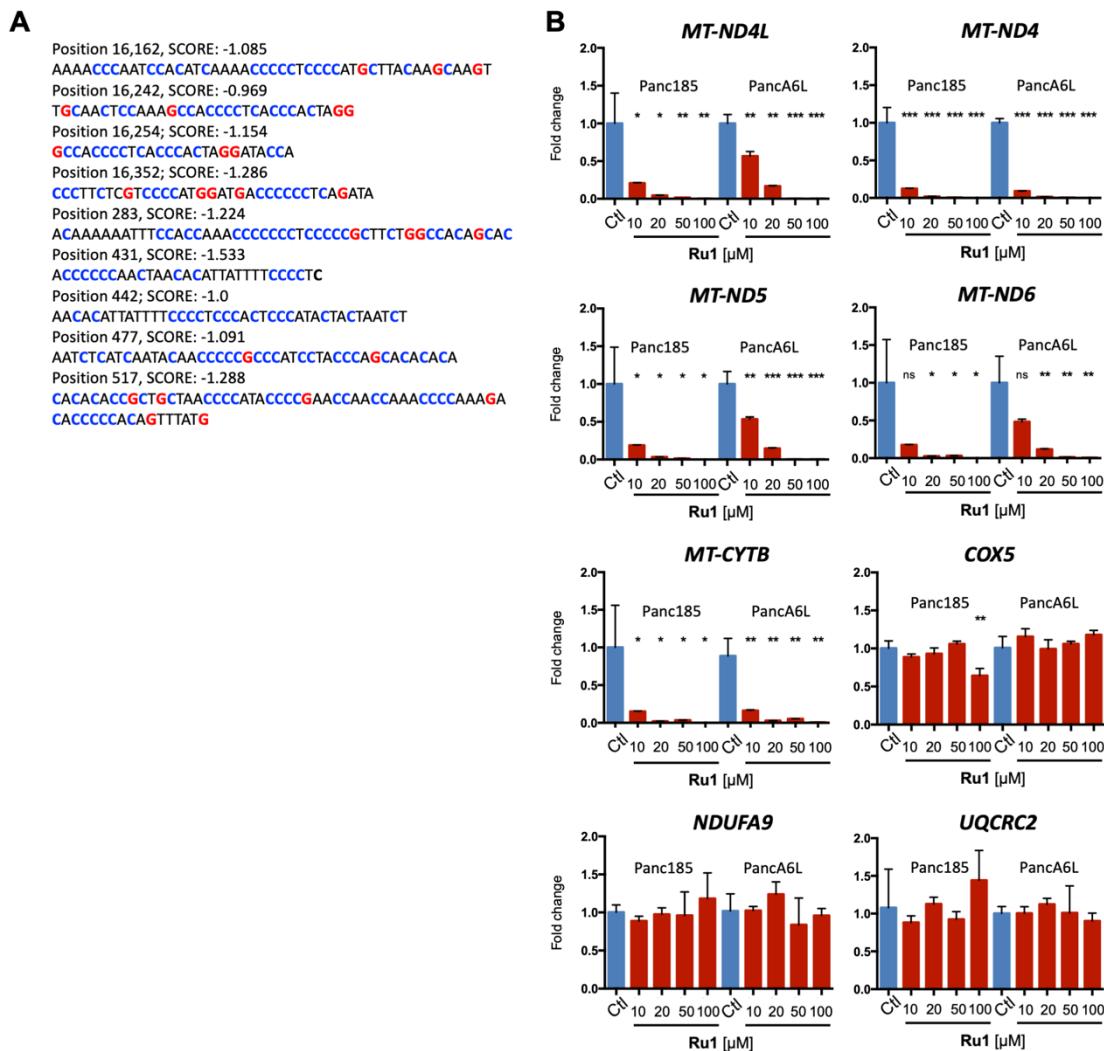


Fig. S6 *Ru1* inhibits mtDNA transcription. **A** Sequence of 9 predicted GQs in the D-loop region, their positions and G4Hunter score indicating G4-prone structures. Isolated guanines (**G**) are shown in red, and cysteines (**C**) in blue. **B** Mean fold-change \pm SD in the relative mRNA expression of the indicated mtDNA- and nuclear-encoded genes as a function of increasing concentrations of *Ru1* in Panc185 or PanA6L cells (48h treatment). Values were normalized to β -actin levels. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, as determined by one-way ANOVA with Dunnett post-test, compared to untreated (Ctl).

nuclear-encoded genes as a function of increasing concentrations of *Ru1* in Panc185 or PanA6L cells (48h treatment). Values were normalized to β -actin levels. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, as determined by one-way ANOVA with Dunnett post-test, compared to untreated (Ctl).

