### **Online resource 1 : Supplementary figures**

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### Distinct molecular profile of diffuse cerebellar gliomas

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Fig. S1 – S12

# DCGs with H3F3A K27M



DCG\_01





DCG\_11

# DCGs with SETD2 mutation



DCG\_02

DCG\_03



DCG\_06

# Other DCGs



Fig. S1 Magnetic resonance imaging (MRI) findings of representative DCGs. Gadolinium-enhanced T1-weighted axial MRI of three DCGs with H3F3A K27M, four DCGs with SETD2 mutation, and four other DCGs are shown.



Wild type



Fig. S2 Immunohistochemical staining of DCGs using anti-H3 K27M antibody. All three cases with nuclear positivity are shown on the left, and three of 24 cases without nuclear staining are shown on the right. Scale bar 50  $\mu$ m.



**Fig. S3** Mutation load of DCGs. Pie charts showing the number of different types of somatic mutations in 15 non-hypermutator samples and in two hypermutator samples (DCG\_04 and DCG\_17).



Fig. S4 WES results for DCGs. Genes mutated in more than two cases except for the two hypermutator cases are shown.WHO grade, hypermutator, and sample ID are indicated at the top. Types of alteration are indicated as colored boxes.Frequencies of alteration in each gene are shown on the right.



DCG\_01, RNA-seq *H3F3A*, Missense Chr.1: 226252135, A to T



DCG\_02, RNA-seq *NF1*, Frameshift del Chr.17: 29676180-81 del



DCG\_04, RNA-seq SETD2, Nonsense Chr.3: 47162252, G to A

	Т		
	T		
	T		
G	Т		
	T		
	Т		
	Т		
	_	_	_

A C T T C T T G C DCG\_04, RNA-seq *NF1*, Missense Chr.17: 29588778, C to T

				Α				
				A				
				A				
				Α				
				Α				
				Α				
				A				
				A				
_				Α				
				A				
Δ.	G	G	Δ	G	G	G	G	C

DCG\_05, RNA-seq *TP53*, Missense Chr.17: 7578280, G to A



DCG\_01, RNA-seq *TP53*, Missense Chr.17: 7577548, C to T



DCG\_03, Sanger-seq SETD2, Splicing Chr.3: 47098981, C to G



DCG\_04, Sanger-seq *TP53*, Splice site Chr.17: 7579591, C to G



DCG\_04, RNA-seq *MSH3*, Missense Chr.5: 79974908, G to C



DCG\_05, RNA-seq BRAF, Missense Chr.7: 140501299, C to A



DCG\_02, RNA-seq SETD2, Nonsense Chr.3: 47165534, G to A



DCG\_03, RNA-seq PPM1D, Frameshift del Chr.17: 58740467-89 del



DCG\_04, RNA-seq *PDGFRA*, Missense Chr.4: 55141062, C to T



T T A G C T T G T DCG\_04, RNA-seq *PMS2*, Missense Chr.7: 6029468, C to G



DCG\_06, RNA-seq SETD2, Nonsense Chr.3: 47142990, G to C



DCG\_02, RNA-seq *TP53*, Missense Chr.17: 7577115, A to C



DCG\_03, RNA-seq *PDGFRA*, Missense Chr.4: 55136819, C to G



DCG\_04, Sanger-seq *EGFR*, Splice site Chr.7: 55228019, G to A



DCG\_05, RNA-seq *H3F3A*, Missense Chr.1: 226252135, A to T

С	Α	G	Α	С	G	G	Α	Α
				Α				
				Α				
				A				
				A				
				A				
				A				
				A				
				A				
				A				
				4				

DCG\_06, RNA-seq *TP53*, Missense Chr.17: 7579358, C to A



DCG\_07, RNA-seq *TP53*, Missense Chr.17: 7578208, T to C



DCG\_11, RNA-seq FGFR1, Missense Chr.8: 38272308, T to C



DCG\_15, Sanger-seq FGFR1, Missense Chr.8: 38272308, T to C



DCG\_17, RNA-seq *MLH1*, Frameshift ins Chr.3: 37070349, ins



DCG\_08, RNA-seq *PPM1D*, Nonsense Chr.17: 58740749, C to T



DCG\_12, RNA-seq ATRX, Frameshift del Chr.X: 76907752 del



DCG\_15, Sanger-seq *PIK3R1*, Missense Chr.5: 67591097, A to G



DCG\_17, RNA-seq *MSH6*, Frameshift del Chr.2: 48027637 del



DCG\_10, RNA-seq *TP53*, Missense Chr.17: 7577153, C to A



DCG\_11, RNA-seq *H3F3A*, Missense Chr.1: 226252135, A to T

DCG\_14, Sanger-seq BRAF, Missense Chr.7: 140453136, A to T



DCG\_17, RNA-seq *TP53*, Nonsense Chr.17: 7578263, G to A



DCG\_17, RNA-seq *PMS1*, Nonsense Chr.2: 190718671, C to T



DCG\_15, Sanger-seq *TP53*, Missense Chr.17: 7578448, G to A

Т
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Т
T

G G C A C G G T G DCG\_17, RNA-seq *EGFR*, Missense Chr.7: 55241726, C to T

Fig. S5 Validation of mutations identified by WES. All mutations in Fig.1a were confirmed by corresponding RNA-sequencing reads or Sanger sequencing. Sample number, methods used for validation, and mutation information are shown below each graph. *RNA-seq* RNA-sequencing, *Sanger-seq* Sanger sequencing, *ins* insertion, *del* deletion.



