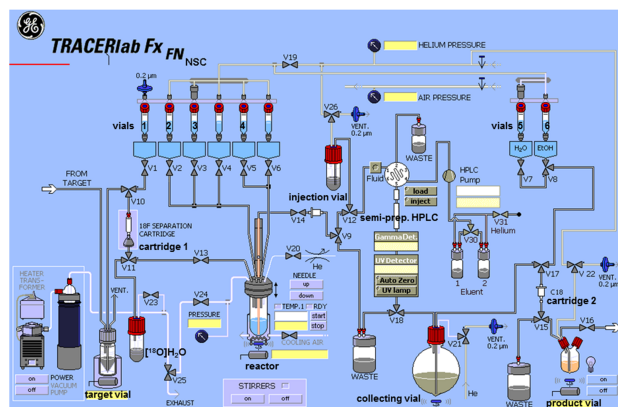


**Supplemental material to the paper by Röttering et al. entitled:  
Imaging of  $\alpha 7$  nicotinic acetylcholine receptors in brain and cerebral vasculature of juvenile pigs  
with [ $^{18}\text{F}$ ]NS14490**

## Supplement to Material and Methods

### Automated synthesis of [ $^{18}\text{F}$ ]NS14490

Automated radiosynthesis of [ $^{18}\text{F}$ ]NS14490 has been developed as follows. Figure 1 presents the set-up of vials and containers of the automatic module, and Table 1 shows which vials were used and how they were filled, and which cartridges were used.



**Fig. 1** Configuration of the TRACERlab<sup>TM</sup> FX F-N automate for the synthesis of [ $^{18}\text{F}$ ]NS14490

An activity of 6.5 - 15.8 GBq n.c.a. [ $^{18}\text{F}$ ]fluoride in aqueous solution placed in the target vial (2 mL, Figure 1), was trapped by passage through an anion exchange cartridge (cartridge 1, Figure 1) on the TRACERlab<sup>TM</sup> FX-N synthesis module. Activity was released into the reactor upon addition of  $\text{K}_2\text{CO}_3$  (1.8 mg, 13 mmol) diluted in 400  $\mu\text{L}$  acetonitrile/water 1:1 v/v (vial 1, Figure 1). Next, Kryptofix K<sub>222</sub> (11.2 mg, 28 mmol) in 1 mL anhydrous acetonitrile (vial 2, Figure 1), was added, and the  $\text{K}^{[18\text{F}]}\text{F-K}_{222}$  complex was formed under a flow of helium at 55°C for 6 min. After evaporation at 85°C under vacuum for 2 min, the reactor was cooled to 60°C and the tosylate precursor **1** dissolved in 750 - 800  $\mu\text{L}$  anhydrous acetonitrile (vial 3, Figure 1) was added to the reactor. Nucleophilic substitution was performed by heating to 85°C for 20 min under a helium atmosphere. The reactor was then cooled to 35°C and a sample was taken for radio-TLC. The process was continued by the addition of 3 mL of acetonitrile/water 35%/65% (vial 4, Figure 1) into the reactor and transferring the reaction mixture to the injection vial. After completion of the transfer, the loop was loaded and the mixture injected to the RP-column of the semi-preparative HPLC column. The fraction containing [ $^{18}\text{F}$ ]NS14490 was collected in a vial (Suppl. Figure 1) containing 10 mL water, and the solution was passed through the HR-X cartridge (cartridge 2, Suppl. Figure 1), trapping the radiotracer. The cartridge was washed by 3 mL water (vial 5, Figure 1) and the radiotracer was released into the product vial by passing 1.5 mL acetonitrile/1.5% formic acid (vial 6, Figure 1) through the cartridge. This solution was manually

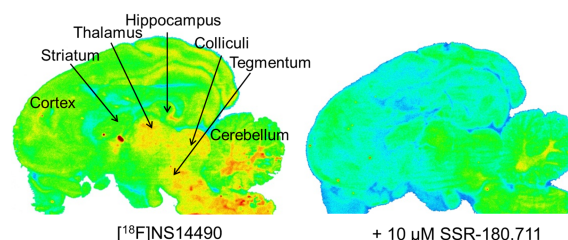
evaporated at 65°C - 70°C and formulated in 200  $\mu\text{L}$  - 400 mL phosphate-buffered saline (PBS) containing 5% ethanol.

