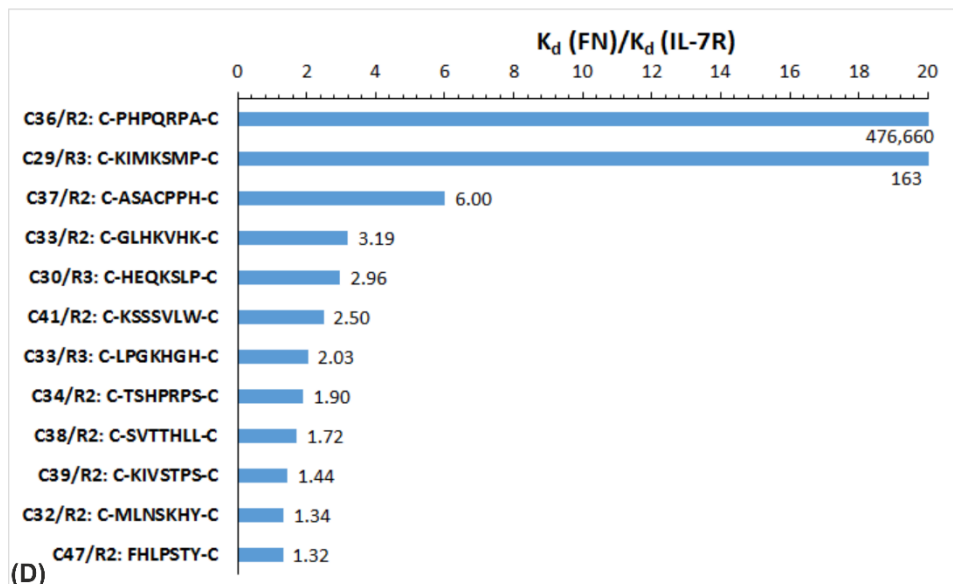
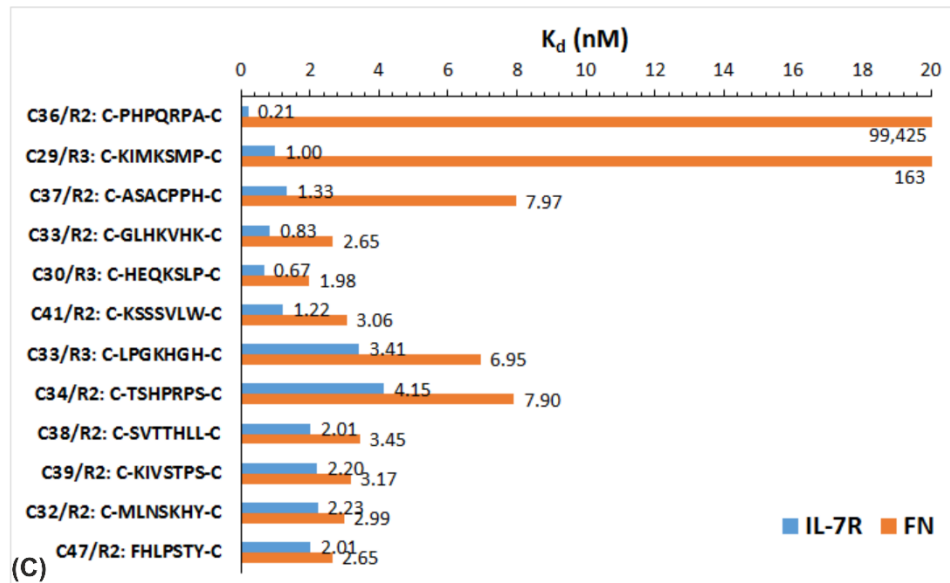
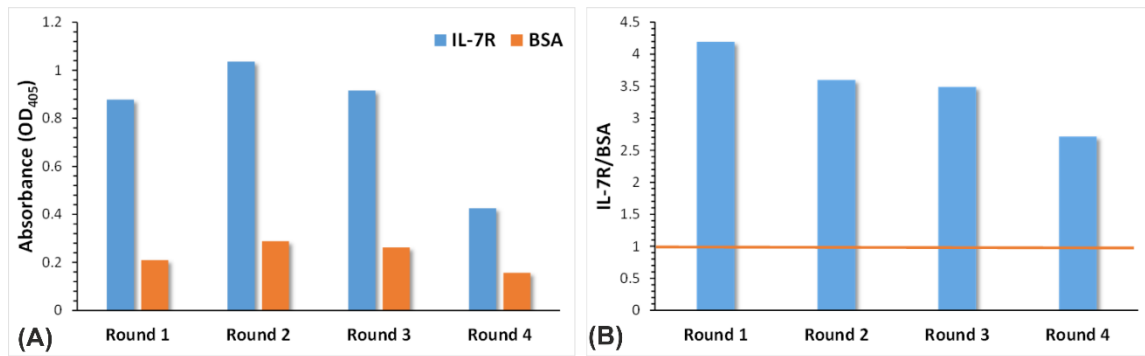


## **Additional File 2: Figures**

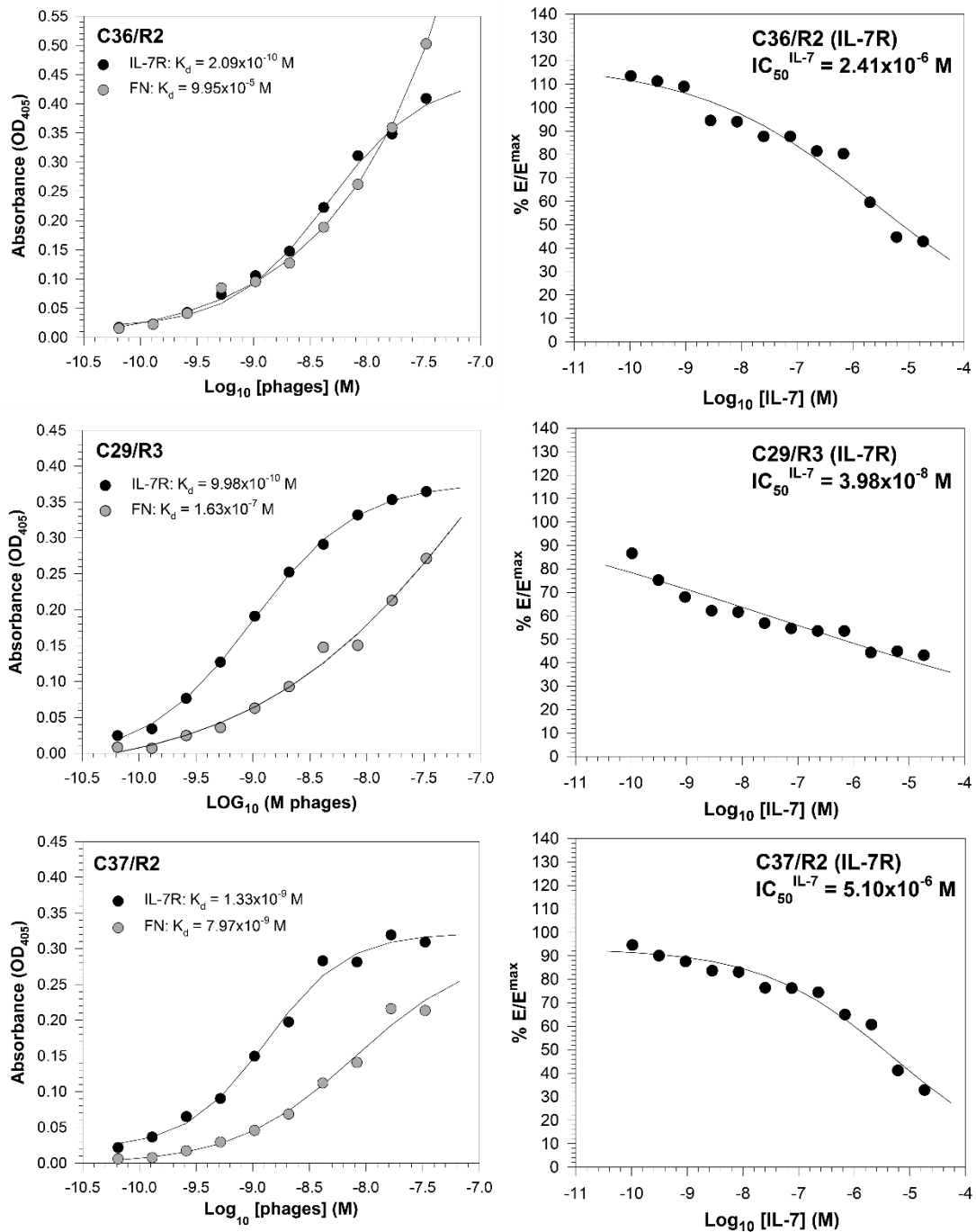
### **Screening for peptides targeted to IL-7R $\alpha$ for molecular imaging of rheumatoid arthritis synovium**

