**Supplemental Information:** Monoamine oxidase binding would not be expected to significantly affect [<sup>18</sup>F]flortaucipir PET interpretation

Target Journal: European Journal of Nuclear Medicine and Molecular Imaging

Authors: Justin P. Wright<sup>1</sup>, Jason R. Goodman<sup>1</sup>, Yin-Guo Lin<sup>1</sup>, Brian P. Lieberman<sup>1</sup>, Jennifer Clemens<sup>1</sup>, Luis

F. Gomez<sup>1</sup>, Qianwa Liang<sup>1</sup>, Adam T. Hoye<sup>1</sup>, Michael J. Pontecorvo<sup>1</sup>, Kelly A. Conway<sup>1\*</sup>

\*Corresponding author

Affiliations: <sup>1</sup>Avid Radiopharmaceuticals, a wholly owned subsidiary of Eli Lilly & Company, Philadelphia,

PA, USA.

First author: Justin P Wright

Corresponding author: Kelly Conway

3711 Market Street

Seventh Fl Philadelphia, PA 19104, USA

conway@avidrp.com

#### **Supplemental Methods**

### Quadrupole Time-of-Flight Mass Spectrometer (QTOF-MS)

Samples (15 µL) were injected on a Waters Acquity UPLC system equipped with a Waters Acquity T3 HSS, 2.1 x 50mm, 1.8 µm column at a mobile phase flow rate of 0.5 mL/min, and (MS) response was monitored with a Waters Xevo QTOF scanning 50-1200 Da. For each sample analysis, the column was equilibrated with 95:5 water with 0.1% formic acid (Mobile Phase A): acetonitrile with 0.1% formic acid (Mobile Phase B) for 0.25 minutes. Following injection, the gradient was changed to 70:30 Mobile Phase B: Mobile Phase A in a linear fashion over 1.75 minutes, and then to 100% Mobile Phase B over 0.2 minutes, held at 100% Mobile Phase B for 0.8 minutes, followed by 95:5 Mobile Phase A:Mobile Phase B over 0.1 minutes and held for 0.4 minutes. Masslynx QuanLynx<sup>™</sup> software was used to analyze MS data. An internal standard was added to all samples to correct for instrument and sample processing variability.

# Supplemental Results

### [<sup>18</sup>F]Flortaucipir autoradiography in rat brain tissue using mild wash conditions

Experimental conditions as described in the Materials and Methods section of the manuscript "[<sup>18</sup>F]Flortaucipir Autoradiography: Comparison of Stringent and Mild Wash Conditions". Under mild wash conditions, high background ARG signal is seen in rat brain tissue samples (male wild-type Sprague Dawley). The MAO-B inhibitor deprenyl (1  $\mu$ M) has no blocking effect on [<sup>18</sup>F]flortaucipir ARG signal. The MAO-A/B inhibitor pargyline (10  $\mu$ M), MAO-A inhibitor, clorgyline (1  $\mu$ M), and flortaucipir (1  $\mu$ M) partially block [<sup>18</sup>F]flortaucipir ARG signal.



Figure S1. Autoradiography of [<sup>18</sup>F]flortaucipir in rat brain tissue with blocking by MAO-A/MAO-B inhibitors and flortaucipir.

# Possible artifactual saturation binding curve

Saturation binding for [<sup>18</sup>F]flortaucipir against recombinant monoamine oxidase-B (MAO-B) when using 1  $\mu$ M [<sup>18</sup>F]flortaucipir to define non-specific binding (A) generates an artifactual [<sup>18</sup>F]flortaucipir:MAO-B saturation binding curve with a K<sub>d</sub> of 28 nM by artificially reducing the [<sup>18</sup>F]flortaucipir nonspecific binding below that of the control microsomes. A similar artifactual binding curve with a K<sub>d</sub> of 16 nM is generated for [<sup>18</sup>F]flortaucipir curves run in binding buffer only, in which 10  $\mu$ M flortaucipir is used to define NSB (B). This signal represents background binding of [<sup>18</sup>F]flortaucipir to the filter, which can be reduced by adding saturating amounts of non-radioactive flortaucipir.





Abbreviations: CPM, counts per minutes; MAO, monoamine oxidase; NSB, Non-specific binding; TB, Total Binding.