### In vitro autoradiography of piglet brain

In vitro autoradiography with [ $^{18}\text{F}$ ]NS14490 on sagittal sections (20  $\mu\text{m}$ ) of the right brain hemispheres of 3 female piglets was performed based on previously published procedures [1] with slight modifications. The brain slices were incubated for 60 minutes at room temperature in assay buffer (50 mM TRIS-HCl, 120 mM NaCl + 5 mM KCl pH 7.4/21°C) containing 13.7 MBq [ $^{18}\text{F}$ ]NS14490 (0.228 MBq mL<sup>-1</sup>). Non-specific binding was determined on adjacent sections in the presence of 300  $\mu\text{M}$  (-)-Nicotine hydrogen tartrate salt. Specific binding was calculated as difference between total and non-specific binding. Competition of non-radioactive  $\alpha 7$ -selective ligands with [ $^{18}\text{F}$ ]NS14490 was investigated with 10  $\mu\text{M}$  of the partial agonist SSR-180,711 and 10  $\mu\text{M}$  of the antagonist methyllycaconitine citrate salt hydrate (MLA). After washing and air-drying all slides were exposed to BAS-SR2325 imaging plates (Fuji Film, Tokyo, Japan) for 24 h. Quantitative analysis of the scan data was performed by computer-assisted microdensitometry. Irregular regions of interest were drawn over selected areas of the brain. The brain regions of interest were confirmed by Nissl and Gallays staining [2] on adjacent sections with the help of a pig brain atlas [3].

## Supplemental information to Results

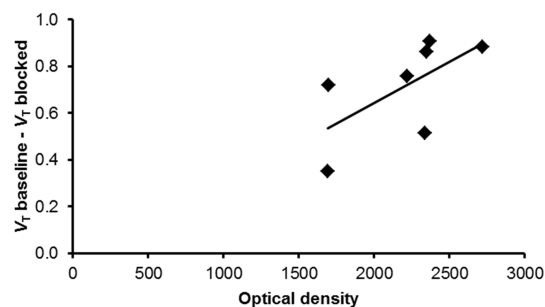
An in vitro autoradiograph of [ $^{18}\text{F}$ ]NS14490 binding on a sagittal sections of piglet brain is shown in Fig.2 (left). Co-incubation with 10  $\mu\text{M}$  of the partial agonist SSR-180,711 resulted in a strong reduction of [ $^{18}\text{F}$ ]NS14490 binding throughout the brain (Fig. 2, right). The highest inhibition was found in the hippocampus while the lowest was observed in the cerebellar white matter. Almost identical values were found when 10  $\mu\text{M}$  of the antagonist MLA was added to the incubation solution.



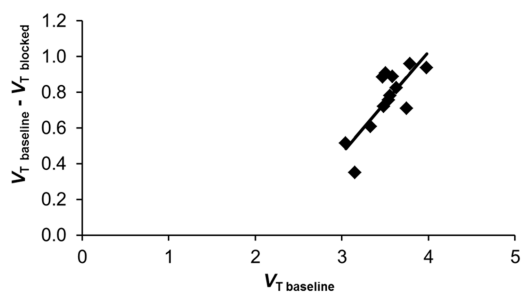
**Fig. 2** In vitro autoradiographs of [ $^{18}\text{F}$ ]NS14490 binding on sagittal sections of piglet brain, left: control, right: co-incubation with the  $\alpha 7$  nAChR- selective SSR-180,711.

The values of specific binding estimated in the brain regions indicated in Fig. 2 were correlated to the specific binding estimated by PET as difference of  $V_T$  under

baseline and blocked condition. A highly significant correlation ( $p < 0.002$ ) was found (Fig. 3).