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Sébastien Sauvage<sup>2,3,7</sup>, Marie-Claire Beckers<sup>4,8</sup>, Sandrine Rorive<sup>2,3</sup>, Isabelle Salmon<sup>2,3</sup>,  
Luce Vander Elst<sup>1,5</sup>, Bernard R. Lauwerys<sup>6</sup>, Robert N. Muller<sup>1,5</sup>

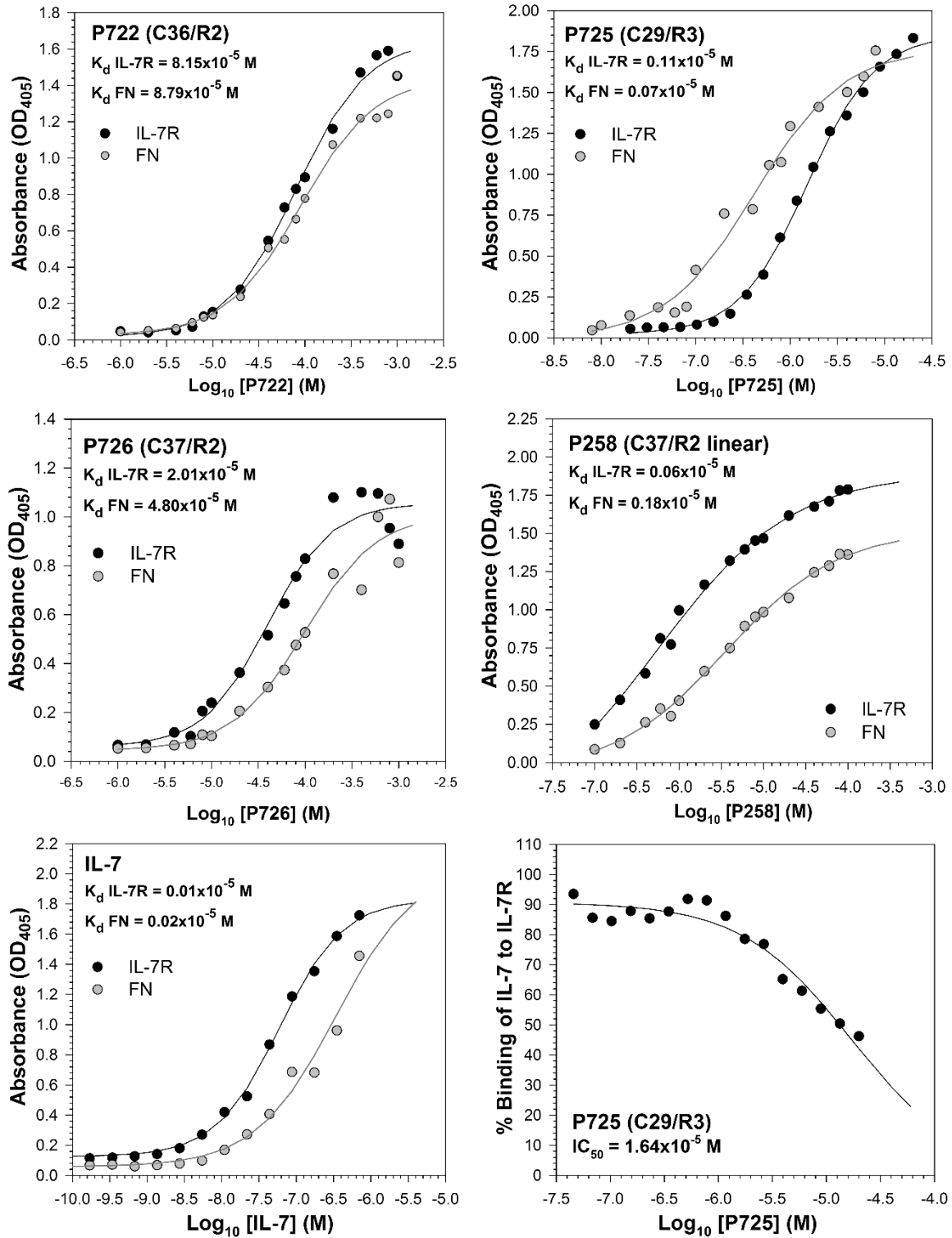
<sup>1</sup> Department of General, Organic and Biomedical Chemistry, NMR and Molecular Imaging Laboratory, University of Mons, Avenue Maistriau 19, Mendeleev Building, B-7000 Mons, Belgium; <sup>2</sup> Department of Pathology, Erasme Hospital, Université Libre de Bruxelles, Route de Lennik 808, 1070 Brussels, Belgium; <sup>3</sup> DIAPath, Center for Microscopy and Molecular Imaging, 8, rue Adrienne Bolland - 6041 Gosselies, Belgium; <sup>4</sup> Eurogentec S.A. Liège Science Park, Rue du Bois Saint-Jean 5, B-4102 Seraing, Belgium; <sup>5</sup> Center for Microscopy and Molecular Imaging, 8, rue Adrienne Bolland - 6041 Gosselies, Belgium; <sup>6</sup> Pôle de pathologies rhumatismales inflammatoires et systémiques, Institut de Recherche Expérimentale et Clinique, Université Catholique de Louvain, Avenue Mounier 53, 1200 Brussels, Belgium; <sup>7</sup> Present address: Molecular Pathology Laboratory, ONCODNA – The Cancer Theranostic Company, 25 Av. Georges Lemaître, 6041 Gosselies, Belgium; <sup>8</sup> Present address: ASIT Biotech s.a., 3 Rue des Chasseurs Ardennais, 4031 Angleur, Belgium