SUPPLEMENTARY TABLES**Table S1 Antibodies**

1 ^a Abs-Epitope	Source	Dilution	Application	Manufacturer (Catalog no.)
α-hu/mu-GAPDH	Mouse monoclonal	1:5000	WB	Abcam (Cat no. ab8245)
Total OXPHOS Human WB Antibody Cocktail	Mouse monoclonal	1:500	WB	Abcam (Cat no. ab110411)
α-hu-CD133/1-APC	Mouse monoclonal	1:50	FC	Miltenyi Biotec (Cat no. 130-090-826)
α-hu-EpCAM-FITC	Mouse monoclonal	1:50	FC	Miltenyi (Cat no. 130-110-998)
α-hu-CXCR4-PE	Mouse monoclonal	1:50	FC	Miltenyi (Cat no. 130-117-354)
α-hu-CD90-APC	Mouse monoclonal	1:20	FC	Molecular Probes (Cat no. A15726)

2 ^a Abs-Epitope	Source	Dilution	Application	Manufacturer (Catalog no.)
Anti-mouse-HRP	Sheep	1:5000	WB	Amersham (NA931)
Anti-rabbit-HRP	Donkey	1:5000	WB	Amersham (NA934)

Table S2 RTqPCR primer sequences

Gene	Species	Primer sense	Primer antisense
β-ACTIN	human	GCGAGCACAGGAGCCTCGCCTT	CATCATCCATGGTGAGCTGGCGG
POU5F1	human	CTTGCTGCAGAAGTGGGTGGAGGAA	CTGCAGTGTGGGTTTCGGGCA
SOX2	human	AGAACCCCCAAGATGCACAAC	CGGGGCCGGTATTATAATC
PGC1A	human	TGACTGGCGTCATTCAAGGAG	CCAGAGCAGCACACTCGAT
MT-ND1	human	GCACTGCGAGCAGTAGCCCCA	TGGCCAAGGGTCATGATGGCA
MT-ND2	human	AACTCCAGCACCACGACCCCT	AAAAGCCGGTTAGCGGGGGC
MT-CO1	human	CTCTTCGTCATGCCCTCCT	ATTCCGAAGCCTGGTAGGAT
MT-CO2	human	CGCCCTCCCATCCCTACGCA	CCGCCGTAGTCGGTGTACTCG
MT-ATP8	human	CCACCTACCTCCCTCACCAAAGC	TGGGGGCAATGAATGAAGCGAAC
MT-ATP6	human	TCCCCTTATGAGCGGGCACAG	TAGGCGTACGCCAGGGCTA
MT-CO3	human	GCCCTCCTAATGACCTCCGGC	TGGACAGGTGGTGTGGTGG
MT-ND3	human	TCGACCCTATATCCCCCGCC	TGGTAGGGTAAAGGAGGGCA
MT-ND4L	human	TCGCTCACACCTCATCCTCCCTA	AGAGGGAGTGGTGTGGAGGGTT
MT-ND4	human	TCGCCCACGGGCTTACATCC	AGGCGAGGTTAGCGAGGCCTT
MT-ND5	human	TCCGCTTCCACCCCTAGCA	GGCGCAGACTGCTGCGAACAA
MT-ND6	human	GATTGTTAGCGGTGTGGTCGGGT	GACCTAACCCCTGACCCCCA
MT-CYTB	human	ACCAAGACGCCCTAACCGCC	GCCTCGCCCCATGTGTAGGAA
COX5	human	CTTTCGCGGCATGCAGACGG	AGCCCCATCCATGCGGTTTACACT
NDUFA9	human	AGTGGAGCGGATGCACATCACA	GACGGTCTGGCCGGCTTCA
UQCRC2	human	GGCCACAGCTGCTGGAGATGTTA	GCAACTAGAGCCTGGGACCCG
Hprt	mouse	TCCTCCTCAGACCGTTTT	CCTGGTTCATCATCGCTAACATC
Atp6	mouse	TCCCAATCGTTGTAGCCATCA	AGACGGTTGTTGATTAGGCCT
Cox1	mouse	ATCACTACCAGTGCTAGCCG	CCTCCAGCGGGATCAAAGAA
Drp1	mouse	ATGCCAGCAAGTCCACAGAA	TGTTCTCGGGCAGACAGTTT

Table S3 PCR primer sequences

Gene	Species	Primer sense	Primer antisense
D-loop	human	CTCACCCATCAACAACCGCT	TATGGGGTGATGTGAGCCCG
MT-RNR2	human	TTCAAGCTCAACACCCACTACC	GGAGCCATTACAGGTCCATT