

# Calpain system protein expression and activity in ovarian cancer

Journal of Cancer Research and Clinical Oncology

Siwei Zhang<sup>1</sup>, Suha Deen<sup>2</sup>, Sarah J Storr<sup>1</sup>, Panagiota S Chondrou<sup>1,3</sup>, Holly Nicholls<sup>1</sup>, Anqi Yao<sup>1</sup>, Ployphailin Rungsakaolert<sup>1</sup>, Stewart G Martin<sup>1,4</sup>

1 Academic Clinical Oncology, Nottingham Breast Cancer Research Centre, School of Medicine, University of Nottingham, Nottingham University Hospitals NHS Trust, City Hospital Campus, Nottingham, NG5 1PB, UK.

2 Department of Histopathology, Queen's Medical Centre, Nottingham University Hospitals NHS Trust, Nottingham, NG7 2UH, UK

3 Current address: Lungs for Living Research Centre, UCL Respiratory, University College London, 5 University Street London, WC1E 6JF United Kingdom.

4 Corresponding author: Academic Clinical Oncology, Nottingham Breast Cancer Research Centre, School of Medicine, University of Nottingham, Nottingham University Hospitals NHS Trust, City Hospital Campus, Nottingham, NG5 1PB, UK

Telephone: +44(0)115 823 1846

Email: [stewart.martin@nottingham.ac.uk](mailto:stewart.martin@nottingham.ac.uk).

## Supplements

**Table S1 Treatment received by the ovarian cancer patients in present study**

<b>Treatment</b>	<b>Number of Patients</b>	<b>Frequency (%)*</b>
<b>Carboplatin</b>	148	25.7
<b>Carboplatin and paclitaxel (taxol ®)</b>	160	27.8
<b>Carboplatin and docetaxel (Taxotere ®)</b>	2	0.3
<b>Carboplatin, docetaxel and irinotecan (Campto ®)</b>	1	0.2
<b>Carboplatin, paclitaxel and topotecan (Hycamtin ®)</b>	3	0.5
<b>Carboplatin, paclitaxel and bevacizumab</b>	1	0.2
<b>Carboplatin and cyclophosphamide</b>	5	0.9
<b>Carboplatin, cyclophosphamide and doxorubicin (adriamycin)</b>	21	3.7
<b>Carboplatin and topotecan</b>	9	1.6
<b>Carboplatin and radiotherapy (HGSC)</b>	1	0.2
<b>Carboplatin, paclitaxel and radiotherapy (HGSC)</b>	1	0.2
<b>CAP: cyclophosphamide, doxorubicin and cisplatin</b>	5	0.9
<b>Chlorambucil (Leukeran ®)</b>	2	0.3
<b>Topotecan (Hycamtin ®)</b>	2	0.3
<b>Gemcitabine (Gemzar ®)</b>	1	0.2
<b>No chemotherapy</b>	74	12.9
<b>Radiotherapy (HGSC and endometrioid carcinoma)</b>	2	0.3
<b>Radiotherapy and progesterone (endometrioid carcinoma)</b>	1	0.2
<b>Dexamethasone and radiotherapy for lung metastasis (endometrioid carcinoma)</b>	1	0.2
<b>BIBF 1120 (Nintedanib)</b>	2	0.3

\* Percentage of the total cohort (n=575).

**Table S2 Spearman's rank correlation coefficient between calpain-1, -2, -4 and calpastatin expression**

		<b>Calpain-1</b>	<b>Calpain-2</b>	<b>Calpain-4</b>
<b>Calpastatin</b>	<b>Correlation Coefficient</b>	<b>0.480**</b>	<b>0.205**</b>	<b>0.514**</b>
	<b>Sig. (2-tailed)</b>	2.4147E-28	0.000008	6.783E-33
<b>Calpain-1</b>	<b>Correlation Coefficient</b>	\	<b>.300**</b>	<b>0.481**</b>
	<b>Sig. (2-tailed)</b>	\	3.3557E-11	1.5002E-28
<b>Calpain-2</b>	<b>Correlation Coefficient</b>	\	\	<b>0.296**</b>
	<b>Sig. (2-tailed)</b>	\	\	6.4717E-11

\*\* Correlation is significant at the 0.01 level (2-tailed).

\* Correlation is significant at the 0.05 level (2-tailed).

Significant P-values are indicated by bold type.

**Table S3 Kaplan–Meier survival analysis in different subgroups of ovarian cancer**

	<i>P</i> value (confidence interval: 99%)			
	Calpastatin	Calpain-1	Calpain-2	Calpain-4
<b>Histological subtypes</b>				
<b>HGSC (n=265)</b>	0.002	0.658	0.011	0.064
<b>Mucinous carcinoma (n=45)</b>	0.497	0.448	0.556	0.014
<b>Endometrioid carcinoma (n=53)</b>	0.415	0.617	0.713	0.899
<b>CCC (n=41)</b>	0.564	0.308	0.826	0.461
<b>LGSC (n=24)</b>	0.930	0.866	0.257	0.987
<b>Stage</b>				
<b>Stage 1 (confined tumour) (n=163)</b>	0.019	0.997	0.634	0.011
<b>Stage 2, 3 &amp; 4 (tumour spread) (n=286)</b>	0.008	0.766	0.025	0.044
<b>Stage 1, 2 &amp; 3 (without distant metastasis) (n=416)</b>	0.019	0.195	0.078	0.004
<b>Stage 4 (with distant metastasis) (n=33)</b>	0.976	0.254	0.024	0.963
<b>Sensitivity to platinum-based chemotherapy</b>				
<b>Sensitive (n=213)</b>	0.050	0.049	0.435	0.036
<b>Resistant (n=39)</b>	0.798	0.815	0.076	0.433
<b>Residual disease</b>				
<b>No residual tumour (n=250)</b>	0.085	0.988	0.244	0.026
<b>residual tumour (n=153)</b>	0.002	0.361	0.594	0.012