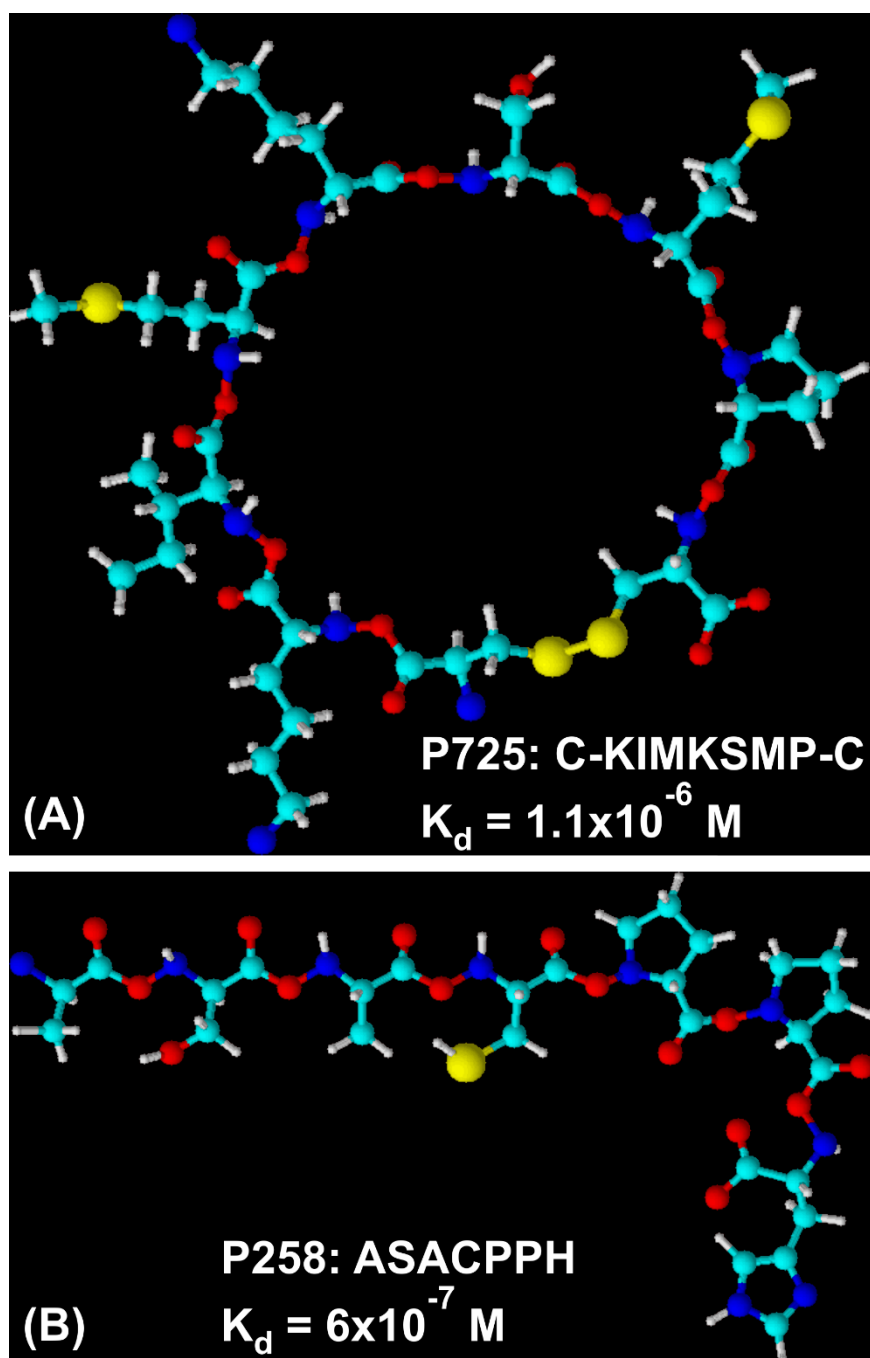
**Figure S1.** The affinity for IL-7R and control proteins (BSA and FN) of the outputs derived from the four rounds of panning (**A** and **B**) and of the selected phage clones (**C** and **D**) isolated from the pools of the second (R2) and the third (R3) rounds of panning. The results are expressed as absolute values (**A** and **C**) or as a ratio between IL-7R and control proteins (**B** and **D**).



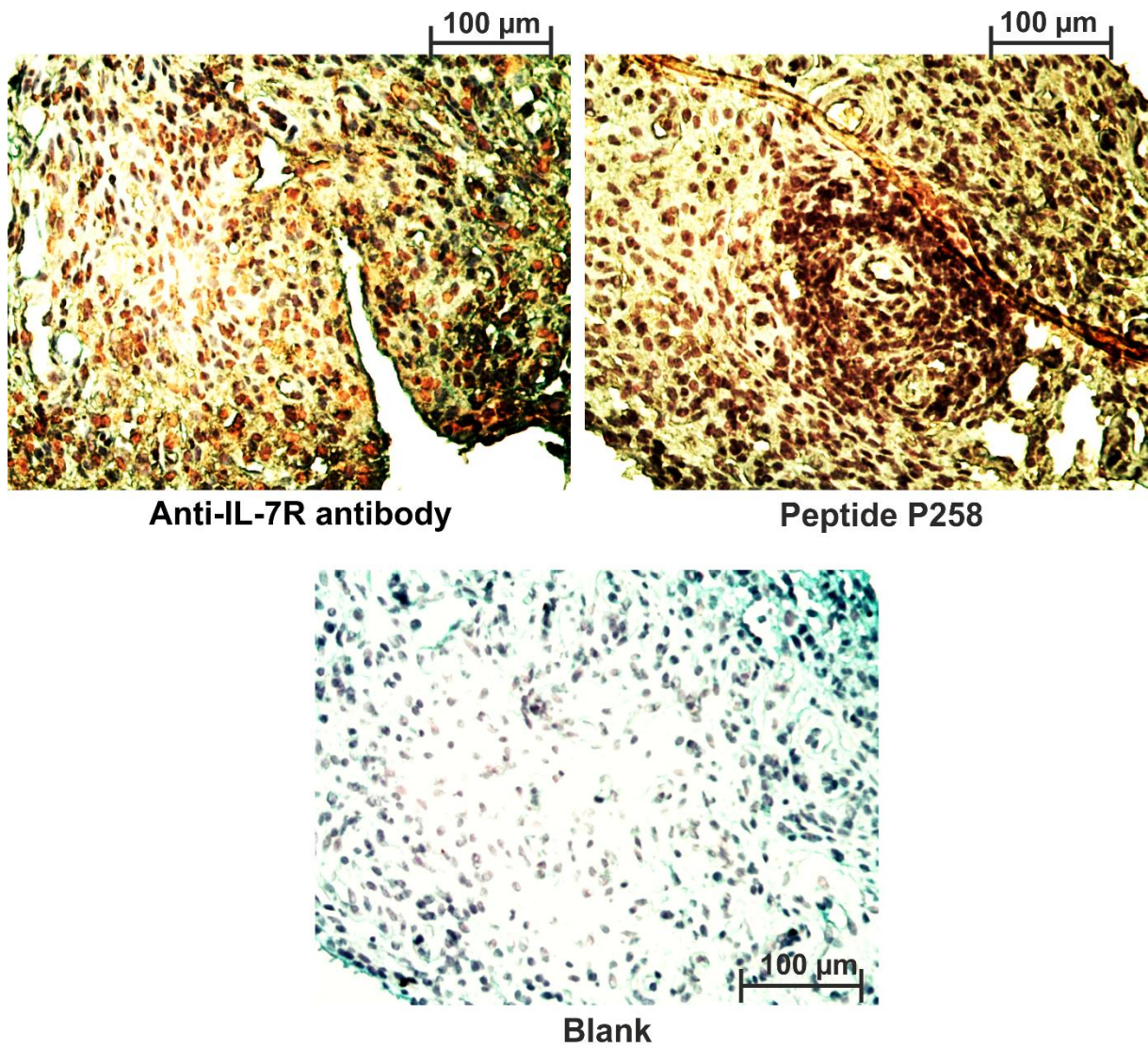
**Figure S2.** The affinity for IL-7R $\alpha$  of the clones 36/R2, 29/R3 and 37/R2 expressed by their own  $K_d^*$  and by the  $IC_{50}$  of IL-7 as determined in competition with each clone.