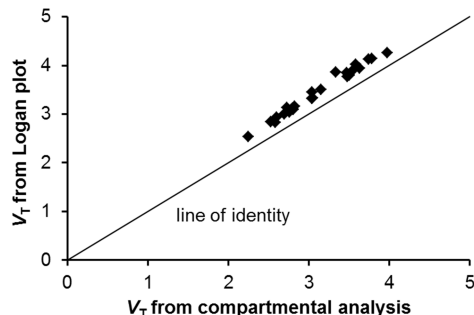


**Fig. 3** Correlation between the specific binding of  $[^{18}\text{F}]\text{NS14490}$  at porcine brain in vitro (optical density measured by autoradiography) and the specific binding measured by PET (difference of  $V_T$  under baseline and blocked conditions). Data are means of the values measured in various brain regions (cortex, striatum, thalamus, hippocampus, colliculi, midbrain tegmentum, cerebellum),  $r = 0.63$ ,  $p < 0.002$ .



**Fig. 4** Occupancy plot of  $[^{18}\text{F}]\text{NS14490}$   $V_T$ s in all investigated brain regions. Fraction of  $\alpha 7$  nAChR occupancy by the blocking agent NS6740 is calculated as slope of the regression line, and of  $[^{18}\text{F}]\text{NS14490}$   $V_{ND}$  as intercept at y-axis.  $r = 0.80$ ,  $p < 0.001$ .

Furthermore it has been shown that the PET estimate of specific  $[^{18}\text{F}]\text{NS14490}$  shows strong correlation to  $V_T$  under baseline conditions. The respective occupancy plot (Fig. 4) allows to estimate the average nonspecific binding,  $V_{ND}$ , of  $[^{18}\text{F}]\text{NS14490}$  in the piglet brain ( $2.2 \text{ mL cm}^{-3}$ ). Parametric images  $V_T$  obtained from Logan Plot represent specific binding of  $[^{18}\text{F}]\text{NS14490}$  in piglet brain because a strong Correlation between  $V_T$  from Logan Plot and  $V_T$  from compartmental analysis has been found (Figure 5).



**Fig. 5** Correlation between  $V_T$  from Logan Plot and  $V_T$  from compartmental analysis in all investigated brain regions together with the line of identity,  $r = 0.99$ .

## REFERENCES

1. Deuther-Conrad W, Wevers A, Becker G, Schildan A, Patt M, Sabri O, Steinbach J, Brust P. Autoradiography of 2- $[^{18}\text{F}]\text{F-A-85380}$  on nicotinic acetylcholine receptors in the porcine brain in vitro. *Synapse*. 2006;59:201-10.
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**Table 1** Experimental set-up for the production of  $[^{18}\text{F}]\text{NS14490}$  in an automated synthesis module. Steps marked with (\*) were optimised within the development of the radiosynthesis.

Element	filled/equipped with	volume	Remarks
target vial	$[^{18}\text{F}]\text{F}^-$ from cyclotron	2 mL, filled with $\text{H}_2\text{O}$	
$[^{18}\text{O}]\text{H}_2\text{O}$	empty		collection of $[^{18}\text{O}]\text{H}_2\text{O}$ for recycling
cartridge 1	$\text{PS-HCO}_3^-$ (unconditioned)		separation of activity from $[^{18}\text{O}]\text{H}_2\text{O}$
reactor	stirrer		
vial 1	1.8 mg $\text{K}_2\text{CO}_3$	400 $\mu\text{L}$ water/MeCN 1:1	desorption of the activity into the reactor
vial 2	11.2 mg Kryptofix $\text{K}_{222}$	1 mL MeCN	filled into reactor with activity
vial 3	2 mg tosylate precursor 1	800 $\mu\text{L}$ MeCN	
vial 4	MeCN/water 35:65	4 mL	for transfer to injection vial
injection vial	empty		for transfer to HPLC
semi-prep. HPLC	RP-column*		
collecting vial	water	10 mL	collection of fraction and dilution for SPE
cartridge 2	HR-X (M, 85 $\mu\text{m}$ ) (conditioned)		
vial 5	water	3 mL	washing of radiotracer trapped on cartridge
vial 6	desorption agent*	1.5- 3 mL	
product vial	empty		product after SPE for final evaporation and formulation