**Table S4 Profiler PCR array results (PEO4 cells vs. PEO1 cells).**

Refseq	Gene	Description	Fold-change			
			4h		24h	
PPH00527A	BMP7	Bone morphogenetic protein 7	259.84	A	351.79	A
PPH00439F	COL3A1	Collagen, type III, alpha 1 chain precursor	75.56	A	107.55	A
PPH00152E	MMP9	Matrix metalloproteinase 9	4.38	A	2.88	B
PPH20529A	NUDT13	Nudix (nucleoside diphosphate linked moiety X)-type motif 13	4.17	A	3.53	B
PPH01944B	NODAL	Nodal homolog (mouse)	11.16	B	16.07	B
PPH09021B	ZEB2	Zinc finger E-box binding homeobox 2	2.11	B	11.02	B
PPH00781A	COL5A2	Collagen, type V, alpha 2	4.47	B	7.66	B
PPH02389A	KRT14	Keratin 14	5.22	B	5.40	B
PPH00215F	SERPINE1	Serpin peptidase inhibitor, clade E, member 1	4.68	B	\	\
PPH00235F	MMP3	Matrix metalloproteinase 3	4.24	B	\	\
PPH02459B	SNAI1	Snail homolog 1 (Drosophila)	3.22	B	\	\
PPH02399C	WNT11	Wingless-type MMTV integration site family, member 11	3.15	B	2.57	B
PPH00531F	TGFB3	Transforming growth factor, beta 3	2.72	B	\	\
PPH00526C	NOTCH1	Notch 1	2.48	B	\	\
PPH11409A	TMEM132A	Transmembrane protein 132A	6.29	A	-2.27	A
PPH02410A	WNT5A	Wingless-type MMTV integration site family, member 5A	-13.74	A	-10.28	A
PPH06022B	JAG1	Jagged 1	-11.73	A	-7.15	A
PPH00524B	TGFB2	Transforming growth factor, beta 2	-2.05	\	-9.22	A
PPH02475A	SNAI2	Snail homolog 2 (Drosophila)	-2.72	\	-6.98	A
PPH02447C	WNT5B	Wingless-type MMTV integration site family, member 5B	-5.32	\	-5.47	A
PPH00176C	ITGA5	Integrin, $\alpha$ 5 (fibronectin receptor)	-3.90	A	-2.25	A
PPH00582E	SPP1	Secreted phosphoprotein 1	\	\	-3.23	A
PPH00515B	BMP1	Bone morphogenetic protein 1	-2.86	A	-2.41	A
PPH00555G	IL1RN	Interleukin 1 receptor antagonist	-2.70	A	-2.44	A
PPH00636F	CDH2	N-cadherin	-2.21	A	-17.91	B
PPH00151B	MMP2	Matrix metalloproteinase 2	-13.11	B	-8.84	B
PPH01971A	FOXC2	Forkhead box C2	-5.70	B	\	\
PPH01922A	ZEB1	Zinc finger E-box binding homeobox 1	\	\	-3.85	B
PPH15155C	SNAI3	Snail homolog 3 (Drosophila)	-2.02	B	\	\

A: This gene's expression is relatively low (threshold cycle > 30) in one sample and reasonably detected (threshold cycle < 30) in the other sample suggesting that the actual fold-change value is at least as large as the calculated and reported fold-change result.

B: This gene's relative expression level is low (threshold cycle > 30), in both control and test samples.

**Table S5 Profiler PCR array results for PEO1 and PEO4 cells (calpeptin-treated group vs. vehicle control group at two time points).**

Gene	Description	Fold-change	
		4h	24h
<b>Up-regulated gene expression in PEO1</b>		<b>4h</b>	<b>24h</b>
<b>ZEB2</b>	Zinc finger E-box binding homeobox 2	3.21 (B)	4.32 (B)
<b>BMP2</b>	Bone morphogenetic protein 2	\	2.71 (B)
<b>BMP7</b>	Bone morphogenetic protein 7	2.04 (B)	\
<b>Up-regulated genes in PEO4</b>		<b>4h</b>	<b>24h</b>
<b>CDH2</b>	Cadherin 2, type 1, N-cadherin (neuronal)	\	8.76 (B)
<b>KRT14</b>	Keratin 14	2.72(B)	2.18 (B)
<b>MMP2</b>	Matrix metalloproteinase 2 (gelatinase A, 72kDa gelatinase, 72kDa type IV collagenase)	2.40(B)	\
<b>SNAI3</b>	Snail homolog 3 (Drosophila)	2.39(B)	\
<b>ZEB2</b>	Zinc finger E-box binding homeobox 2	2.37(B)	\
<b>Down-regulated gene expression in PEO1</b>		<b>4h</b>	<b>24h</b>
<b>MMP9</b>	Matrix metalloproteinase 9 (gelatinase B, 92kDa gelatinase, 92kDa type IV collagenase)	-3.06 (B)	\
<b>NOTCH1</b>	Notch 1	-2.20 (B)	\
<b>Down-regulated gene expression in PEO4</b>		<b>4h</b>	<b>24h</b>
<b>NODAL</b>	Nodal homolog (mouse)	-2.99 (B)	-9.83 (B)
<b>WNT11</b>	Wingless-type MMTV integration site family, member 11	-2.03 (B)	-2.06 (B)
<b>ZEB1</b>	Zinc finger E-box binding homeobox 1	-2.40 (B)	
<b>ZEB2</b>	Zinc finger E-box binding homeobox 2		-4.71 (B)