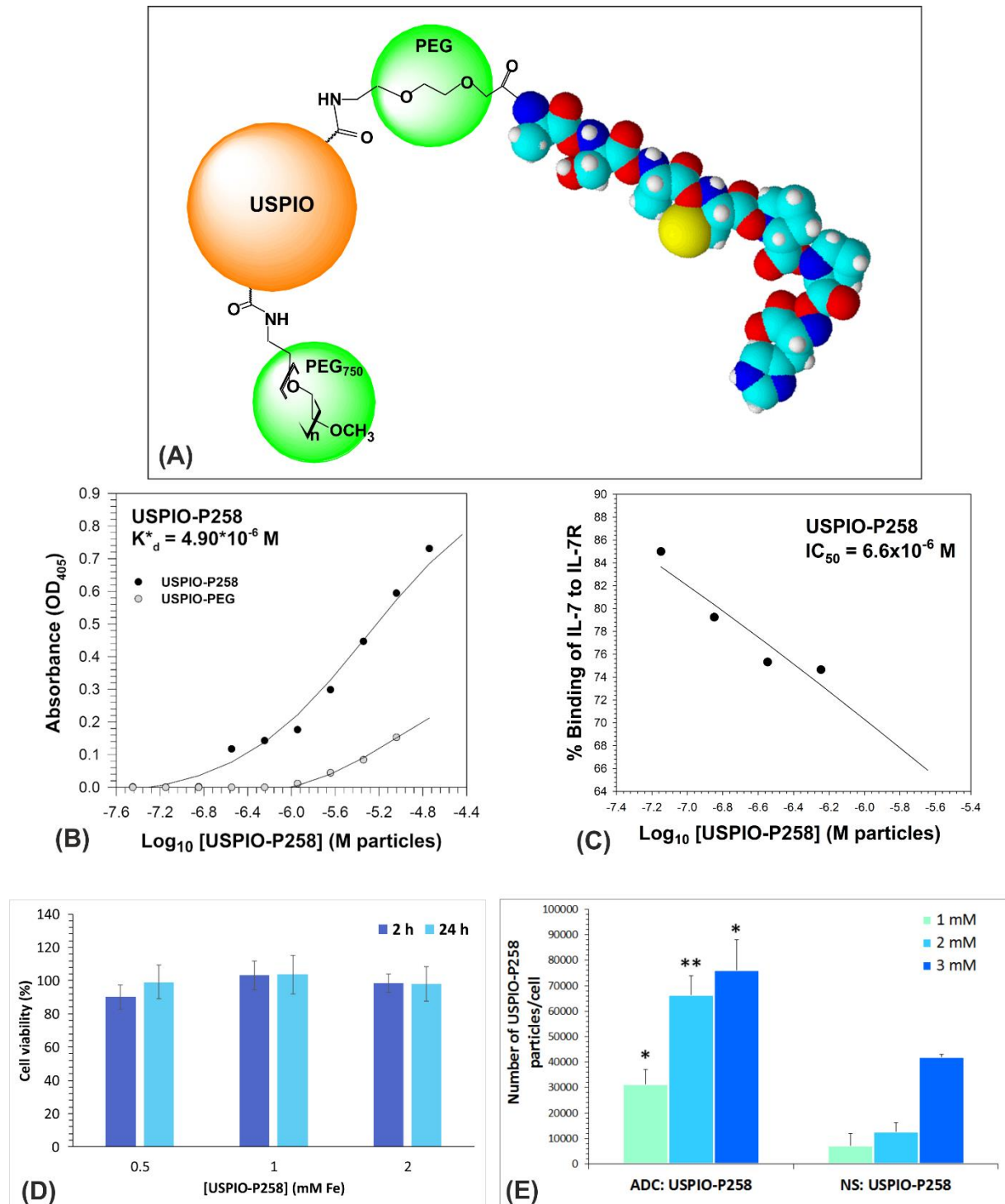
**Figure S3.** The affinity for IL-7R $\alpha$  and FN of the candidate peptides compared to that of IL-7. The  $IC_{50}$  of P725 as determined in competition with IL-7 is also shown.



**Figure S4.** 3D chemical structure of peptides P725 (A) and P258 (B) (drawn with ACD/ChemSketch 2.0 software) and their  $K_d$  for IL-7R $\alpha$  binding.