**Fig. S6** Chromosome copy number alteration profiles of each sample. Chromosome copy number status is color-coded for each chromosomal site band for each patient. *Chr* chromosome, *cnLOH* copy-neutral LOH.

SETD2 mutant (DCG\_04)

SETD2 wild type (DCG\_05)



SETD2 mutant (DCG\_06)

SETD2 wild type (DCG\_07)



**Fig. S7** Representative immunohistochemistry of H3K36 trimethylation in DCGs. Representative immunohistochemical staining of H3K36 trimethylation (H3K36me3) and hematoxylin and eosin (HE) staining of *SETD2* mutant (DCG\_04 and DCG\_06) and wild type (DCG\_05 and DCG\_07) DCGs. Nuclei of vascular endothelial cells are indicated by the white arrows as the internal positive control for H3K36me3 staining. Scale bar, 50 μm.



**Fig. S8** Detection of *PPM1D*-noncoding (antisense *RPS6KB1* isoform 2 and 3) fusion and validation PCR. **a** Split-reads are shown aligning on *PPM1D* exon 5 and the intragenic region in *RPS6KB1*. Reads on the intragenic *RPS6KB1* region were transcribed reciprocally and included a few isoforms. *Chr* chromosome. **b** Validation PCR of the *PPM1D*-noncoding (antisense RPS6KB1 isoform 2 and 3) fusion. PCR primers were designed to specifically amplify fusion products (top). PCR bands of the estimated size of each isoform were detected in DCG\_12 (middle). In a negative control lane (N), PCR product amplified without template DNA was electrophoresed. Predicted sequences were confirmed by Sanger sequencing (bottom).



Fig. S9 Consensus *k*-means clustering of 224 gliomas. Consensus cumulative distribution function (CDF) and consecutive differences of areas under the CDF curves from *k*-means clustering for different numbers of methylation clusters are indicated. In our study, k = 6 was also selected as previously reported [48].









**Fig. S10** Gene expression analysis of DCGs and cerebral GBMs. GSEA showed that two gene sets were up-regulated in 14 DCGs in both the "RTK I" group and the "K27" group compared with eight cerebral GBMs. One gene set was overexpressed in "PDGFRA-amplified GBMs" (left), and the other gene set was overexpressed in "Proneural GBMs" (right). The false discovery rate (q) and normalized enrichment score (NES) are shown on top. The top 30 significantly up-regulated genes of each gene set in DCGs are shown at the bottom. Cases with *H3F3A* K27M are indicated with green colored boxes.



**Fig. S11** Methylation level of *SOX10*, *FOXG1*, *OLIG1*, and *OLIG2* loci in 224 samples including 18 DCGs. The map of the chromosomes (Chr) of these four genes, the position of CpG islands, and the positions of the Infinium probes are shown at the top. Each row represents a sample, and each vertical bar represents an Infinium probe. Methylation clusters in this study and brain regions of each sample are shown on the left.



Rank	Motif	<i>p</i> -value	% of	% of
			Targets	Backgrounds
1	<b><u><u><u></u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u>C</u><u></u></u></b>	1x10 <sup>-26</sup>	54.9%	38.7%
2	CCATTRACAG	1x10 <sup>-21</sup>	57.2%	42.3%
3	<b>GGGITTCATTT</b> <u>G</u>	1x10 <sup>-19</sup>	0.95%	0.01%
4	ATCGATCG	1x10 <sup>-19</sup>	42.9%	29.7%
5	<b>LATTCACG</b>	1x10 <sup>-18</sup>	65.5%	51.9%

**Fig. S12** Motifs enriched in the hypomethylated DNA region of the "*SOX10* promoter hypomethylation" group. **a** Glioma samples (n = 224) were classified into three groups depending on the *SOX10* promoter methylation level. **b** A volcano plot comparing the beta-value of each probe more than 1,500 bp from the TSS between 18 DCGs and 123 cerebral high-grade gliomas is shown. One dot represents one probe. The *q*-values that were calculated using a paired two-sided moderated Welch's *t*-test were plotted on the y-axis. Methylation differences expressed as beta-values are plotted on the x-axis. Significantly hypomethylated probes (1,070; *q*-value <  $1 \times 10^{-10}$  and methylation difference > 0.25) in the "*SOX10* promoter hypomethylation" group were selected. **c** Top five enriched motifs in sequences around the 1,070 hypomethylated probes of the "*SOX10* promoter hypomethylation" group are shown.