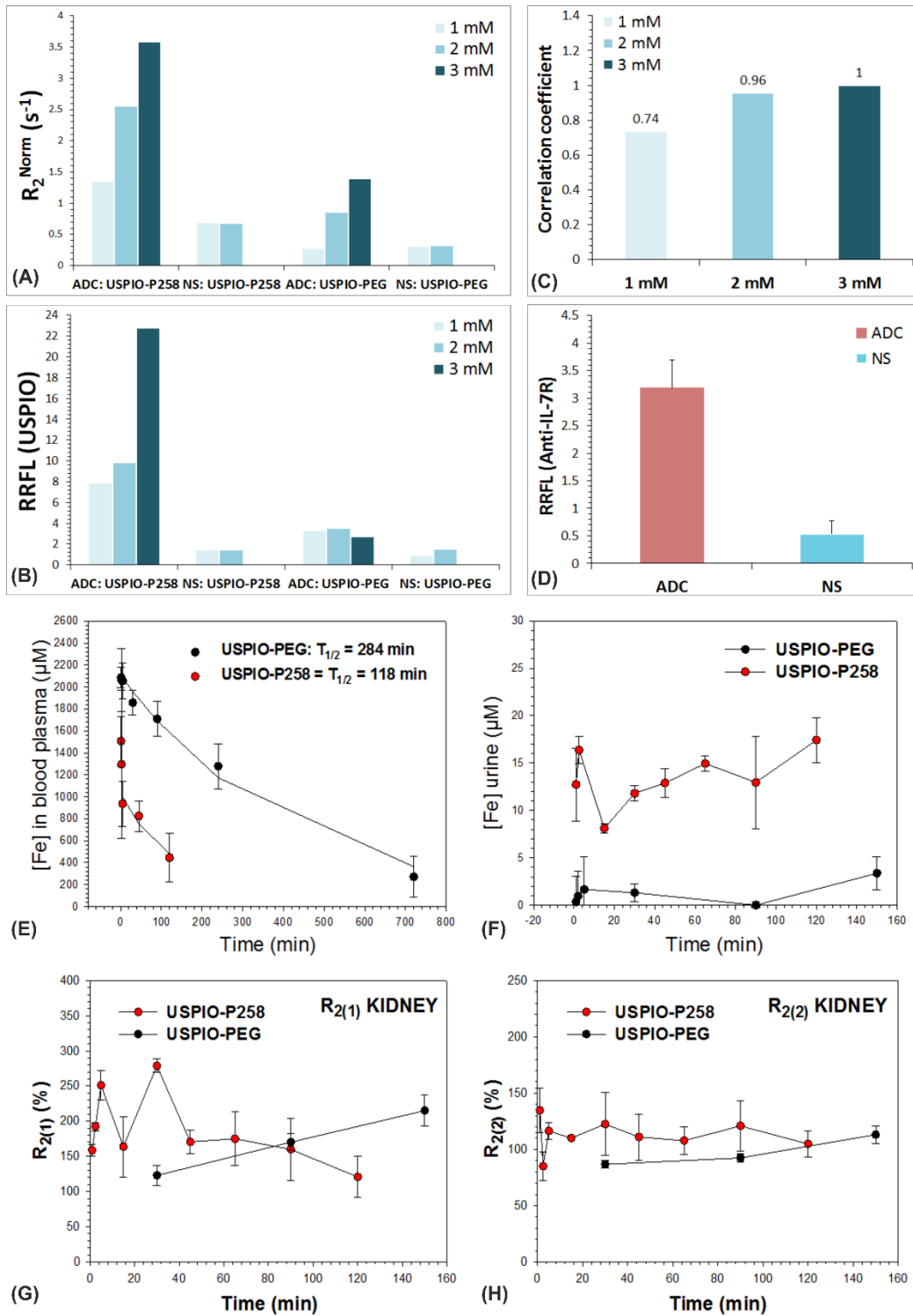


**Figure S5.** The expression of IL-7R $\alpha$  in RA synovial tissue and the binding of biotinylated peptide P258 to the same histological tissue; each microphotograph is representative of three stained sections.



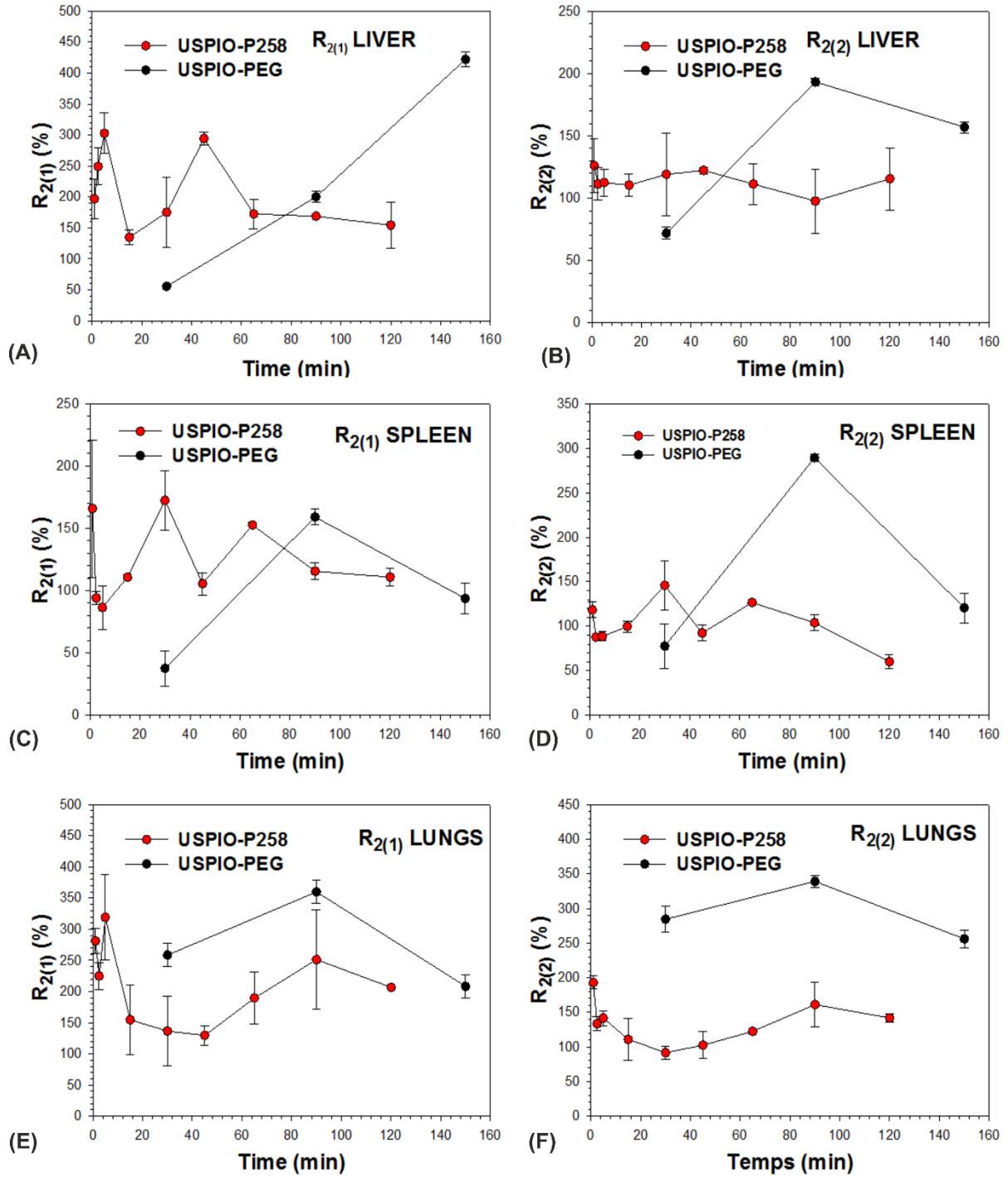
**Figure S6.** Affinity parameters, cytotoxicity and binding to Jurkat cells of USPIO-P258. Schematic representation of USPIO-P258 is shown in (A). The binding of USPIO-P258 to IL-7R $\alpha$  compared to that of USPIO-PEG (B), its IC<sub>50</sub> determined in competition with IL-7 (C) and its eventual cytotoxic effects evaluated on HepaRG<sup>TM</sup> cell line as determined by MTT assay (D). The number of USPIO-P258 particles bound per ADC or NS cell is shown in (E); \* =  $p < 0.05$ , \*\* =  $p < 0.01$  for ADC cells vs. NS cells.



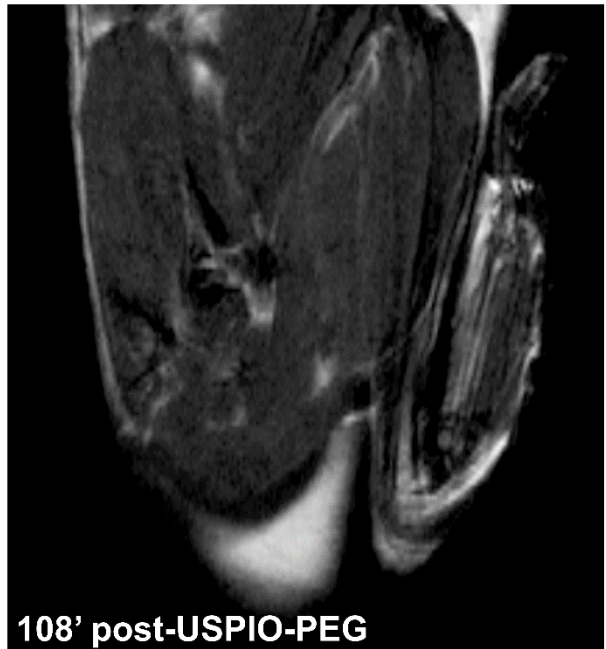
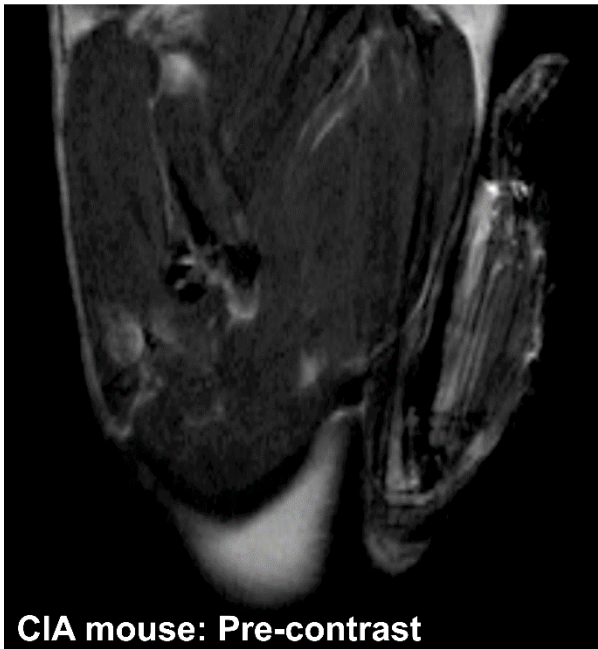
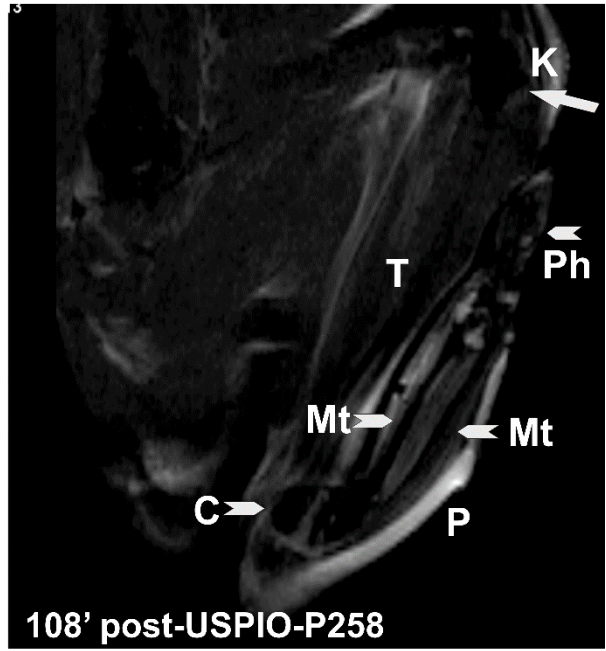
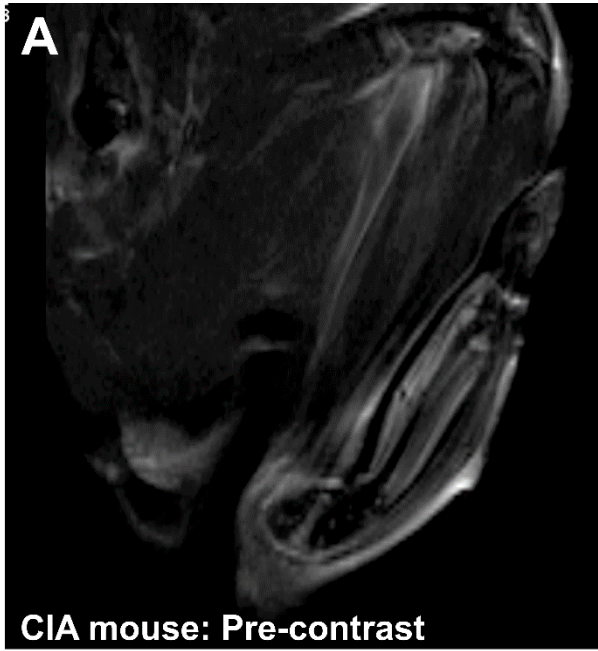


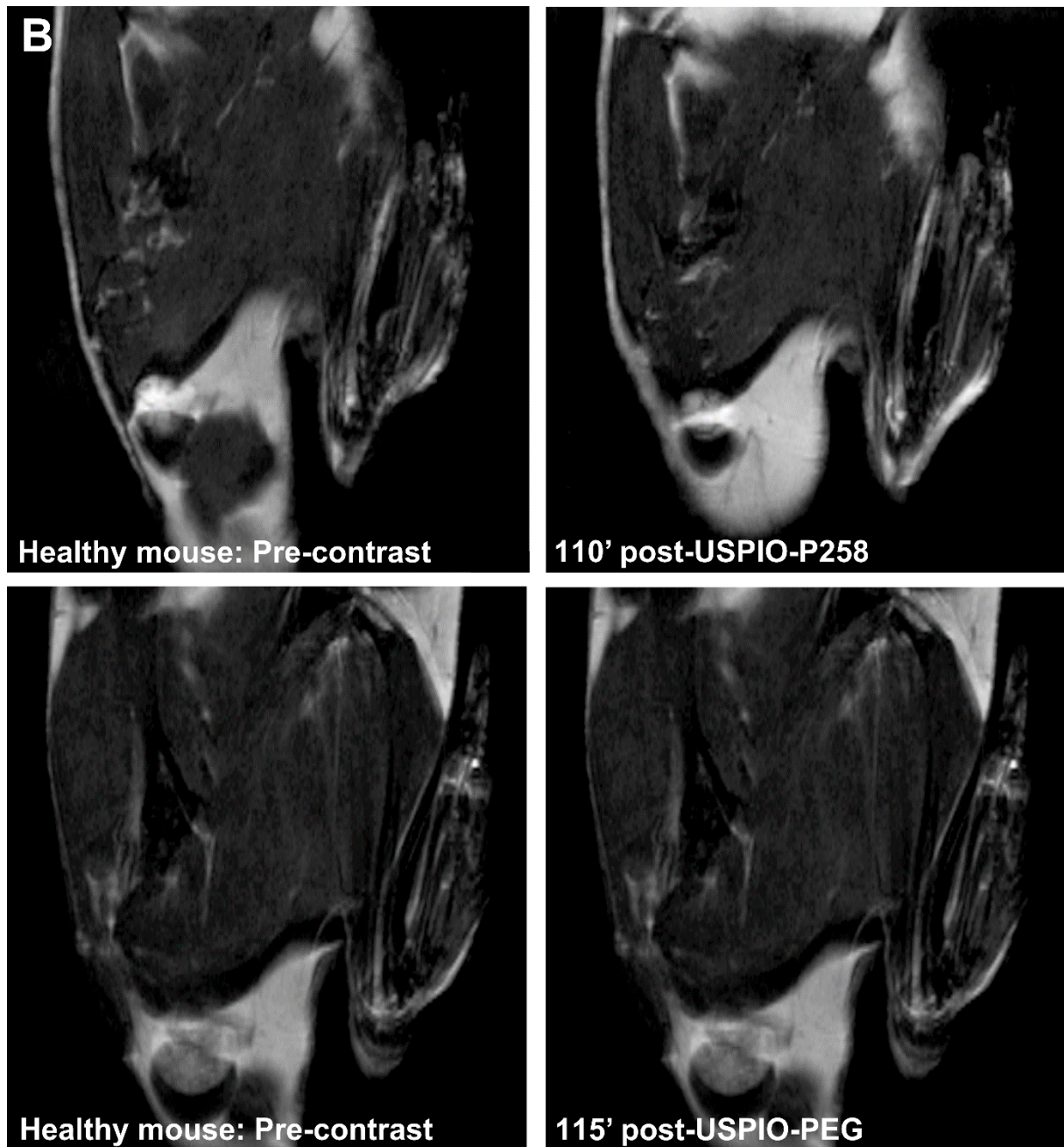
**Figure S7.**  $R_2^{\text{Norm}}$  (A) and RRFL (B) of ADC-stimulated (ADC) or non-stimulated (NS) Jurkat cells after incubation with USPIO-P258 or USPIO-PEG. The correlation coefficient between  $R_2^{\text{Norm}}$  and RRFL (C) as well as the IL-7R expression are also shown. The graphs presented in E-F illustrate the pharmacokinetics (E) and biodistribution of USPIO-P258 in urine (F) and kidney (G for  $T_{2(1)}$  and H for  $T_{2(2)}$ ) as compared to USPIO-PEG. The results for USPIO-PEG are cited from Ansciaux E et al. *Contrast Media Mol Imaging*. 2015; 10(3): 211-24.





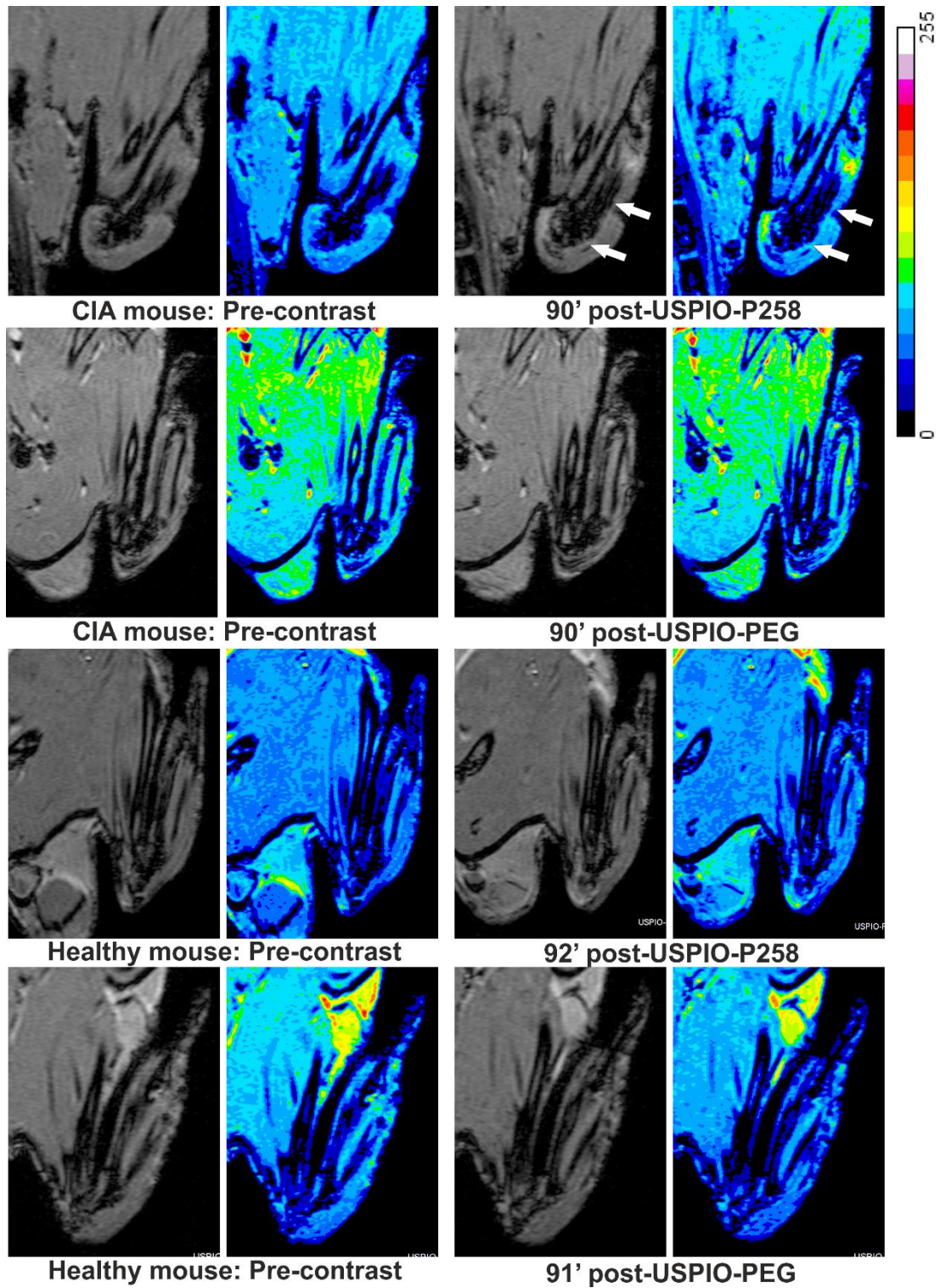
**Figure S8.** The biodistribution of USPIO-P258 in liver (A for  $T_{21}$  and B  $T_{22}$ ), spleen (C and D) and lungs (E for  $T_{2(1)}$  and F for  $T_{2(2)}$ ) as compared to USPIO-PEG. The results for USPIO-PEG are cited from Ansciaux E et al. Contrast Media Mol Imaging. 2015; 10(3): 211-24.



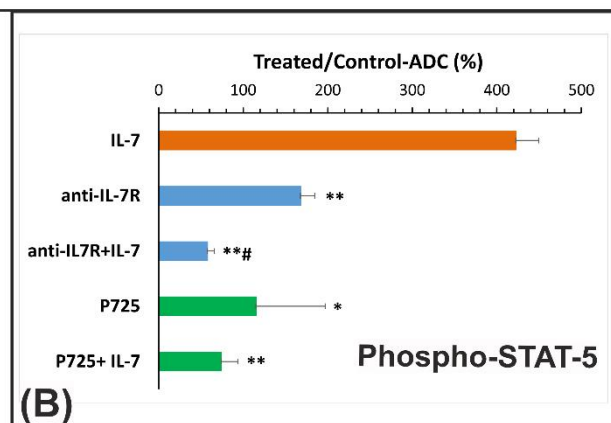
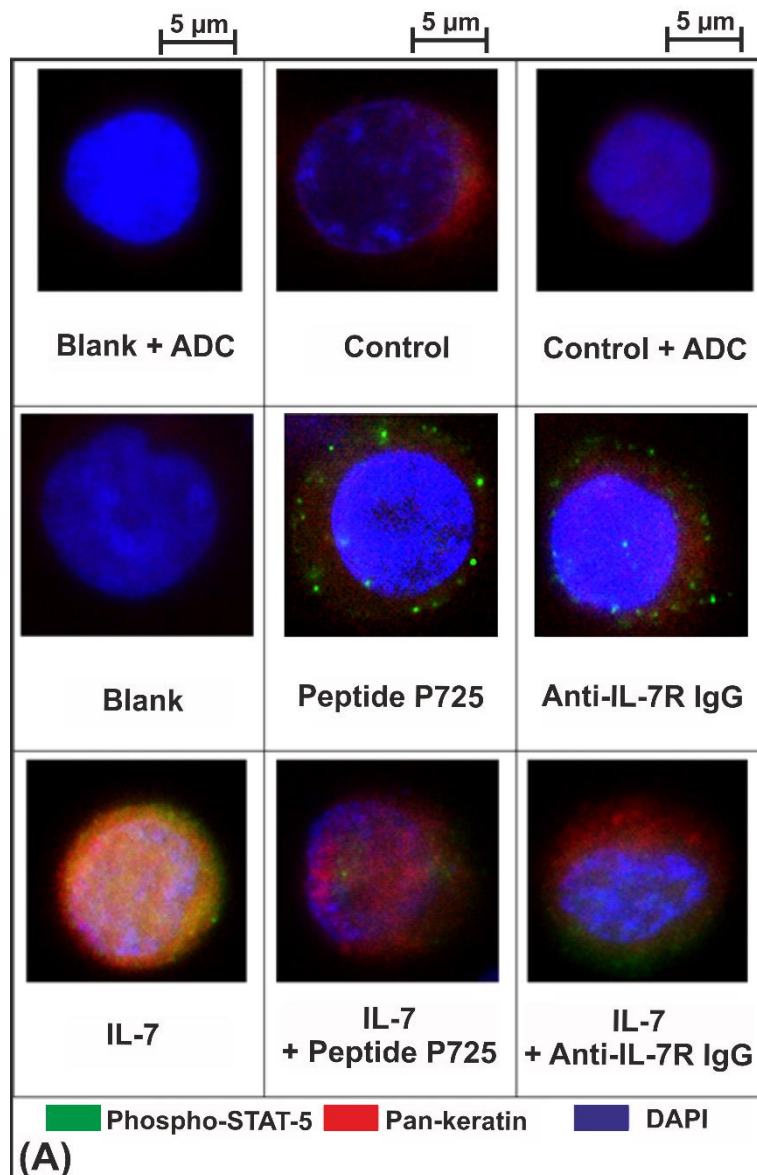


**Figure S9.** RARE MR images of the hind limbs of CIA (A) or healthy (B) mice injected with either USPIO-P258 or USPIO-PEG in pre-contrast and about 2 hours post-contrast. The arrowheads point to the hind limb; the arrow points to the knee. C = calcaneus; CB = coxal bones; K = knee; P = paw; Ph = phalanx; Mt = metatarsal bones; T = tibia.

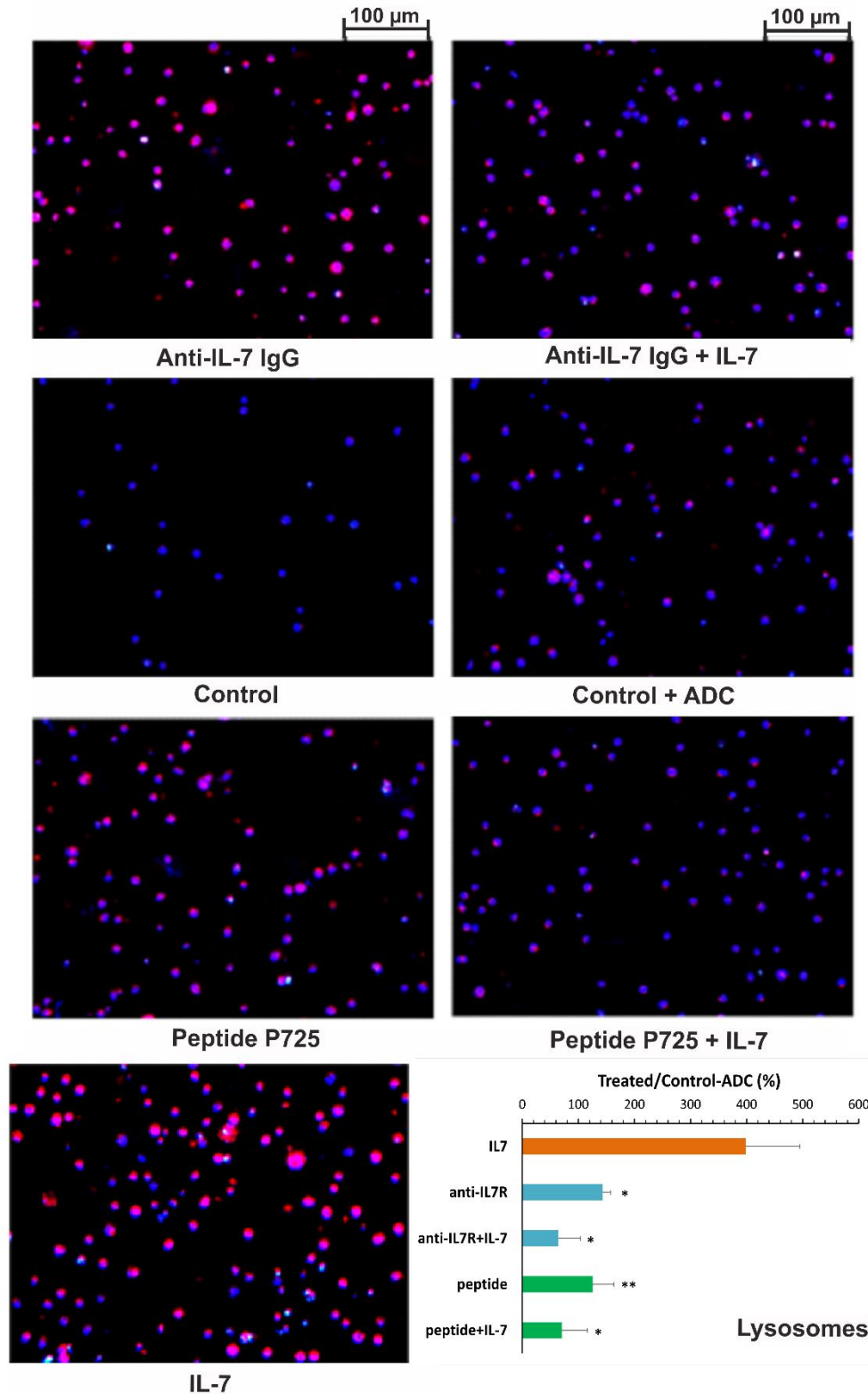




**Figure S10.** 3D FISP MR images of the hind limbs of CIA or healthy mice injected with either USPIO-P258 or USPIO-PEG in pre-contrast and at different times post-contrast. The color-coded signal enhancement is shown at the right side of each raw image. The arrows point to the hind limb.



**Figure S11.** Immunofluorescent labeling of phospho-STAT-5 in ADC-stimulated Jurkat cells is shown in (A); the blank and controls are ADC-stimulated or non-stimulated Jurkat cells and incubated with (controls) or no (blank) anti-phospho-STAT-5 antibody. The semi-quantitative analysis of phospho-STAT-5 is shown in (B). \* =  $p < 0.05$ , \*\* =  $p < 0.01$  vs. IL-7; # =  $p < 0.05$  vs. anti-IL-7R.



**Figure S12.** Lysosome detection by fluorescence on ADC-stimulated Jurkat cells and incubated with IL-7, with the competitor (anti-IL-7 antibody or peptide P725) or concomitantly with the competitor and IL-7. Controls are Jurkat cells stimulated or not with ADC. Lysosomes are stained in red with Lysotracker® Red DND-99, whereas nuclei are stained in blue with Hoechst 33342. The semi-quantitative analysis of microphotographs is shown in the histogram. \* =  $p < 0.05$ , \*\* =  $p < 0.01$  vs. IL-7.